

## Distribution of lichens and lichenicolous fungi in the Świętokrzyski National Park

ANNA ŁUBEK<sup>1</sup> and STANISŁAW CIEŚLIŃSKI<sup>2</sup>

<sup>1</sup>Department of Algology and Mycology, University of Łódź, Banacha 12/16,  
PL-90-237 Łódź, adonica@biol.uni.lodz.pl

<sup>2</sup>Institute of Biology, Świętokrzyska Academy, Świętokrzyska 15  
PL-25-406 Kielce, sciesl@pu.kielce.pl

Lubek A., Cieśliński S.: *Distribution of lichens and lichenicolous fungi in the Świętokrzyski National Park*. Acta Mycol. 39 (2): 173-252, 2004.

This paper presents the distribution of 405 taxa of the lichens and 26 species of lichenicolous fungi in the Świętokrzyski National Park.

**Key words:** lichenized and lichenicolous fungi, distribution, Świętokrzyski National Park, Świętokrzyskie Mts.

### INTRODUCTION

The Świętokrzyski National Park (SNP) covers the central area of the Świętokrzyskie Mountains – the oldest and the northernmost and easternmost mountain range in Poland, uplifted during the Caledonian and Variscan orogenetic uplift. These mountains stand out because of a characteristic geological structure consisting in, among others, uncovering of layers of rocks from 500 million years ago that form boulder fields (Wróblewski 2000) (Fig. 1). At the same time, the SNP is one of the oldest national parks in Poland – founded in 1950. The rich and diversified lichen biota of the Świętokrzyskie Mountains has long been of interest to lichenologists. Lichenological research in this area dates back to the 19<sup>th</sup> century and has continued with varied intensity until the present day. As regards the SNP, there have been four periods describing the degree of research of the lichen biota: up to 1960, 1961-1980, 1981-1985, 1999-2002. The first period is characterized by a substantial concentration of research in the area of the highest hills of the Łysogóry Range (Berdau 1876; Błoński 1890; Kobendza and Motyka 1928, 1929; Kobendza 1939). Rich herbarium collections of B. Halicz and S. Kuziel (from 1955-1959) and M. Sroczyński (from 1960) dating from that period, deposited in the Herbarium of Lichens and Lichenicolous Fungi at the University of Łódź (LOD) were published after 1960 (Halicz and Kuziel 1965, 1966). The findings concerning lichens found in the second period of research come from the

selected areas of the Łysogóry and Klonowskie Ranges, the Wilkowska Valley, as well as from the protection zone and may often be considered as complementary (Tobolewski 1962; Halicz and Cieśliński 1967; Bystrek and Cieśliński 1976; Cieśliński and Halicz 1971; Toborowicz 1977; Cieśliński 1975, 1981). Some publications from that period were based on previous herbarium collections (Cieśliński and Bystrek 1982). The third period includes the spatially vastest research. It concerned the whole area of the ŚNP in its borders from before 1996, it did not, however, took the area of the protection zone into consideration (Cieśliński and Toborowicz 1989; Cieśliński 1985, 1991, 2000). The latest research conducted between 1999-2002 covered the whole area of the ŚNP and its protection zone (Łubek 2004).

The lichen biota of the area inside the borders of the national park should be considered as well studied within the Świętokrzyskie Mts. This article presents a distribution of all species of lichens and lichenicolous fungi found in the area of the ŚNP and its protection zone.

#### MATERIAL AND METHODS

The research material – lichens and lichenicolous fungi – comes from three sources: published and unpublished literature, herbarium collections deposited in the Herbarium of Lichens and Lichenicolous Fungi at the University of Łódź (LOD) and the Herbarium of Lichens at the Świętokrzyska Academy (KTC), as well as the field studies conducted between 1999-2002 (Łubek 2004).

The research included all habitats, both natural and antrophogenic, within the area of the ŚNP and its protection zone where lichens may be found. Epiphytic lichens (growing on the bark of living trees, bushes and shrubs), epixylic lichens (found on wood in various stages of decay), epilithic lichens (saxicolous) growing on limestone rock and quartritic sandstone, epigaeic lichens (terrestrial) and epibryophytic lichens (growing on bryophytes) have been studied.

The grid square method has been applied. The research area was divided into 88 squares-sites, each side of the square 2 kilometers long. The square grid has been adjusted to the national ATLICHEN grid – acc. to Cieśliński and Fałtynowicz (1993). The studied squares are included in eight ATLICHEN squares: Ee65, Ee66, Ee67, Ee75, Ee76, Ee77, Ee78, Ee87. The spatial arrangement of the squares and their numbering are presented in Fig. 2. Field charting of the lichen and lichenicolous fungi biota was made between 1999-2002. Published, unpublished, and herbarium historical materials, except for those where the exact location of the site was impossible, were charted, too.

The spatial distribution of lichens and lichenicolous fungi is presented in cartograms. Fig. 1 serves to locate the square in which a given taxon is found. A single cartogram (Fig. 2) – a grid of squares, each side of the square 2 kilometers long – is included in a series of cartograms (1-431) and corresponds to one lichen taxon or lichenicolous fungus. A single map contains the following data:

- name of species of a lichen, \*lichenicolous fungus or \*saprobiont [(+) – an optional saprobiont], which constitutes the title of the map;
- total number of present sites (np) and total number of historical sites (nh);

– information about Protection status in Poland (acc. to the Journal of Laws. – Dz. U. No 168, item 1765): P - species under strict protection, Pp - under partial protection;

– Świętokrzyskie Mts/Poland Red List Categories (acc. to Cieśliński and Łubek 2003; Cieśliński, Czyżewska, Fabiszewski 2003): Re - Regionally Extinct, CR - Critically Endangered, EN - Endangered, VU - Vulnerable, NT - Near Threatened, LC - Least Concern, DD - Data Deficient;

– subsequent time periods when a species was found are marked with the following symbols: ● – until 1960, ○ – 1961-1980, ▽ – 1981-1985, ● – a species found in all time periods: until 1960, 1961-1980, 1981-1985 to 2002, ● – 1999-2002, ? – lack of exact location.

The list of lichens (cartograms 1 to 405), lichenicolous fungi and saprobionts (406-431) has been arranged alphabetically. The nomenclature follows Scholz (2000), Věžda and Liška (1999), and Hawksworth (1983).

## RESULTS

Taking into consideration the existing results of the research in the area of the SNP, 405 species, subspecies and varieties of lichens and 26 species of lichenicolous fungi have been found. 382 species of lichens and 18 species of lichenicolous fungi have been determined to exist on the basis of the past of the national park and its protection zone. During present research 261 species of lichens have been found, and 144 species of lichens have not been found. The majority should be assumed as extinct in the territory of the national park, the rest may be still found somewhere, but was overlooked during the research.

The whole lichen biota of the national park consists of 119 genera grouped in 50 families and the genus *Lepraria* not belonging to any family. The most abundant in species are the genera: *Cladonia* (37 species), *Lecanora* (31), *Usnea* (13), *Caloplaca* (13), *Micarea* (12), *Pertusaria* (11) and *Chaenotheca* (10).

Lichenicolous fungi have been classified into 21 genera classified into 11 orders. The most abundant in species are the families *Mycohalicaceae* (4 species) and *Tremellaceae* (3) and the genus *Lichenoconium* (3 species). The most numerous group among lichenicolous fungi are species belonging to *Ascomycetes* – 52%. Participation of species belonging to amorphous fungi (*Hyphomycetes* and *Coelomycetes*) – 32% and to *Basidiomycetes* – 16% is smaller.

The most dominant in the lichens biota are epiphytes – more than 220 species and epixyles – 118, which is connected with the forest character of the SNP. The third most numerous group are epilithes – 117 species which are also characterized by the number of exclusive species (86%). Terrestrial lichens – 72 species find habitats specific for them in the national park primarily on the boulder fields where they grow in the soil accumulated among rock blocks. The least number of lichens have been found on bryophytes – only 7 species. A detailed analysis of the lichen biota in the SNP and its protection zone shall be included in a separate work.

**Acknowledgements.** We would like to thank Prof. Krystyna Czyżewska (University of Łódź) for the access herbarial material of lichens from the Świętokrzyski National Park, determining or revising the determination of taxa and critical analysis of this article. We would like to thank Dr Lucyna Śliwa (PAN, Kraków) for determining or revising the determination of the species of *Lecanora dispersa* and *L. umbrina* groups, Dr Martin Kukwa (University of Gdańsk) for determining or revising by TLC the determination of the species of *Lepraria*.

The work was partly supported by the Committee for Scientific Research, research project no 6 P04G 077 20 and University of Łódź, grants no 505/451, 505/413.

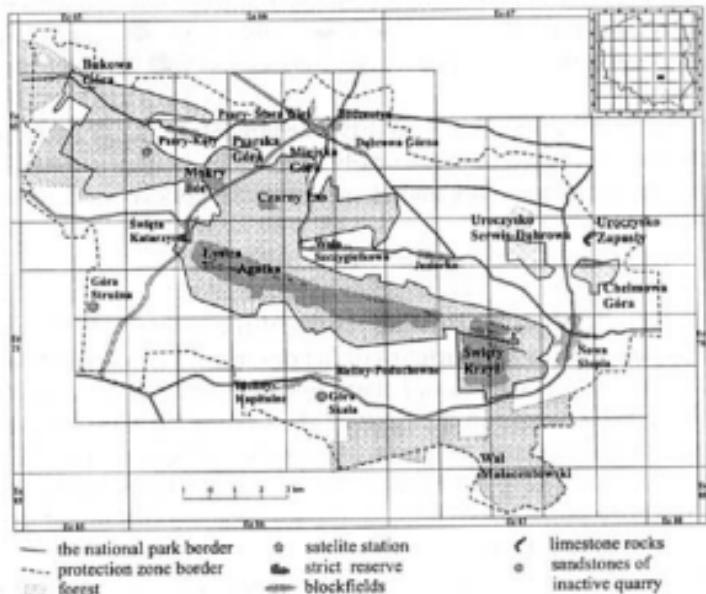


Fig. 1. Świętokrzyski National Park and its protecting zone.

