

COMPARATIVE STUDY OF PHENOLIC ACIDS
FROM UNDERGROUND PARTS
OF *RHEUM PALMATUM* L., *R. RHAPONTICUM* L.
AND *R. UNDULATUM* L.

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ABSTRACT

In three species of *Rheum* L. genus growing in Poland the composition of phenolic acids was determined. By 2D-TLC method the following acids were identified: ellagic, chlorogenic, gallic, protocatechuic, homoprotocatechuic, caffeic, α -resorcilic, p-hydroxyphenylacetic, p-hydroxybenzoic, p-coumaric, syringic, vanillic and ferulic. There are no substantial qualitative differences among the complex of phenolic acids in the investigated species. The RP-HPLC method was used for quantitative determination of phenolic acids. The amount of individual phenolic acids ranged between 2.2 $\mu\text{g/g}$ and 147.8 $\mu\text{g/g}$ in air-dry rhizome. The content of ferulic acid is the highest in all the examined cases. The total amount of the tested phenolic acids in *Rheum undulatum* L., *R. palmatum* L., and *R. raphaniticum* L. was respectively 346.4 $\mu\text{g/g}$, 229.8 $\mu\text{g/g}$, and 195 $\mu\text{g/g}$.

KEY WORDS: Polygonaceae, *Rheum*, rhizome and roots, phenolic acids, 2D-TLC, RP-HPLC.

INTRODUCTION

Rheum genus belonging to the family Polygonaceae is represented by about 70 species. Only three out of the 70 species are growing in Poland. They are: *R. palmatum*, *R. raphaniticum* (*R. rhabarbarum*), *R. undulatum*.

R. palmatum is cultivated for therapeutic properties of roots. The petioles of *Rheum raphaniticum* and *R. undulatum* are used for their nutritious properties.

The main components of *Rheum* are: anthraquinones (emodin, aloe-emodin, chrysophanol, physcion, rhein and its derivatives (Chai et al. 1998; Liu 1999; Li et al. 2000); diantrones (Oshio et al. 1974); stilbenes (resveratrol, piceatannol, raphanigenin) and its derivatives (Matsuda et al. 2001; Kashiwada et al. 1986; Kageura et al. 2001); naphthalenes (Tsuboi et al. 1977); chromones, phenylbutanones (Kashiwada et al. 1986).

Among them, there are compounds possessing chemotaxonomic attributes. The presence of raphanitin in *R. rhabarbarum* is the marker differentiating *R. raphaniticum* from *R. palmatum*. Most phenolic compounds may be used as chemotaxonomic markers.

The aim of this paper is the comparative study of phenolic acids obtained from underground parts of three *Rheum* species cultivated in Poland. Phenolic acids of the mentioned species were investigated in *Rheum palmatum* and

R. undulatum. P-coumaric acid and gallic acid and its derivatives were detected (Kashiwada et al. 1986; Abe et al. 2000), and gallic acid was isolated from *R. undulatum* (Kageura et al. 2001; Matsuda et al. 2001).

MATERIAL AND METHODS

Plant material

The rhizome and roots of three specimens of *Rheum* were used in this study: *Rheum palmatum* was from Herbapol (Lublin, Poland); *Rheum undulatum* and *Rheum raphaniticum* L. were collected in the Botanical Garden of the University Maria Curie-Skłodowska in Lublin in 2003.

Rheum raphaniticum and *Rheum undulatum* are deposited in the Department of Pharmaceutical Botany, Medical Academy in Lublin.

Extraction and isolation

The air-dried plant material (50 g) were extracted three times (15 min. for each extraction) with 80% methanol in ultrasonic bath. The obtained extract was purified and free phenolic acids fractions were isolated in classical way (Ibrahim and Towers 1960). The processes of acid hydrolysis of phenolic acids conjugated with sugar were performed according to Schmidlein and Herrmann (1975). The

TABLE 1. The occurrence of phenolic acids in rhizome and roots of *Rheum palmatum* L., *Rheum undulatum* L. and *Rheum rhaponticum* L.