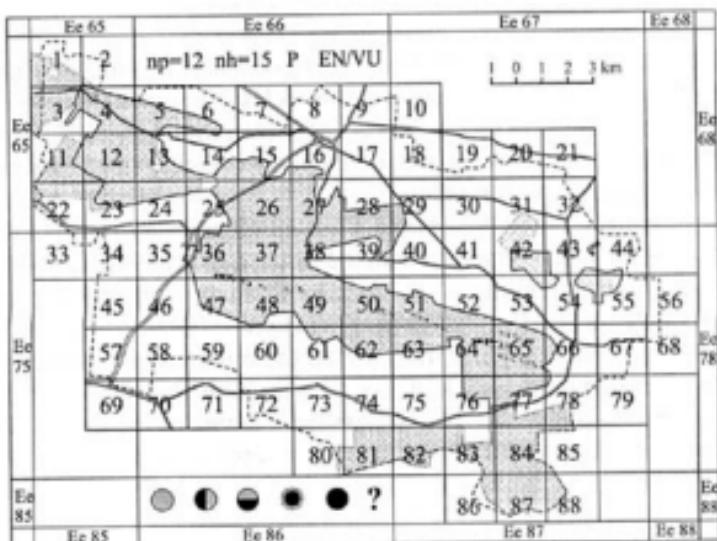
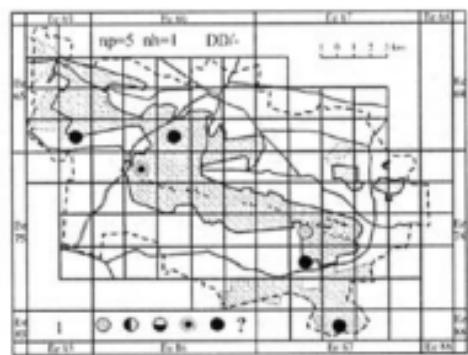
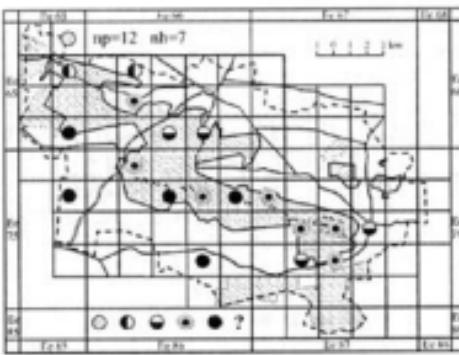
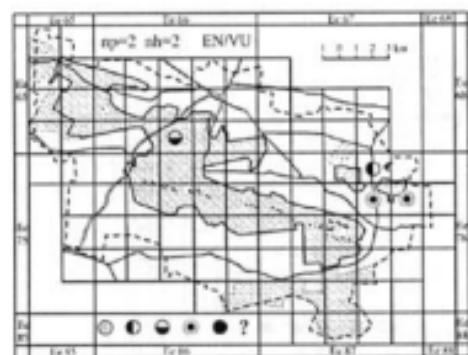
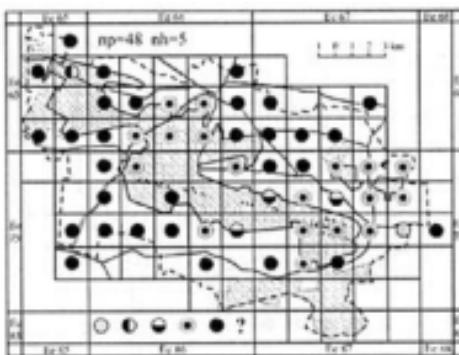
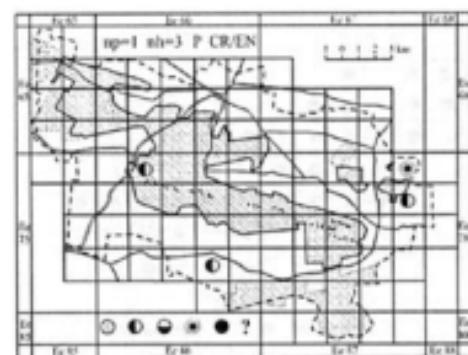
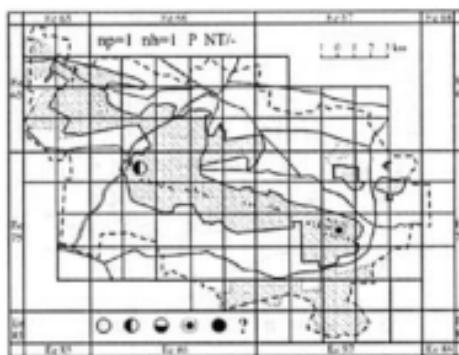
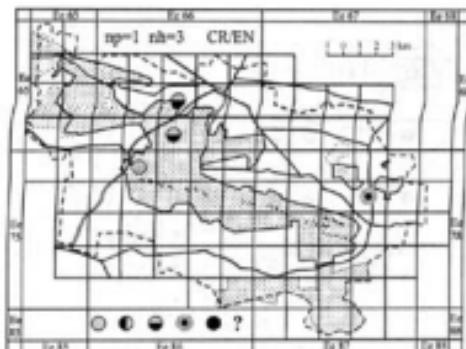
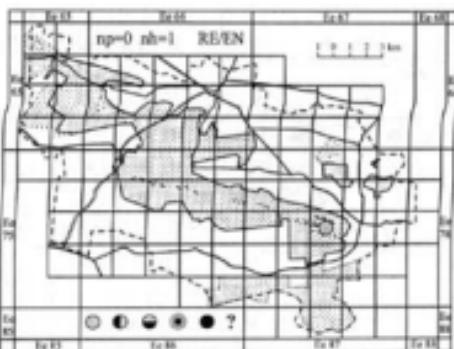
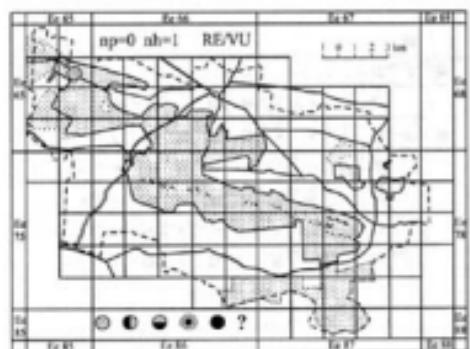
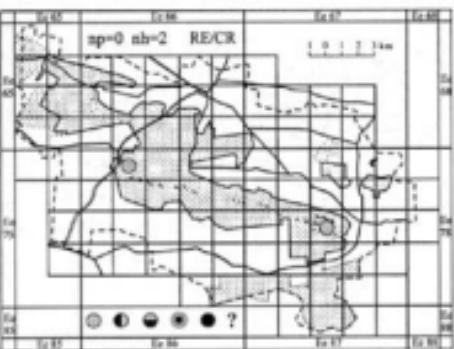
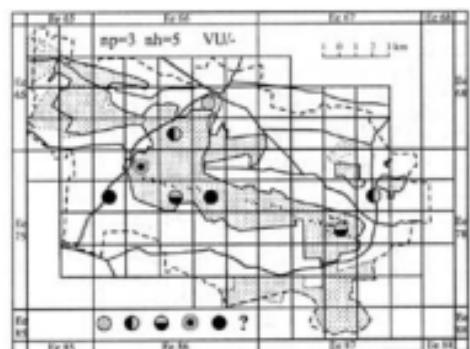
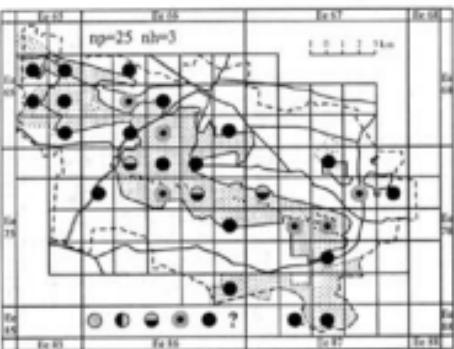
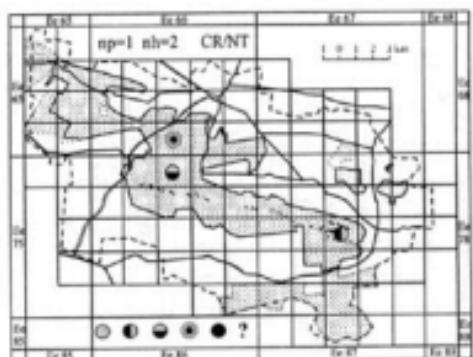
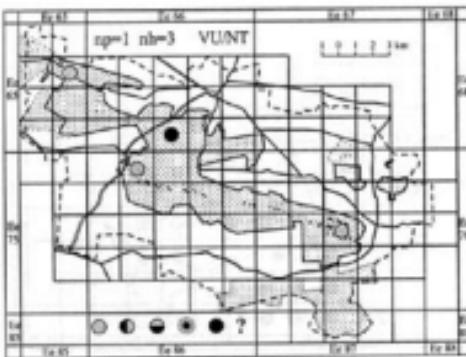
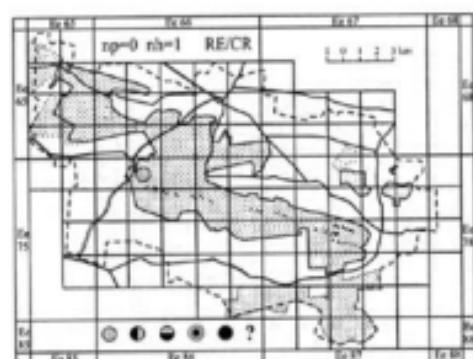
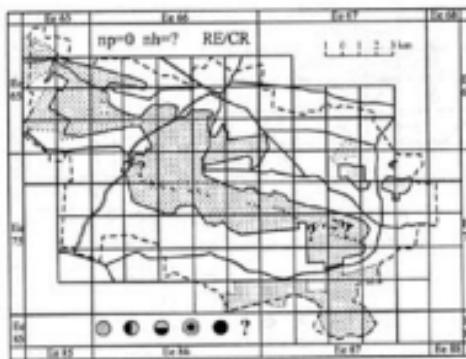
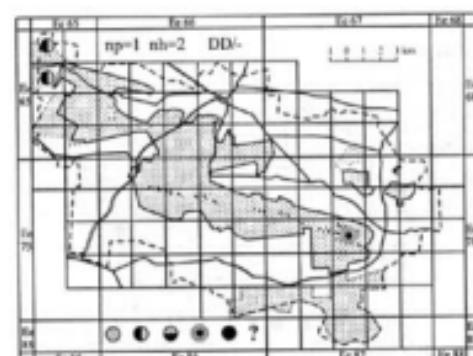
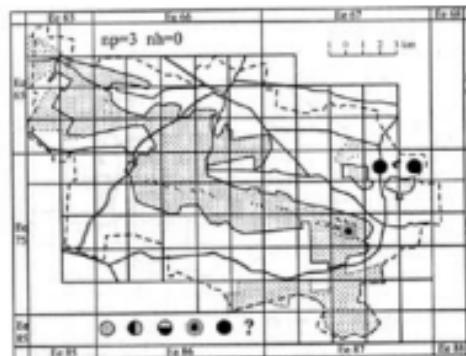


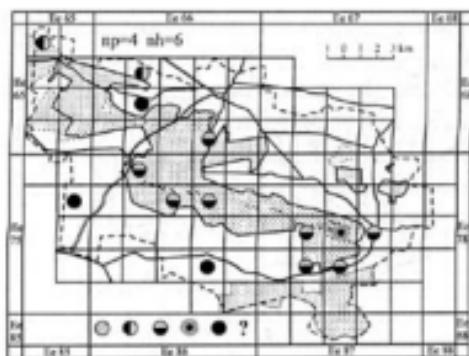
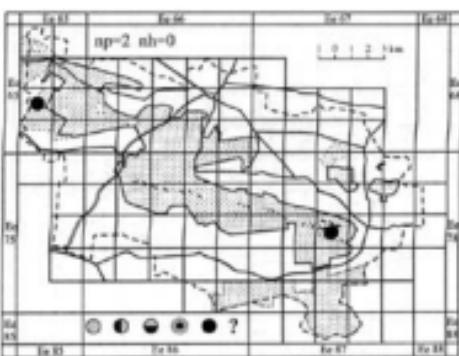
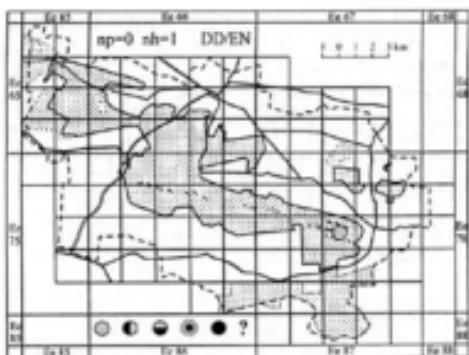
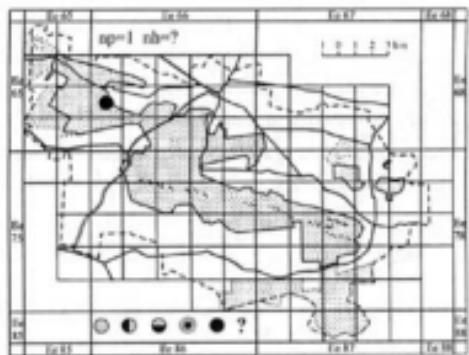
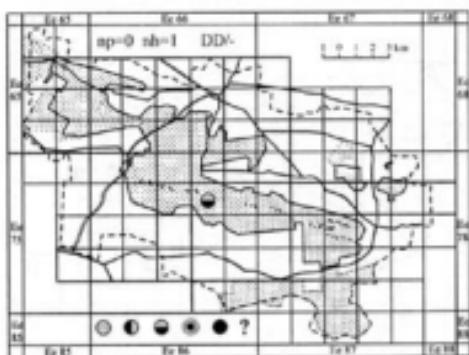
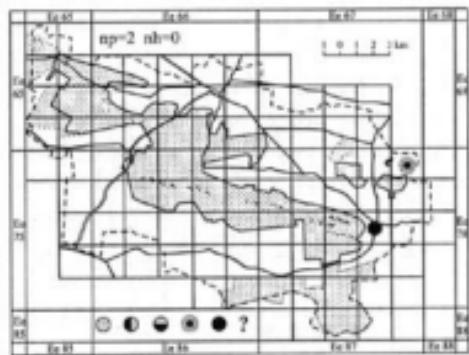
Fig. 2. A single cartogram, and their numbers.

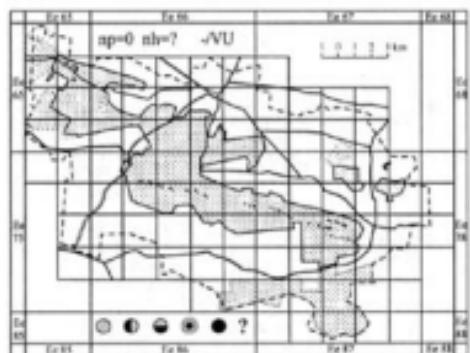
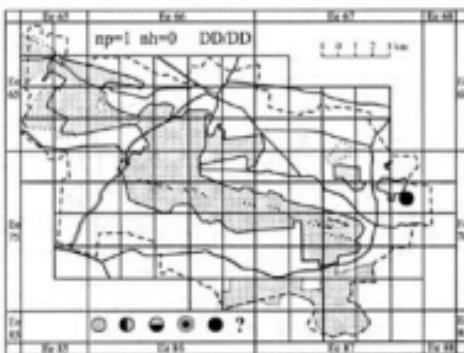
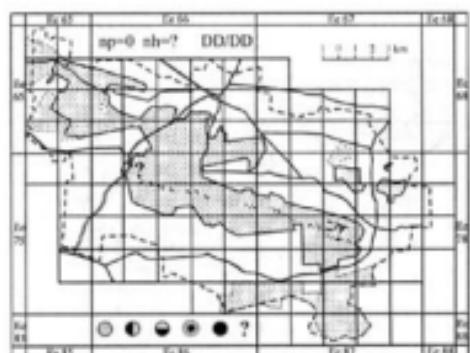
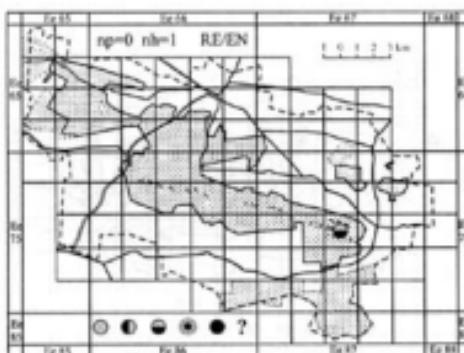
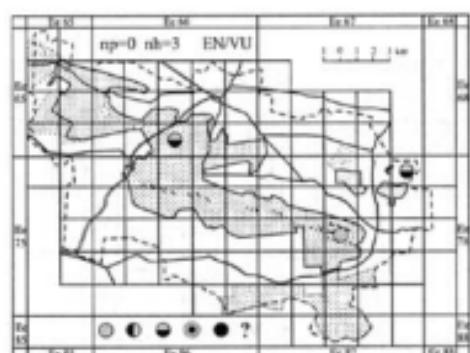
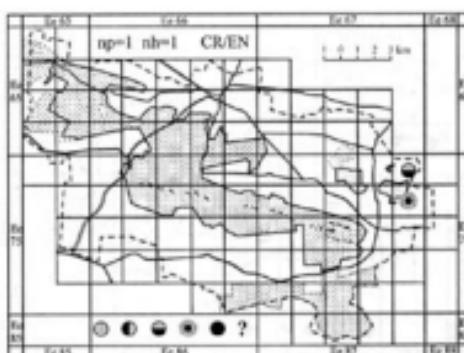
## Distribution of lichens and lichenicolous fungi

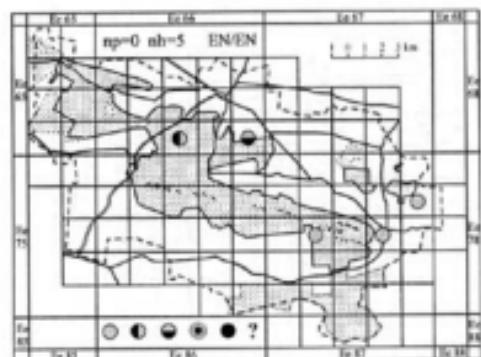
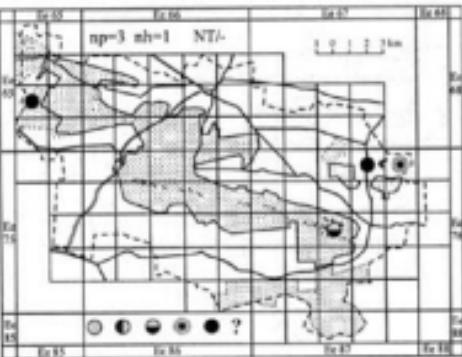
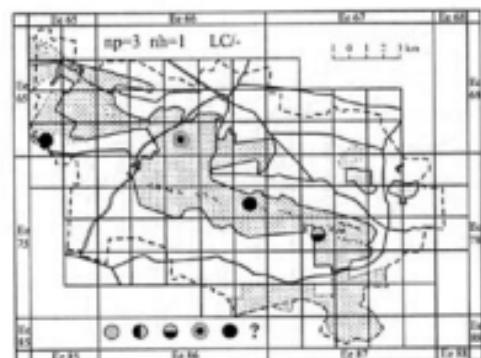
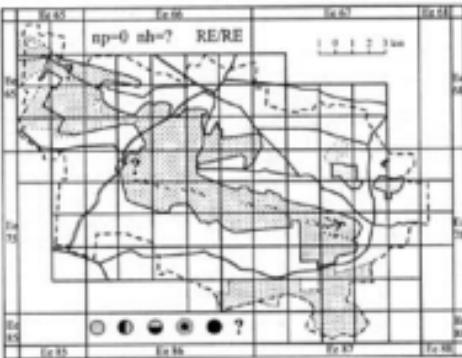
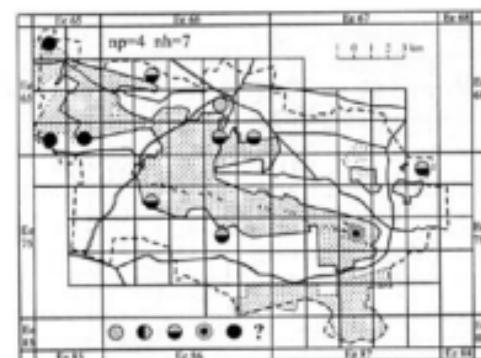
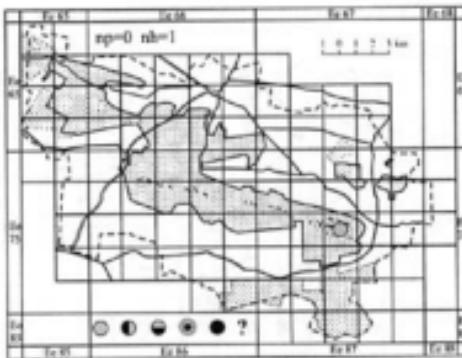
*Absconditella lignicola* Věžda & Pišút*Acarospora fuscata* (Nyl.) Th. Fr.*Acrocordia gemmata* (Ach.) A. Massal.*Amandinea punctata* (Hoffm.) Coppins & Scheid.*Anaptychia ciliaris* (L.) Körb. ex A. Massal.*Arctoparmelia incurva* (Pers.) Hale

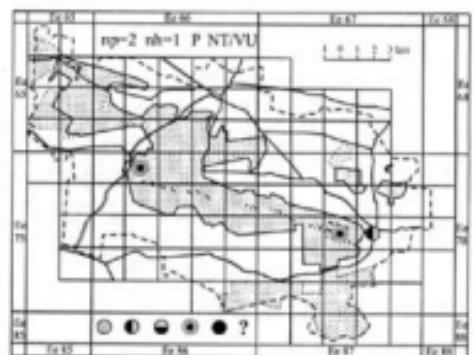
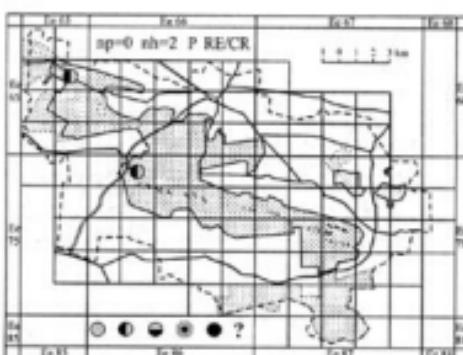
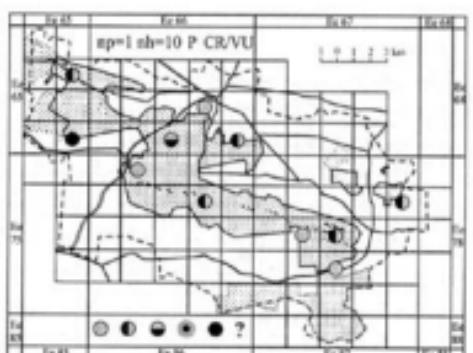
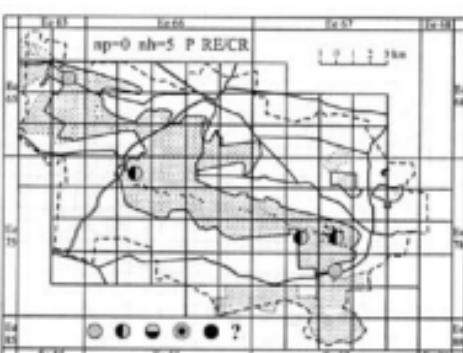
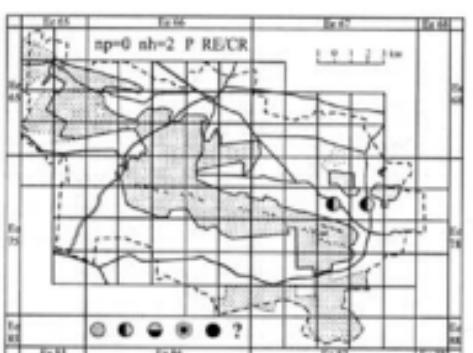
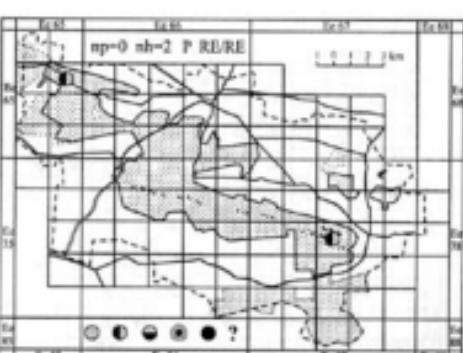
*Arthonia byssacea* (Weigel) Almq.*Arthonia didyma* Körb.*Arthonia dispersa* (Schrad.) Nyl.*Arthonia fuliginosa* (Turner & Borrer) Flot.*Arthonia radiata* (Pers.) Ach.*Arthonia spadicea* Leight.

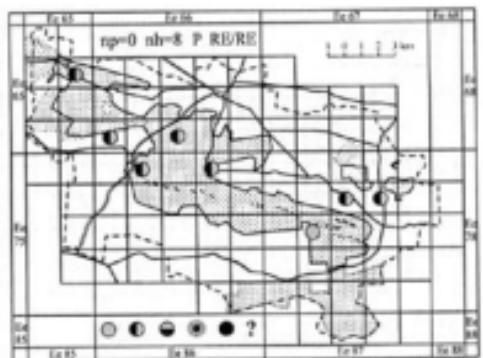
*Arthonia vinosae* Leight.*Arthothelium ruuanum* (A. Massal.) Körb.*Arthothelium spectabile* Flot. ex A. Massal.*Arthrosporum populorum* A. Massal.*Aspicilia caesiocinerea* (Nyl. ex Malbr.) Arnold*Aspicilia calcarea* (L.) Mudd

*Aspicilia cinerea* (L.) Körb.*Aspicilia contorta* (Hoffm.) Kremp.*Aspicilia gibbosa* (Ach.) Körb.*Aspicilia moenium* (Vain.) G. Thor & Timdal*Aspicilia simoënsis* Räsänen*Bacidia bagliettoana* (A. Massal. & De Not.) Jatta

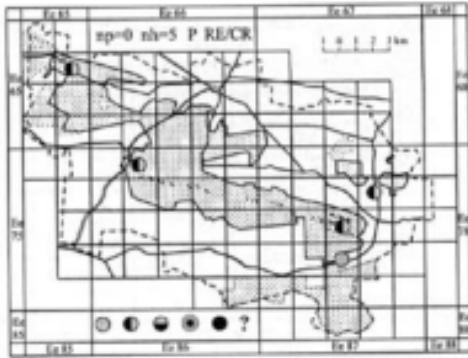
*Bacidia beckhausii* Körb.*Bacidia fraxinea* Lönne.*Bacidia friesiana* (Hepp) Körb.*Bacidia rosella* (Pers.) De Not.*Bacidia rubella* (Hoffm.) A. Massal.*Bacidia subincompta* (Nyl.) Arnold.

*Bacidina assulata* Vězda*Bacidina inundata* (Fr.) Vězda*Bacidina phacodes* (Körb.) Vězda*Baeomyces carneus* Flörke*Baeomyces rufus* (Huds.) Rebent.*Bagliettoa parviflora* (J. Steiner) Vězda & Poelt

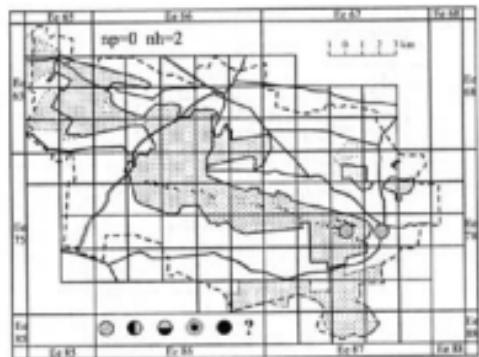
*Brodoa intestiniformis* (Vill.) Goward*Bryoria capillaris* (Ach.) Brodo & D. Hawksw.*Bryoria fuscescens* (Gyeln.) Brodo & D. Hawksw.*Bryoria implexa* (Hoffm.) Brodo & D. Hawksw.*Bryoria mirabilis* (Motyka) Bystr.*Bryoria motykana* (Bystr.) Bystr.



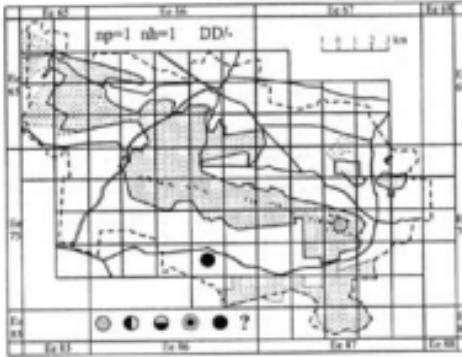
*Bryoria subcana* (Nyl. ex Stizenb.) Brodo & D. Hawksw.



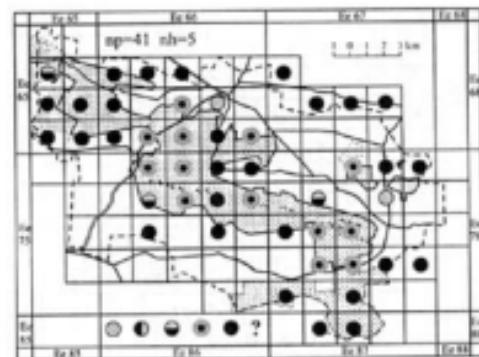
*Bryoria vrangiana* (Gyeln.) Brodo & D. Hawksw.



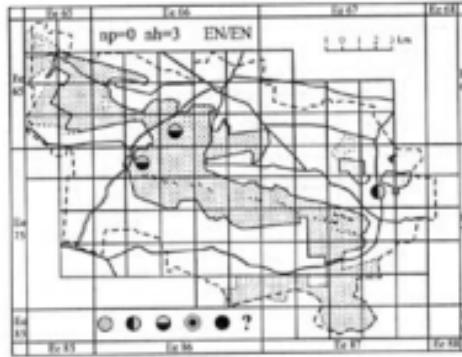
*Buellia alboatra* (Hoffm.) Th. Fr.