No.	Acids	Commonly used names	<i>R. palmatum</i>			<i>R. undulatum</i>			<i>R. rhaponticum</i>		
			A	B	C	A ₁	B ₁	C ₁	A ₂	B ₂	C ₂
1	galloyl gallic	ellagic	+	+	-	+	+	-	+	+	-
2	caffeoyl quinic	chlorogenic	+	-	-	+	-	-	+	-	-
3	3,4,5-trihydroxybenzoic	gallic	-	+	+	+	+	+	+	+	+
4	3,4-dihydroxybenzoic	protocatechuic	+	+	+	+	+	+	+	+	+
5	3,4-dihydroxyphenylacetic	homoprotocatechuic	-	+	+	-	+	+	-	+	+
6	3,4-dihydroxycinnamic	caffeic	-	+	+	-	+	+	-	+	+
7	3,5-dihydroxybenzoic	α -resorcylic	+	-	-	+	-	-	+	-	-
8	4-hydroxyphenylacetic	p-hydroxyphenyl-acetic	+	+	-	+	+	+	+	+	-
9	4-hydroxybenzoic	p-hydroxybenzoic	+	+	+	+	+	+	+	+	+
10	4-hydroxycinnamic	p-coumaric	+	+	+	+	+	+	+	+	+
11	3,4,5-trimethoxybenzoic	syringic	+	+	+	+	+	+	+	+	+
12	3-methoxy-4-hydroxybenzoic	vanillic	+	+	+	+	+	+	+	+	+
13	3-methoxy-4-hydroxycinnamic	ferulic	+	+	+	+	+	+	+	+	+

A – free phenolic acids fraction; B – after acid hydrolysis; C – after alkaline hydrolysis; (+) – present; (-) – absent

alkaline hydrolysis were performed with Ba (OH)₂ in reductive medium NaBH₄ (Świątek and Dombrowicz 1987).

The samples of free phenolic acids (A), phenolic acids liberated by acid hydrolysis (B), and phenolic acids liberated by alkaline hydrolysis (C) were dissolved in 10 mL of ethanol and investigated by 2D-TLC and HPLC methods.

2D-TLC qualitative determination of phenolic acids

Separation and identification of phenolic acids (samples A-C) was performed on 10×10 cm cellulose plates (Merck, Darmstadt, Germany). After spotting of the phenolic acids standard solution or plant extract the plates were developed in DS horizontal chambers (Dzido 1993) in phases according to Smolarz and Waksmundzka-Hajnos (1993). After drying, the chromatograms were observed in UV light ($\lambda=254$ nm) before and after treatment with ammonia vapour. Derivatisation was performed by spraying with one of the two reagents: diasotized sulphanic acid in 20% sodium carbonate solution or 2% aqueous solution of ferric chloride.

HPLC quantitative determination of phenolic acids

The quantitative determination of nine main phenolic acids was carried out using Knauer (Germany) Model nr K-1001 liquid chromatograph with 20 μ L sample injector (Rheodyne, Cotati, CA, USA) and UV detector (UV-VIS) operated at 254 nm.

The stainless-steel column 200×4.6 mm J.D. was filled with 5 nm Hypersil ODS (Shandon, Cheshire, UK). The isocratic mobile phase consisted of methanol-water (25:75) with 5% v/v acetic acid. The flow rate was 1 mL/min. Chromatography was performed at room temperature. The chromatograms were recorded with Chroma 2000 program.