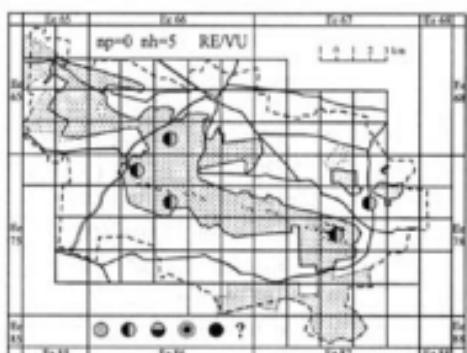
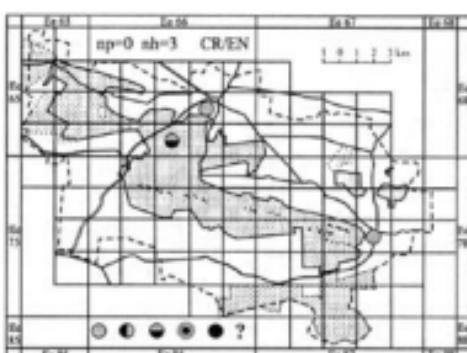
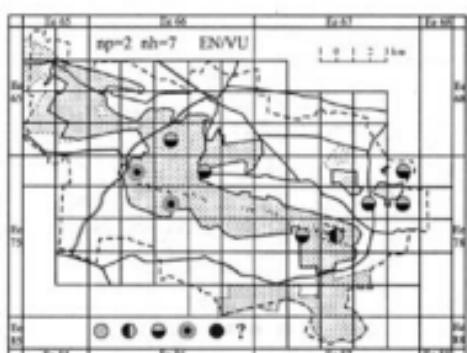
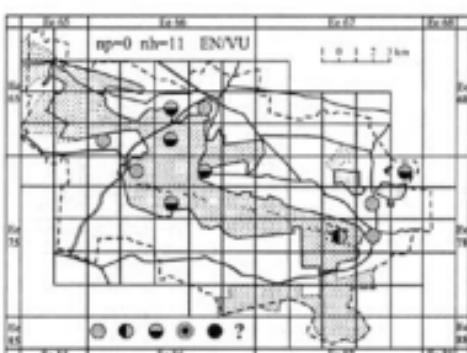
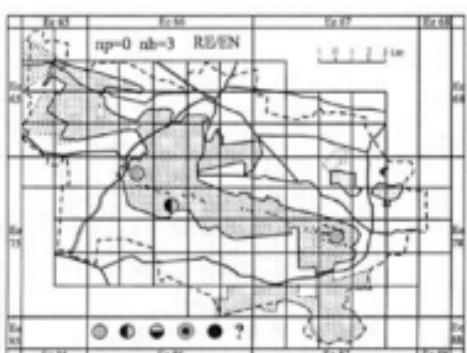
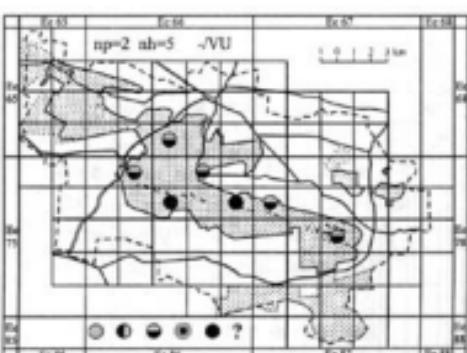
*Buellia aethalea* (Ach.) Th. Fr.

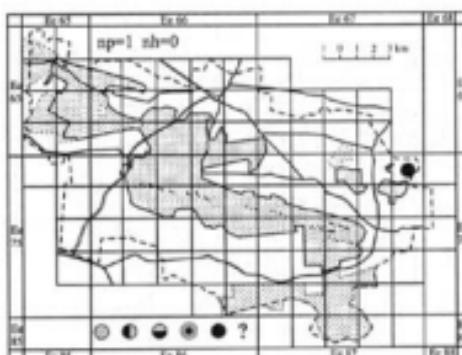
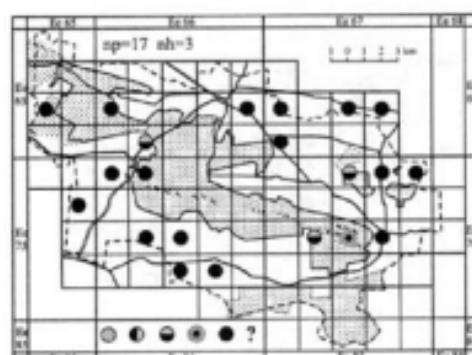
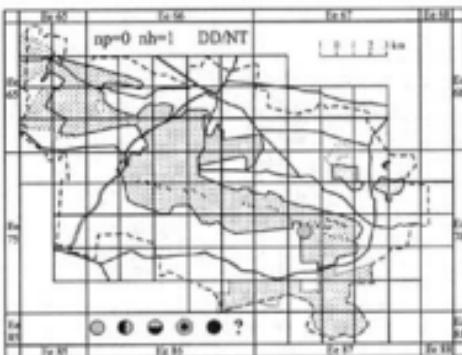
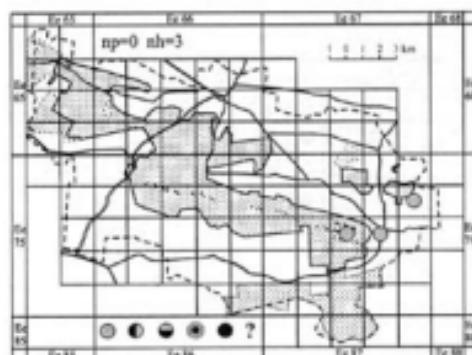
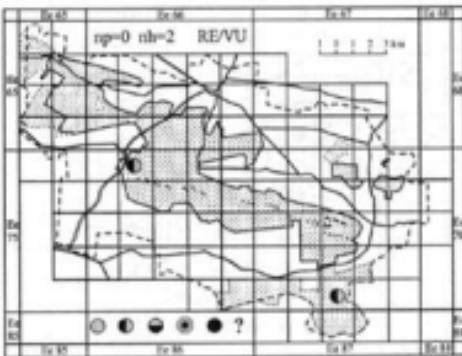
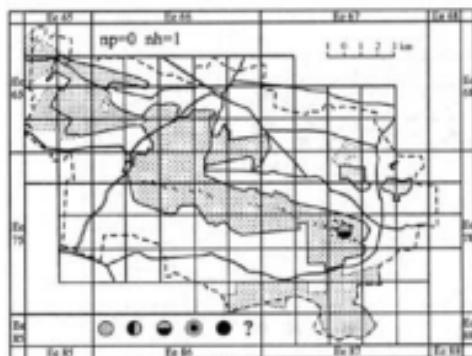


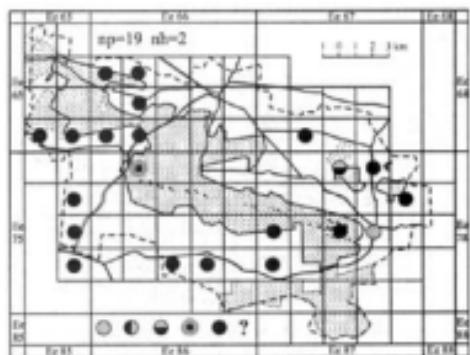
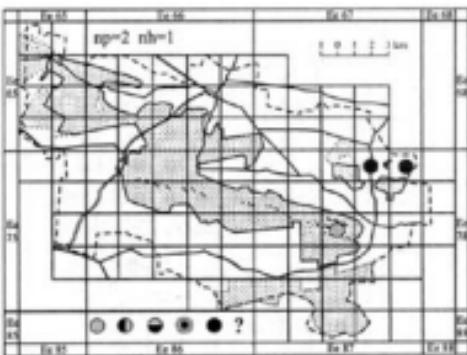
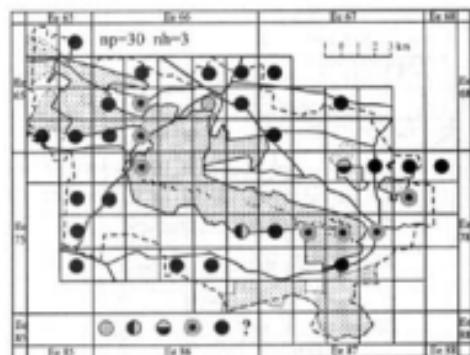
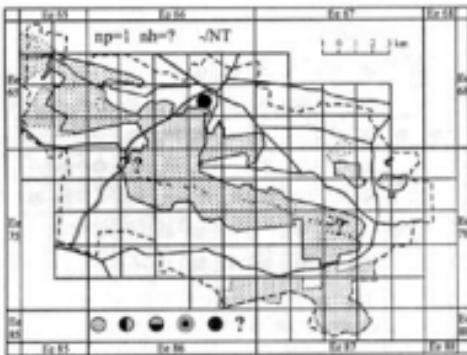
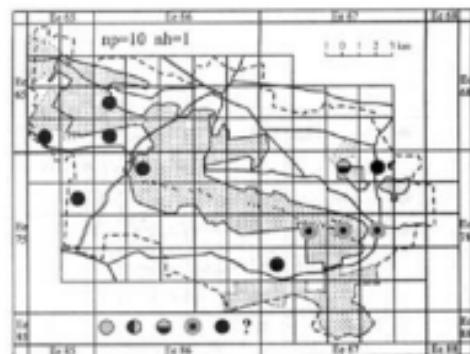
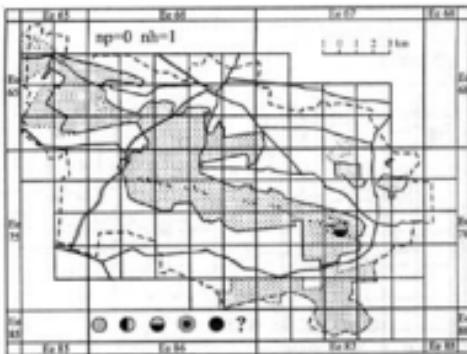
*Buellia griseovirens* (Turner & Borrer ex Sm.) Almib.

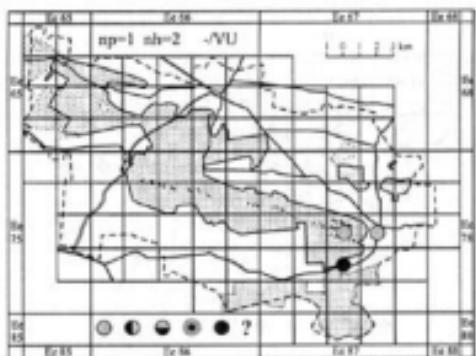
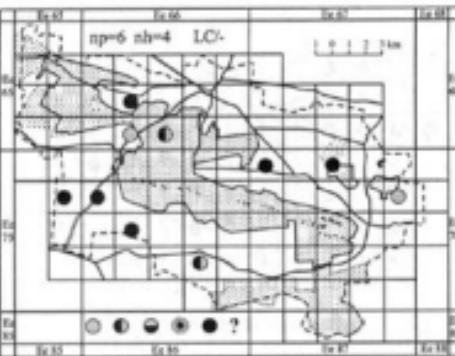
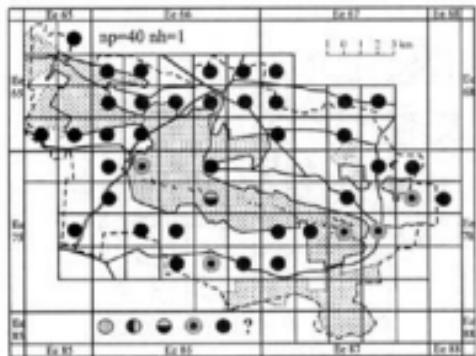
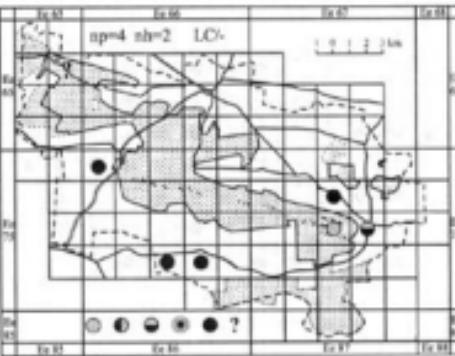
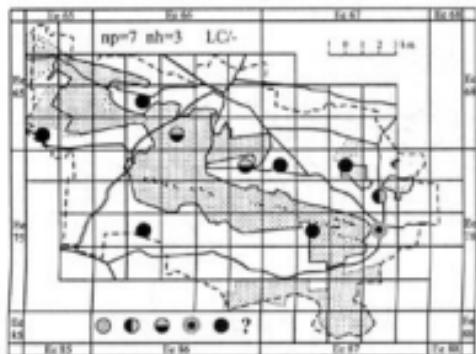
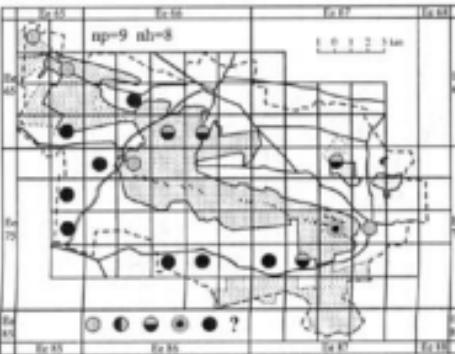


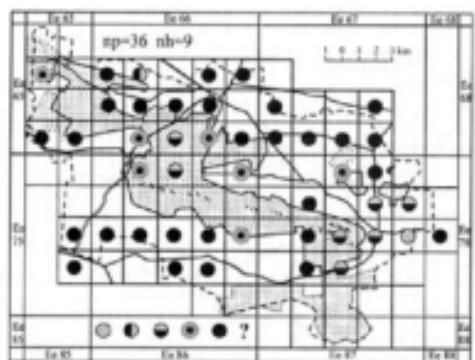
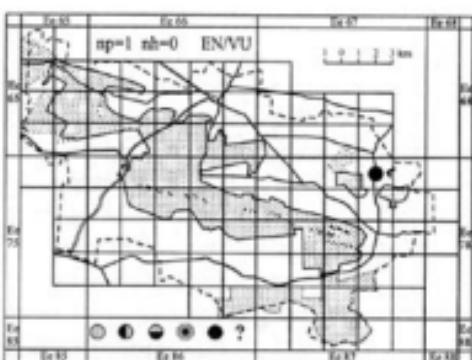
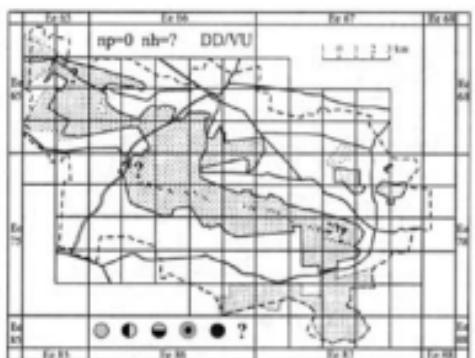
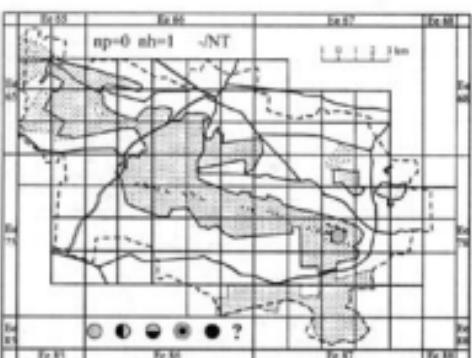
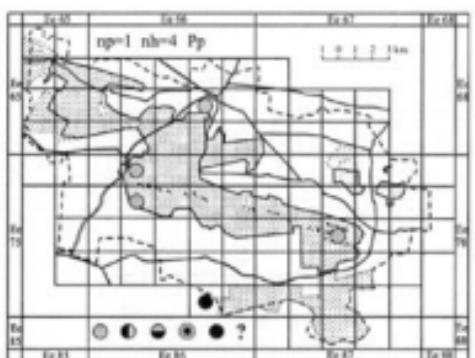
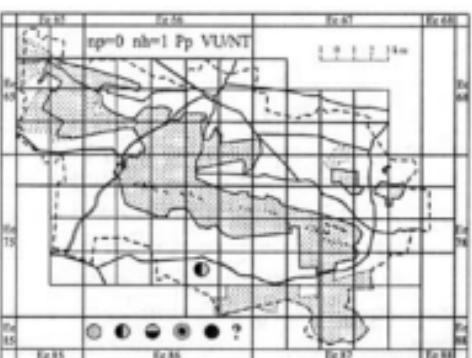
*Buellia schaeferi* De Not.

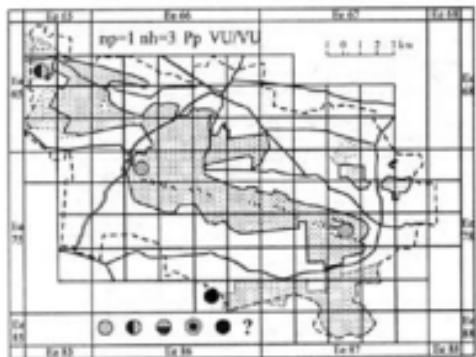
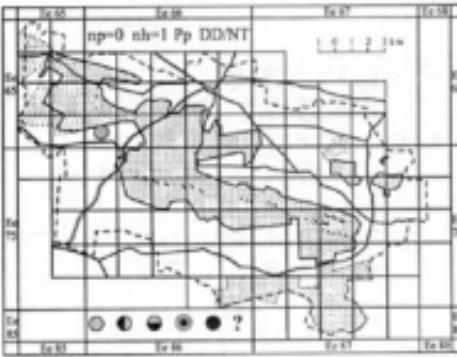
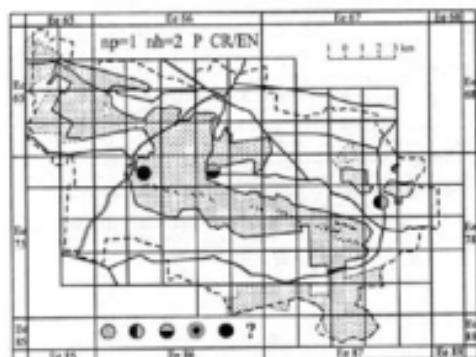
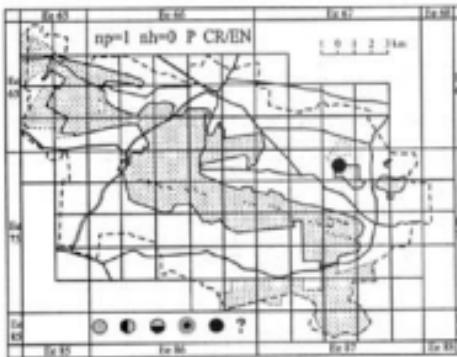
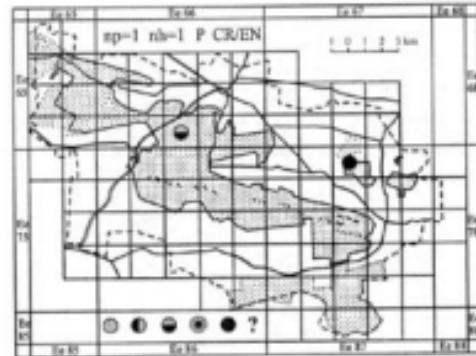
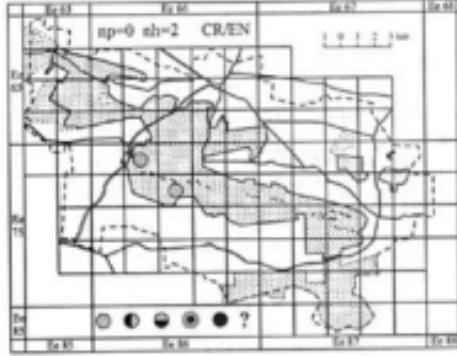
*Calicium abietinum* Pers.*Calicium adspersum* Pers.*Calicium glaucellum* Ach.*Calicium salicinum* Pers.*Calicium trabinellum* (Ach.) Ach.*Calicium viride* Pers.

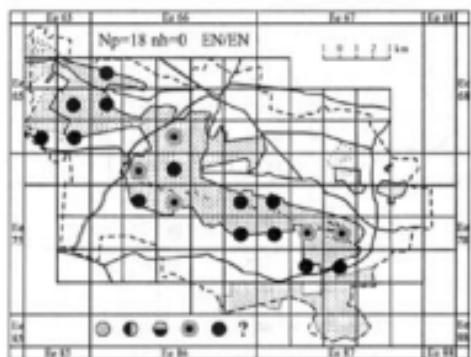
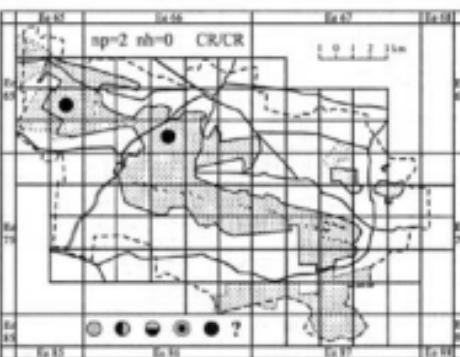
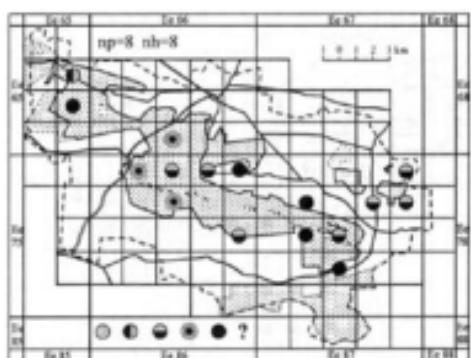
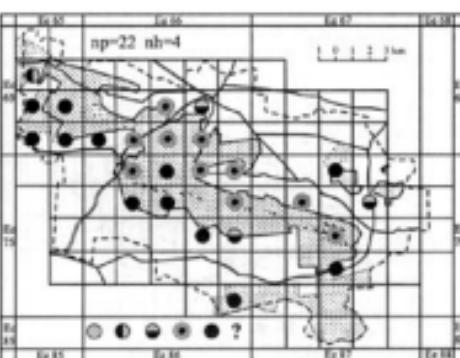
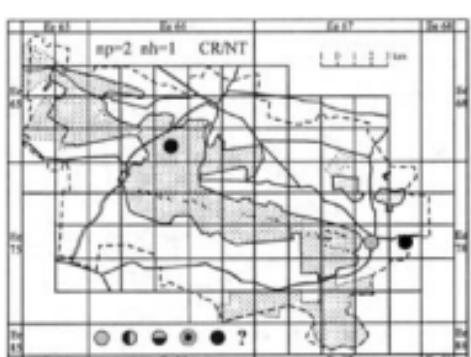
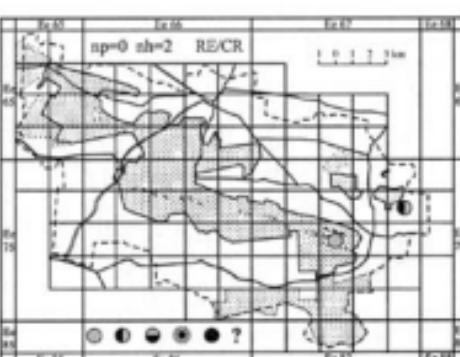


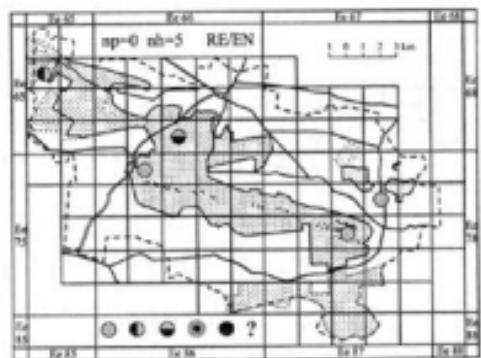
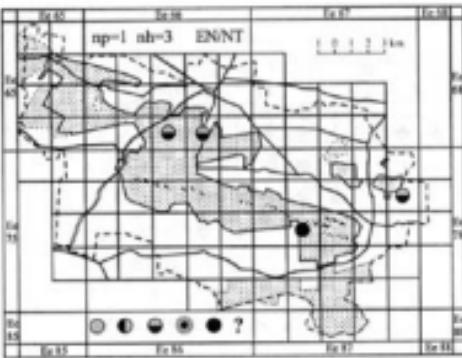
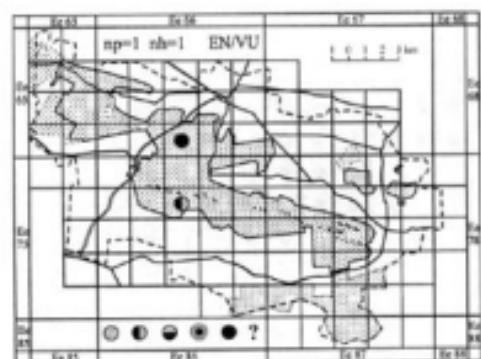
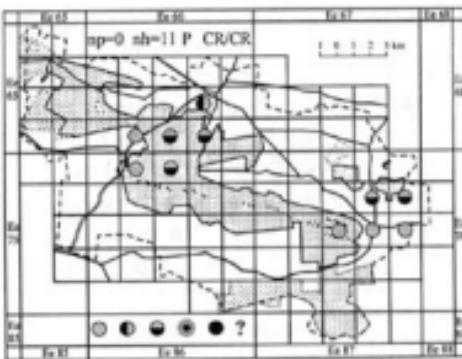
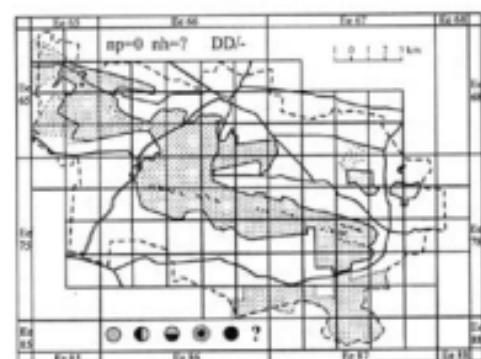
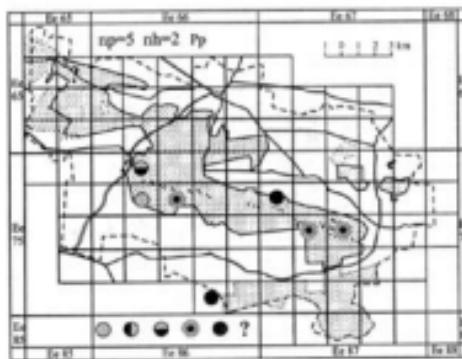
*Caloplaca decipiens* (Arnold) Blomb. & Fössell*Caloplaca dolomitica* (Hue) Zahlbr.*Caloplaca holocarpa* (Hoffm. ex Ach.) Wade*Caloplaca obscurella* (J. Lahm ex Körb.) Th. Fr.*Caloplaca saxicola* (Hoffm.) Nordin*Caloplaca teicholyta* (Ach.) J. Steiner

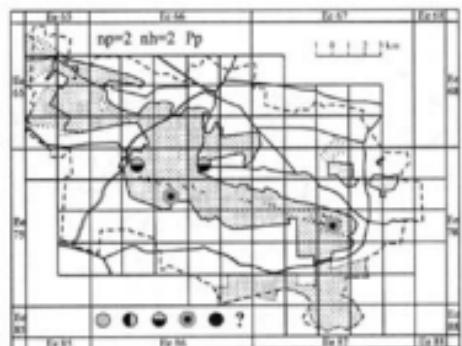
*Caloplaca variabilis* (Pers.) Müll. Arg.*Candelaria concolor* (J. Dicks.) Stein*Candelariella aurella* (Hoffm.) Zahlbr.*Candelariella coralliza* (Nyl.) H. Magn.*Candelariella reflexa* (Nyl.) Lettau*Candelariella vitellina* (Hoffm.) Müll. Arg.