RESULTS

From the qualitative investigations carried out by two-dimensional thin-layer chromatography method it follows that rhizome and roots of the investigated raw material contain derivatives of benzoic, cinnamic and phenylacetic acids. They are the following acids: ellagic, chlorogenic, gallic, protocatechuic, homoprotocatechuic, caffeic, α -resorcylic, p-hydroxyphenylacetic, p-hydroxybenzoic, p-cou-

maric, syringic, vanillic and ferulic. The mentioned acids are commonly present both in elementary and linked state except α -resorcylic acid and chlorogenic acid, which are present only in free state, and caffeic and homoprotocatechuic which are detected only in bounded and liberated form. The distribution of these compounds is shown in Table 1.

The data of chromatographical (2D-TLC) analysis show no substantial qualitative differences among groups of phenolic acids from the investigated species.

RP-HPLC method was used for quantitative determinations of phenolic acids. The amount of each phenolic acid was interpreted from calibration curve obtained for concentrations of standard solution of acid containing 80, 100 and 120% of the expected level of this compound in the extracts. It was found that the calibration plots for all acids were of the type $y=bx+a$, where y is the peak area, x is the concentration of the standard, a is the intercept of the plot on y axis, and b is its slope. The errors in slope and intercept of the regression line, standard deviation and detection limit are shown in Table 2.

The amount of individual phenolic acids in *R. palmatum* (in A+B+C fractions) ranged between 21.2 μ g/g of air dry rhizome for syringic acid and 67.7 μ g/g for ferulic acid (Fig. 1). The minor component of *R. undulatum* is p-hydroxybenzoic acid (7.2 μ g/g) and major component is ferulic acid (147.8 μ g/g) (Fig. 2). The rhizome and roots of *R. rhaponticum* are also rich in ferulic acid (77.7 μ g/g) and poor in p-hydroxybenzoic (2.2 μ g/g) (Fig. 3). In all cases the content of ferulic acid is the highest. Besides, the ferulic acid, concentration of p-coumaric acid and protocatechuic acid is significant in all investigated species. Homoprotocatechuic and vanillic acids occurred in the highest amounts in *R. palmatum*.

We show that phenolic acids concentration is higher in linked than free state. *R. rhaponticum* contains 35.1 μ g/g of free phenolic acids, *R. palmatum* – 45.9 μ g/g and *R. undulatum* – 48.8 μ g/g; higher content of acids linked with sugar (69.8 μ g/g 90.8 μ g/g and 90.9 μ g/g respectively) and most acids as ester derivatives (*R. palmatum* – 105 μ g/g; *R. rhaponticum* – 111.5 μ g/g; *R. undulatum* (185.6 μ g/g).

On the basis of the obtained results one can expect that *R. palmatum* and *R. undulatum* contain similar amounts of phenolic acids in free state and bound with sugars, whereas

TABLE 2. Data of quantitative determination of phenolic acids.

Phenolic acid	Detection limit (L.o.d) (ng)	Standard deviation	Regression line	Errors in slope and intercept
gallic	13	0.9928	$y = 6133 x - 12.6$	$b = 6133 \pm 16.5$ $a = -12.6 \pm 2.4$
protocatechuic	10	0.9454	$y = 8944 x + 39.7$	$b = 8944 \pm 30.5$ $a = 39.7 \pm 25.2$
caffeic	12	0.9992	$y = 2973 x + 12$	$b = 2973 \pm 12.1$ $a = 12 \pm 1.8$
p-hydroxybenzoic	16	0.9998	$y = 13.8 x + 14.8$	$b = 13.8 \pm 0.4$ $a = 14.8 \pm 23.8$
p-coumaric	60	0.9988	$y = 3.6 x + 555$	$b = 3.6 \pm 1.15$ $a = 555 \pm 56.6$
syringic	11	1.0130	$y = 342 x + 7.7$	$b = 342 \pm 5.9$ $a = 7.7 \pm 0.87$
vanillic	10	0.9940	$y = 11044 x + 37$	$b = 11044 \pm 19.3$ $a = 37 \pm 18.6$
ferulic	64	1.0004	$y = 3.9 x + 8.8$	$b = 3.9 \pm 0.92$ $a = 8.8 \pm 46.1$

R. rhaponticum contains more of these compounds after alkaline hydrolysis.

Large total amounts of the nine investigated acids was detected in underground parts of *R. undulatum* (346.4 µg/g), whereas in *R. palmatum* – 229.80 µg/g, and *R. rhaponticum* L. – 195 µg/g.