*Candelariella xanthostigma* (Ach.) Lettau*Catapyrenium cinereum* (Pers.) Körb.*Catillaria chalybeia* (Borrer) A. Massal.*Catillaria lenticularis* (Ach.) Th. Fr.*Cetraria aculeata* (Schreb.) Fr.*Cetraria ericetorum* Opiz

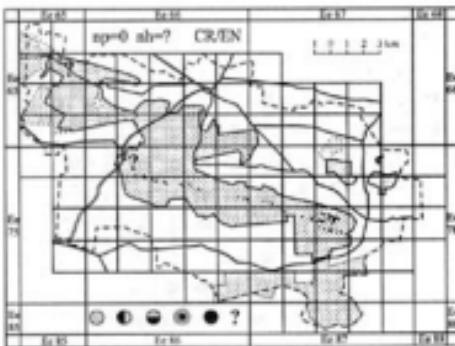
*Cetraria islandica* (L.) Ach.*Cetraria muricata* (Ach.) Eckfeldt*Cetraria sepincola* (Ehrh.) Ach.*Cetrelia cetrarioides* (Delise ex Duby) W.L. Culb. & C.F. Culb.*Cetrelia olivetorum* (Nyl.) W.L. Culb. & C.F. Culb.*Chaenotheca brachypoda* (Ach.) Tibell

*Chaenotheca brunneola* (Ach.) Müll. Arg.*Chaenotheca chlorella* (Ach.) Müll. Arg.*Chaenotheca chrysoccephala* (Turner ex Ach.) Th. Fr.*Chaenotheca ferruginea* (Turner & Borrer) Mig.*Chaenotheca furfuracea* (L.) Tibell*Chaenotheca hispidula* (Ach.) Zahlbr.

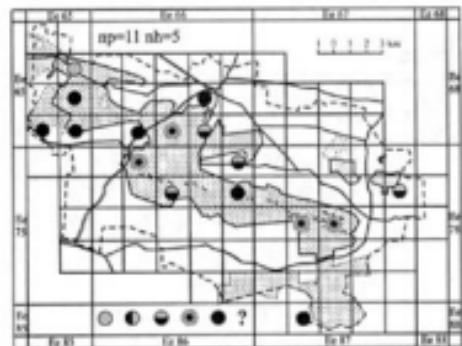
*Chaenotheca phaeocephala* (Turner) Th. Fr.*Chaenotheca trichialis* (Ach.) Th. Fr.*Chaenotheca xylooxena* Nádv.*Chrysotrichia candelaris* (L.) J.R. Laundon*Chrysotrichia chlorina* (Ach.) J.R. Laundon*Cladonia arbuscula* (Wallr.) Flot. ssp. *arbuscula*



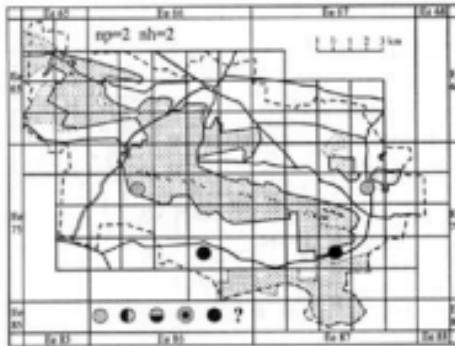
*Cladonia arbuscula* (Wallr.) Flot. ssp. *mitis*  
(Sandst.) Ruoss



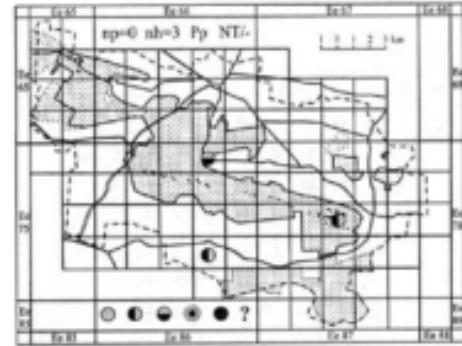
*Cladonia botrytes* (K.G. Hagen) Willd.



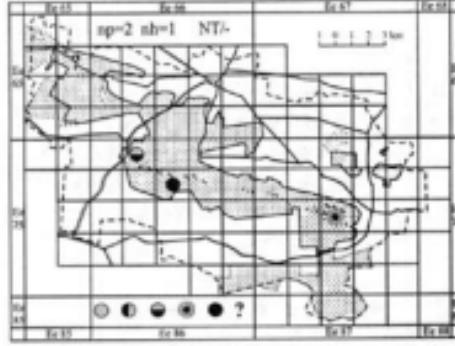
*Cladonia ciliata* (Ach.) Stirt. var. *tenuis* (Flörke) Ahti



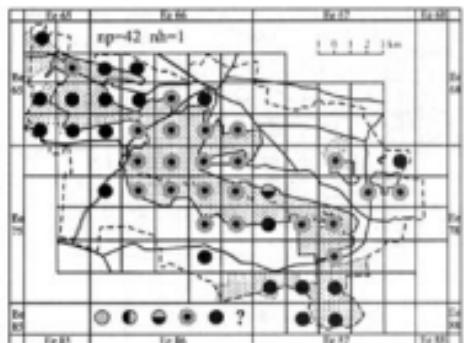
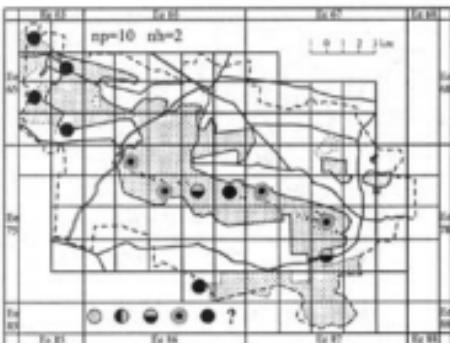
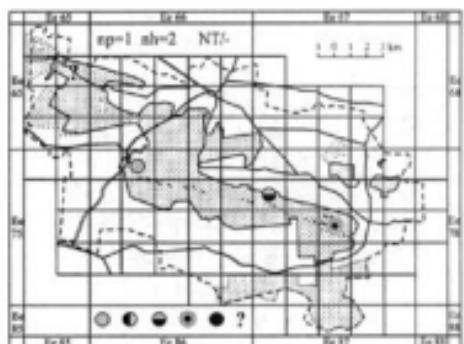
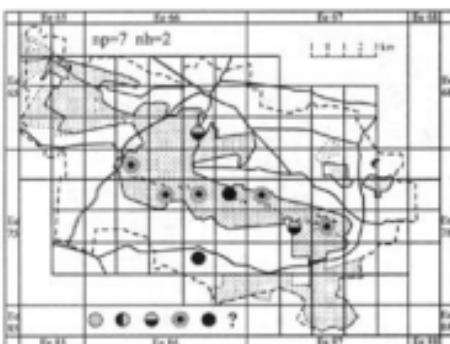
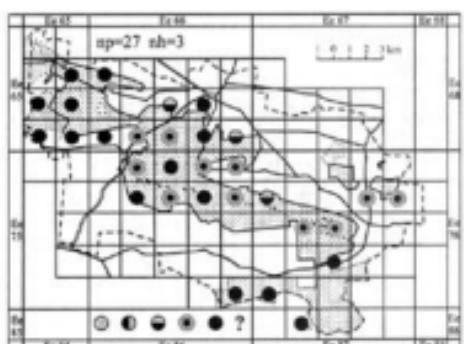
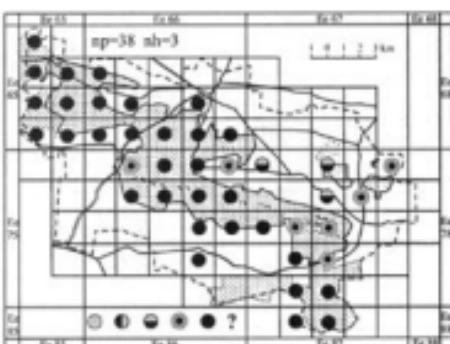
*Cladonia cervicornis* (Ach.) Flot. ssp. *verticillata* (Hoffm.) Ahti

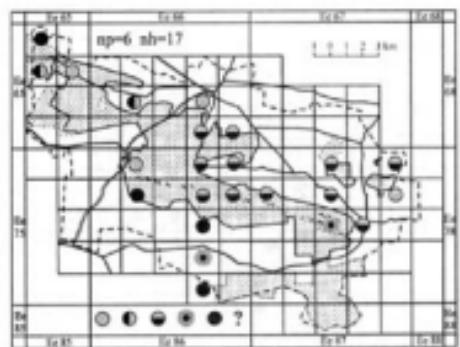
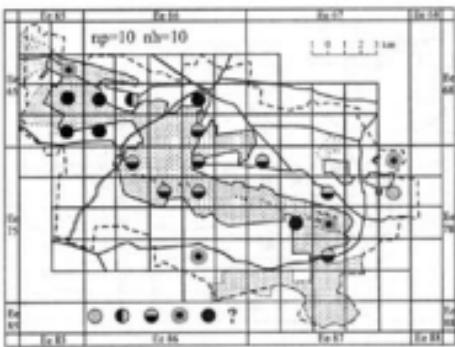
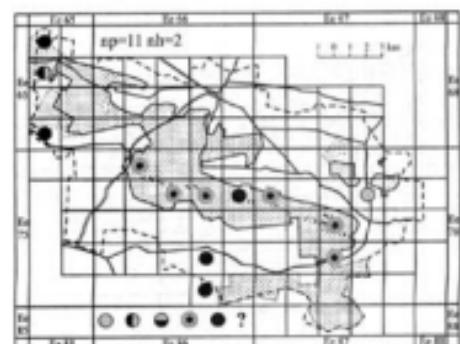
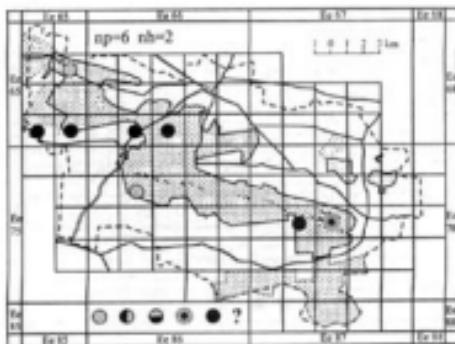
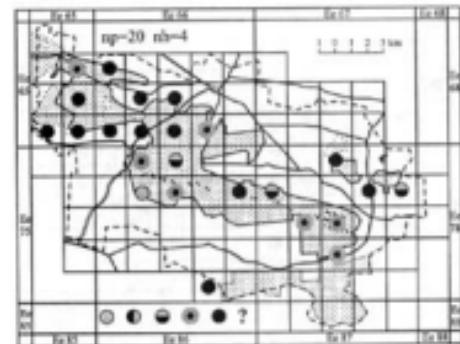
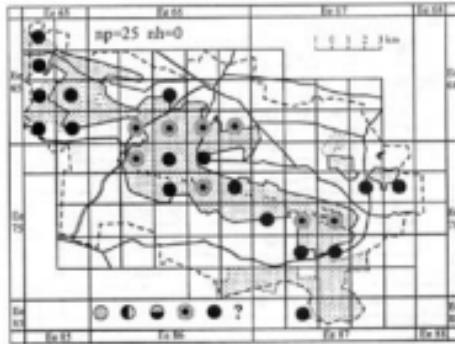


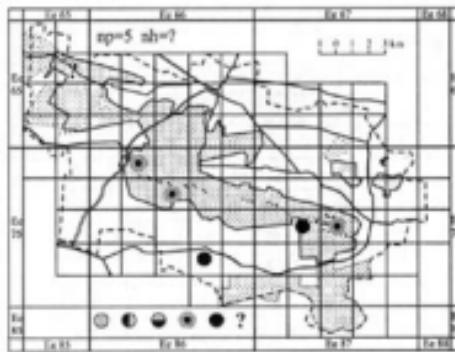
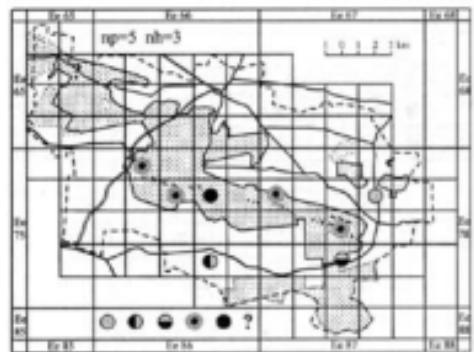
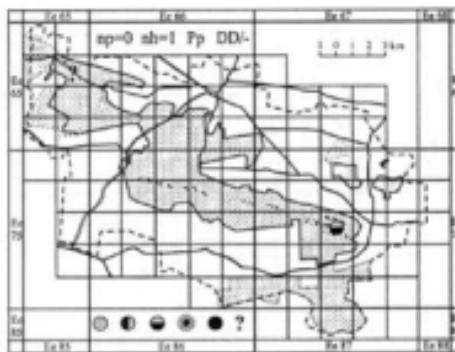
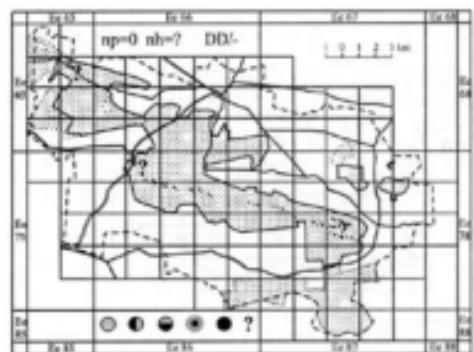
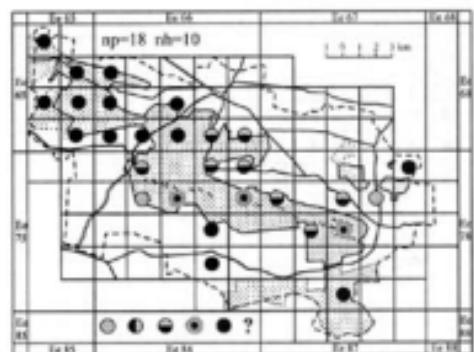
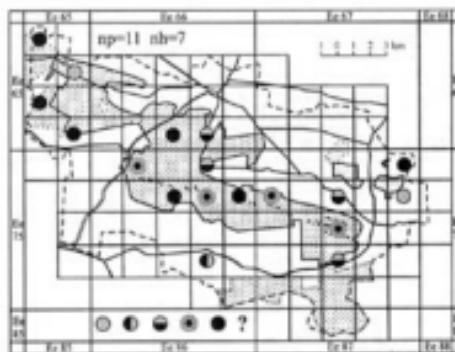
*Cladonia ciliata* Stirt. var. *tenuis* (Flörke) Ahti