The main active components of genus *Rheum* L. are anthraquinones, but the presence of other phenolic compounds such as stilbenes and phenolic acids occurring in these spe-

cies in lower concentration have a synergic influence on the activity of these species.

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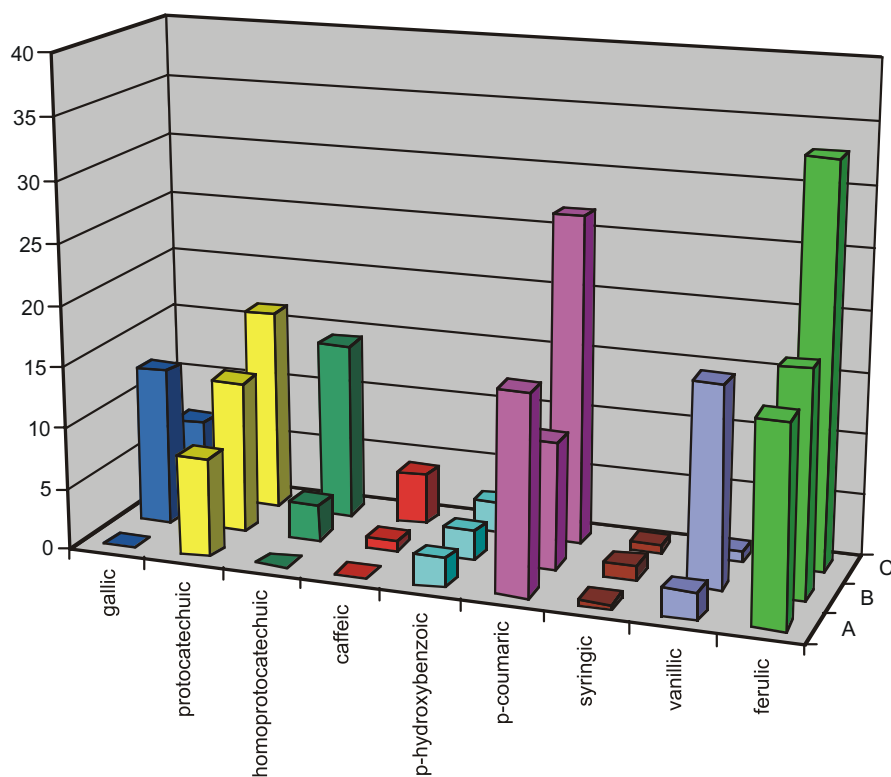


Fig. 1. Content of phenolic acids in *R. palmatum* L. (in µg/g air-dried underground parts).