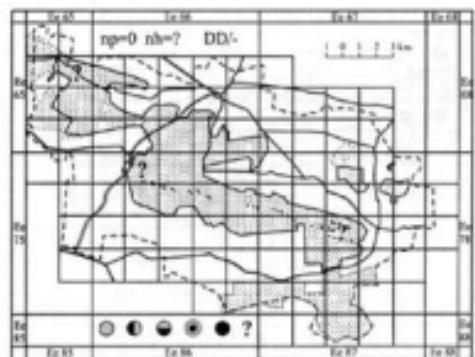
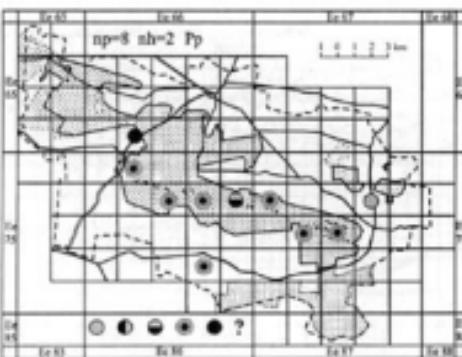
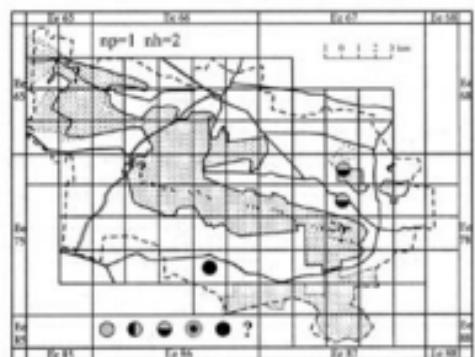
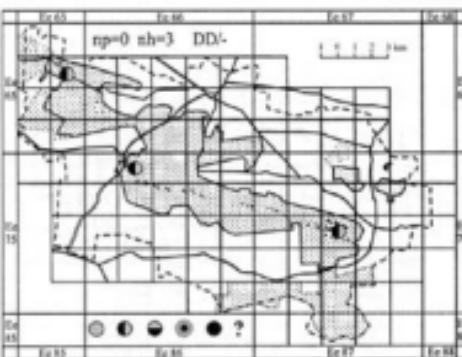
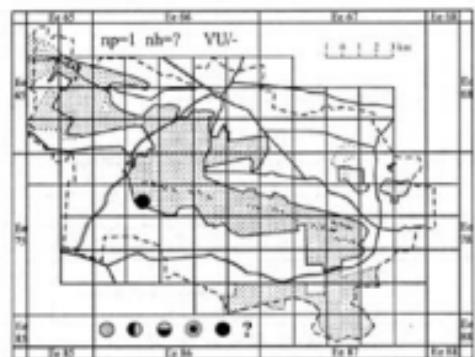
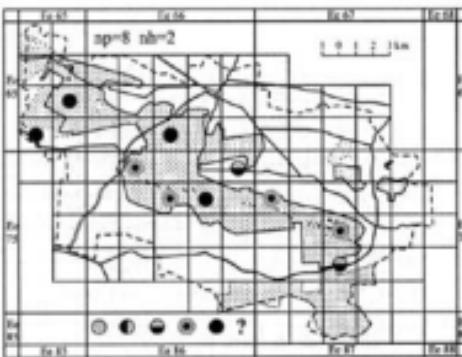


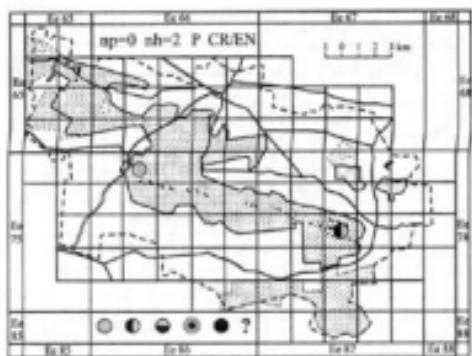
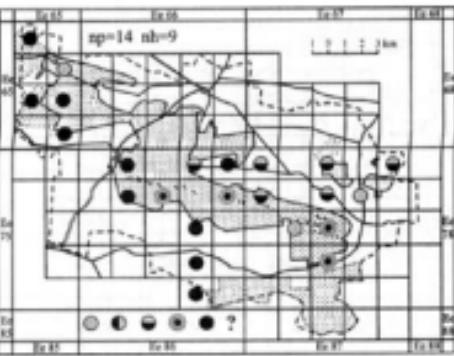
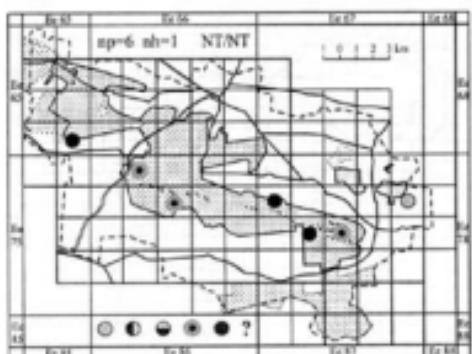
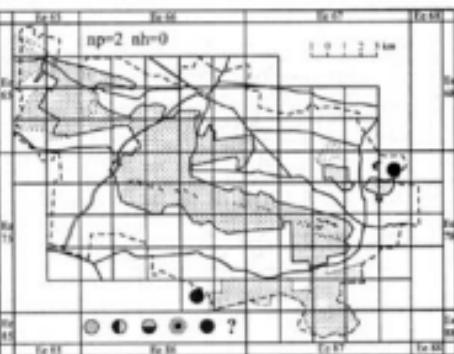
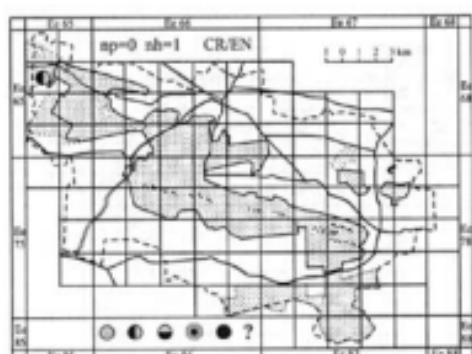
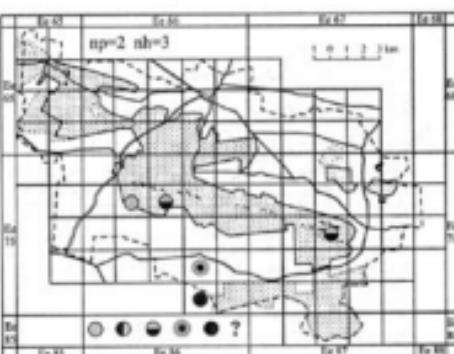
*Cladonia coccifera* (L.) Willd.

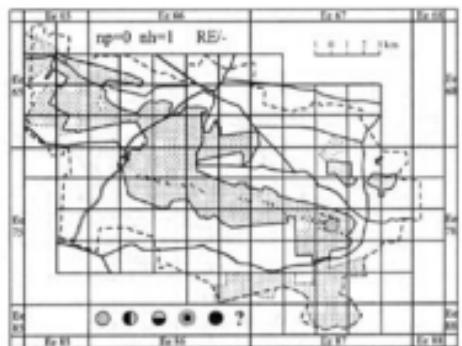
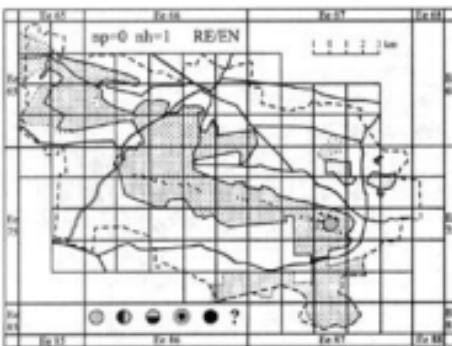
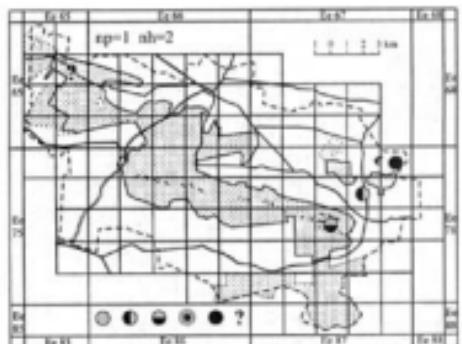
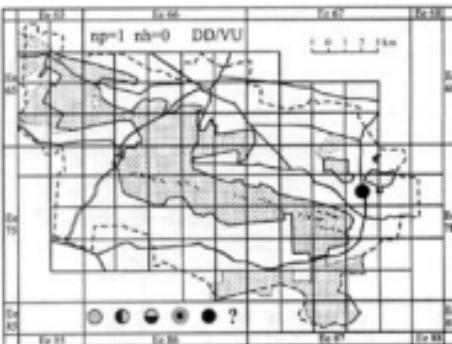
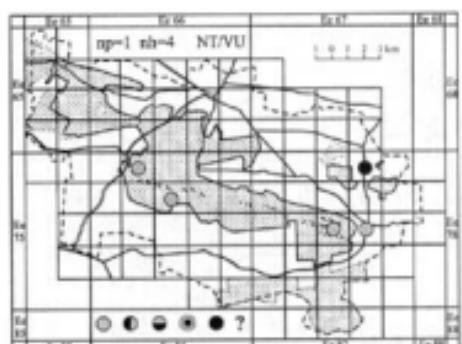
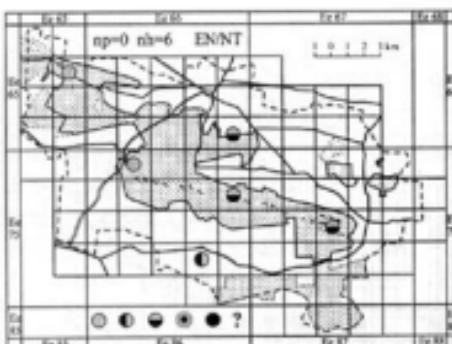
*Cladonia coniocraea* aust.*Cladonia cornuta* (L.) Hoffm.*Cladonia crispata* (Ach.) Flot.*Cladonia deformis* (L.) Hoffm.*Cladonia digitata* (L.) Hoffm.*Cladonia fimbriata* (L.) Fr.