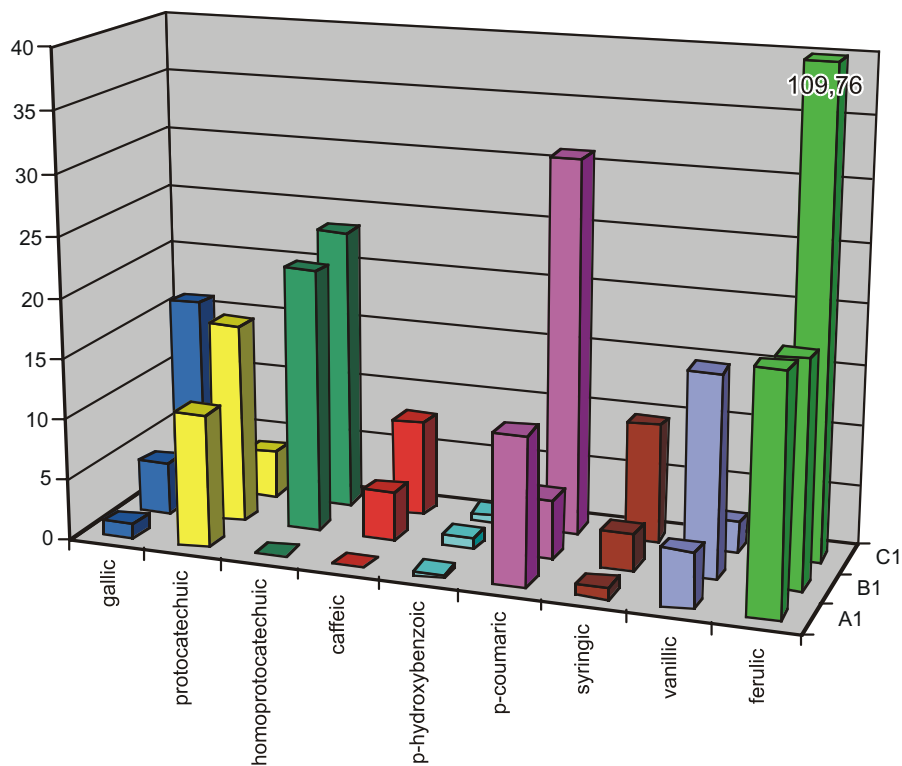


Fig. 2. Content of phenolic acids in *R. undulatum* L. (in µg/g air-dried underground parts).

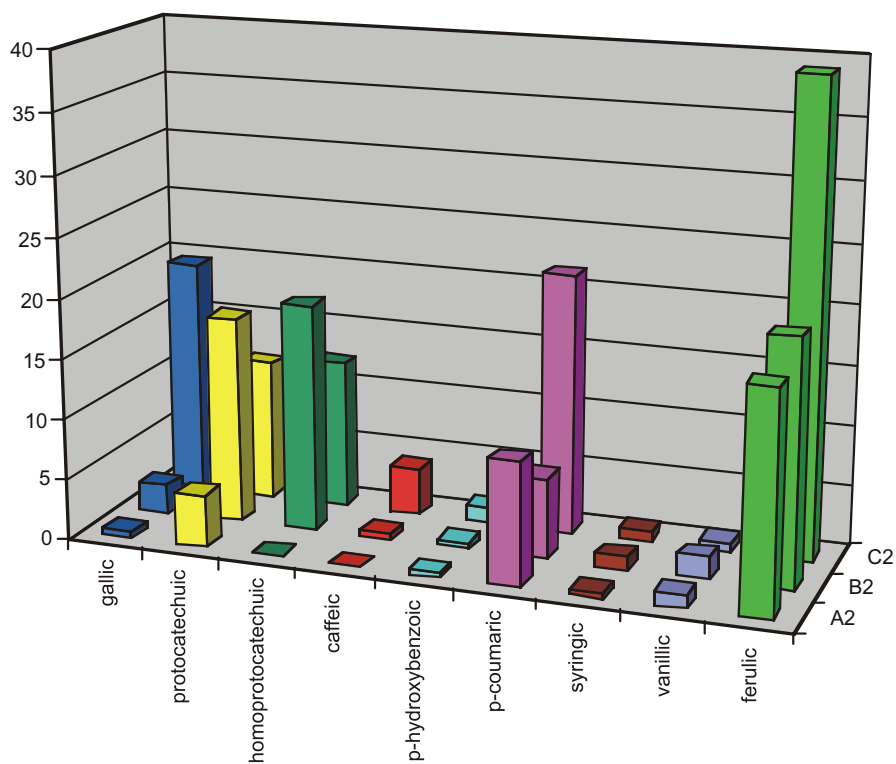


Fig. 3. Content of phenolic acids in *R. rhaponticum* L. (in µg/g air-dried underground parts).

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