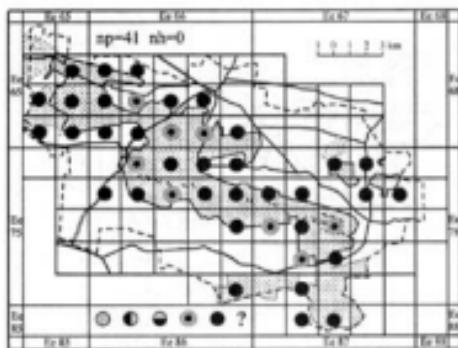
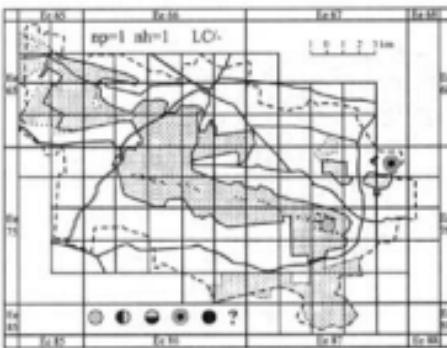
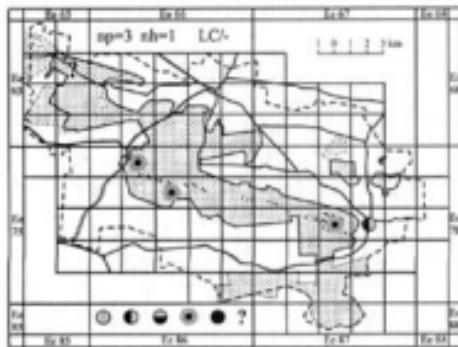
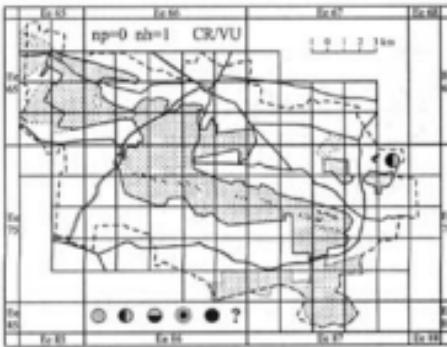
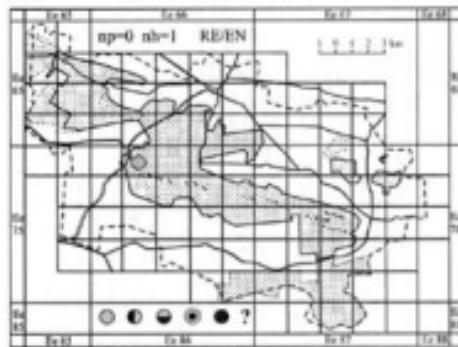
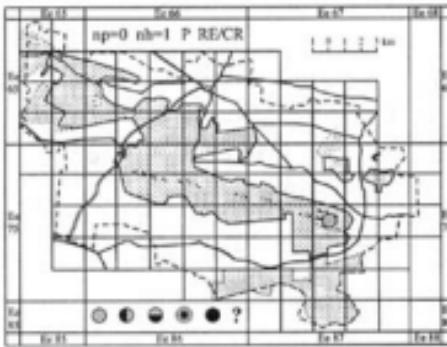
*Cladonia furcata* (Huds.) Schrad.*Cladonia glauca* Flörke*Cladonia gracilis* (L.) Willd.*Cladonia macilenta* Hoffm. ssp. *floerkeana* (Fr.) V. Wirth*Cladonia macilenta* Hoffm. ssp. *macilenta**Cladonia ochrochlora* Flörke

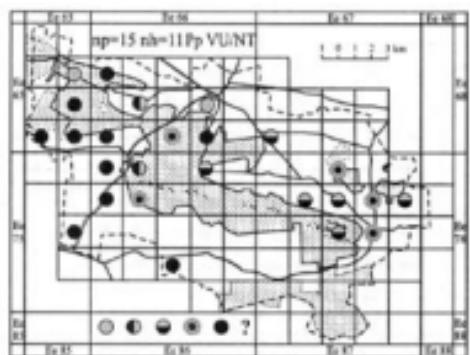
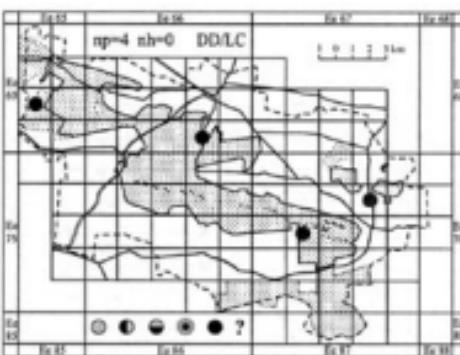
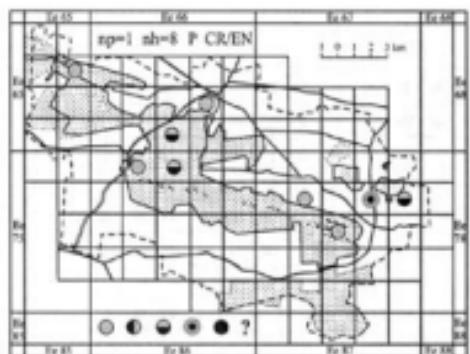
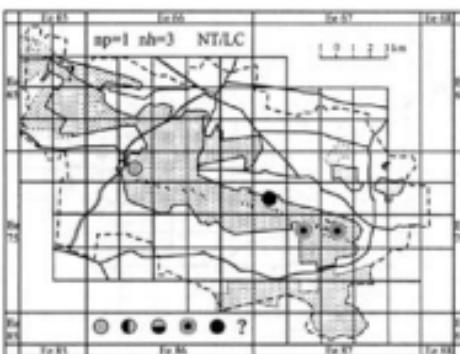
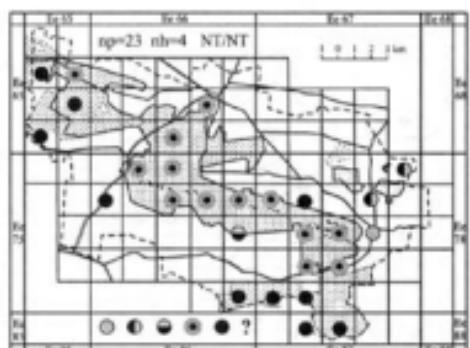
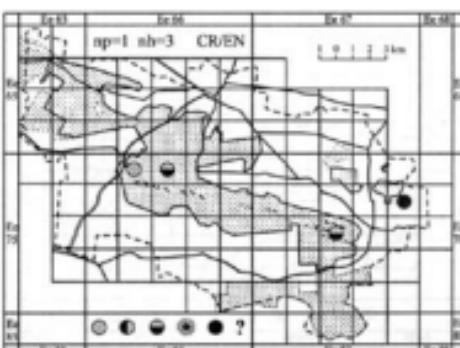
*Cladonia phyllophora* Hoffm.*Cladonia pleurota* (Flörke) Schaeer.*Cladonia polydactyla* (Flörke) Spreng.*Cladonia portentosa* (Dufour) Coem.*Cladonia pyxidata* (L.) Hoffm.  
ssp. *chlorophphaea* (Sommerf.) V. Wirth s.l.*Cladonia pyxidata* (L.) Hoffm. ssp. *pyxidata*

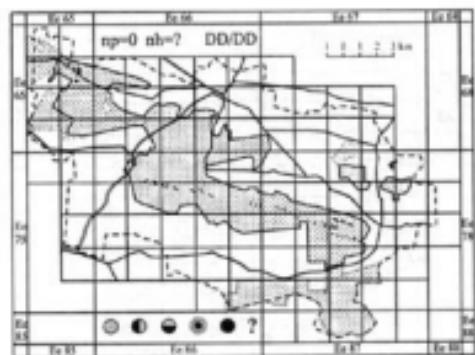
*Cladonia ramulosa* (With.) J.R. Laundon*Cladonia rangiferina* (L.) Weber ex F.H. Wigg.*Cladonia rangiformis* Hoffm.*Cladonia rei* Schaeer.*Cladonia scabriuscula* (Delise) Nyl.*Cladonia squamosa* (Scop.) Hoffm.  
var. *squamosa*

*Cladonia stellaris* (Opiz) Pouzar & Věžda*Cladonia subulata* (L.) Weber ex F.H. Wigg.*Cladonia sulphurina* (Michx.) Fr.*Cladonia symphycarpa* (Flörke) Fr.*Cladonia turgida* Hoffm.*Cladonia uncialis* (L.) Weber ex F.H. Wigg

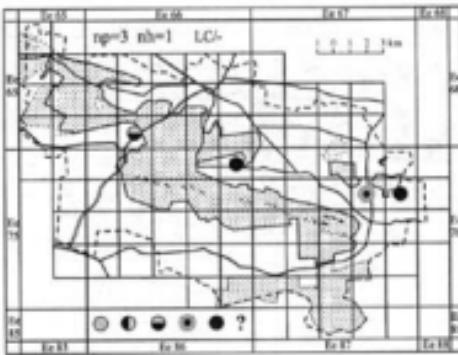
*Collema crispum* (Huds.) Weber ex F.H. Wigg.*Collema flaccidum* (Ach.) Ach.*Collema tenax* (Sw.) Ach. em. Degel.*Cyphellium notarisi* (Tul.) Blomb. & Frossell*Dermatocarpon miniatum* (L.) W. Mann*Dibaeis baeomyces* (L.fil.) Rambold & Hertel

*Dimerella pineii* (Ach.) Vézda*Diploschistes muscorum* (Scop.) R. Sant.*Diploschistes scruposus* (Schreb.) Norm.*Endocarpon pusillum* Hedw.*Ephæbe lanata* (L.) Vain.*Evernia divaricata* (L.) Ach.

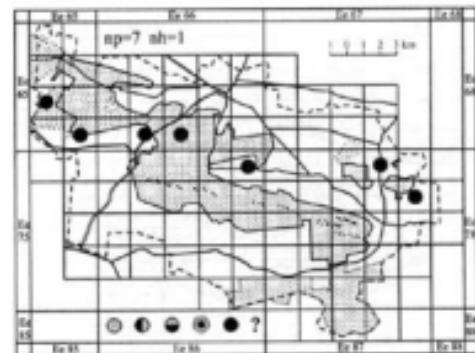
*Evernia prunastri* (L.) Ach.*Fellhaneropsis vezdae* (Coppins et P. James) Sérus. et Coppins*Flavoparmelia caperata* (L.) Hale*Fuscidea kochiana* (Hepp) V. Wirth & Věžda*Graphis scripta* (L.) Ach.*Gylecta truncigena* (Ach.) Hepp.



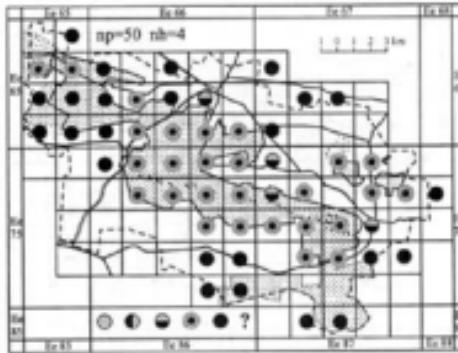
*Haematomma ochroleucum* (Neck.) J.R. Laundon  
var. *ochroleucum*



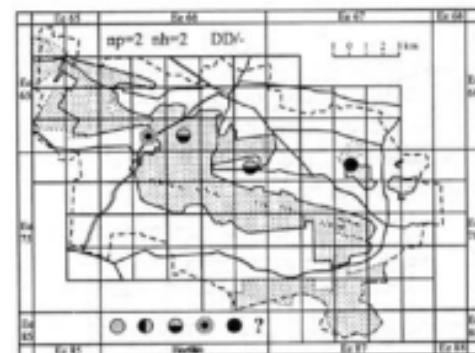
*Hypocenomyce antracophila* (Nyl.) P. James  
& Gotth. Schneid.



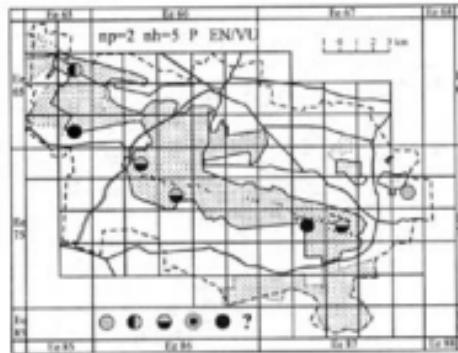
*Hypocenomyce caradocensis* (Leight. ex Nyl.)  
P. James & Gotth. Schneid.



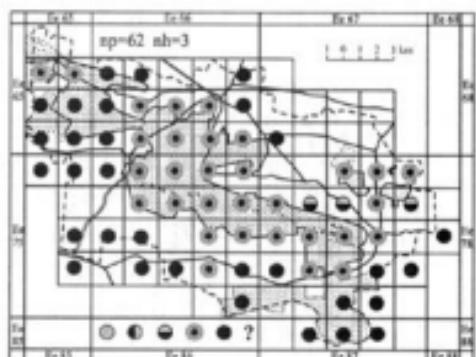
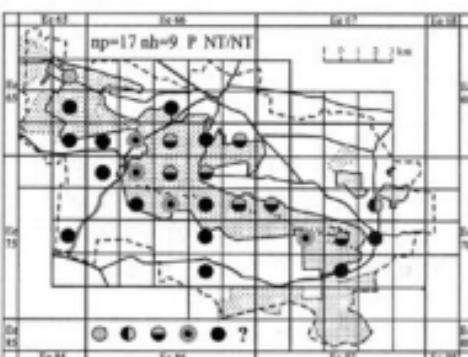
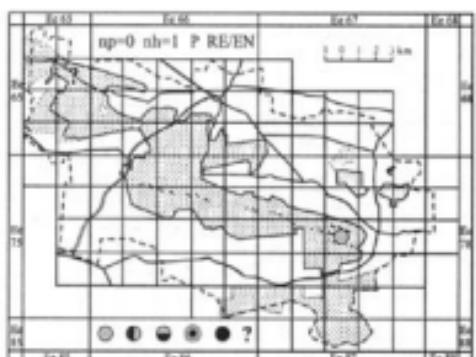
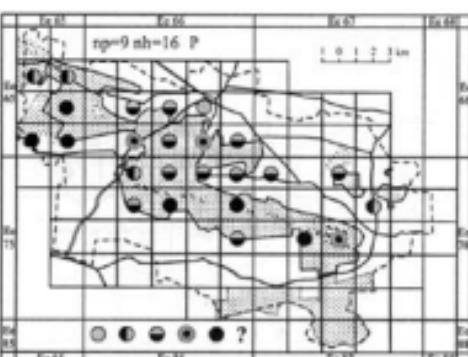
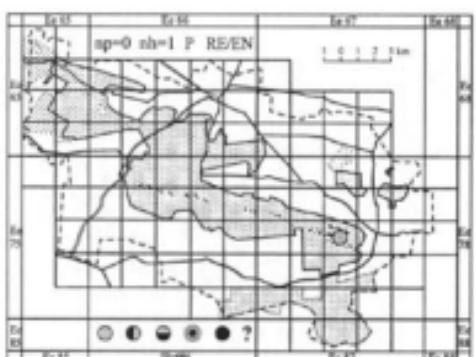
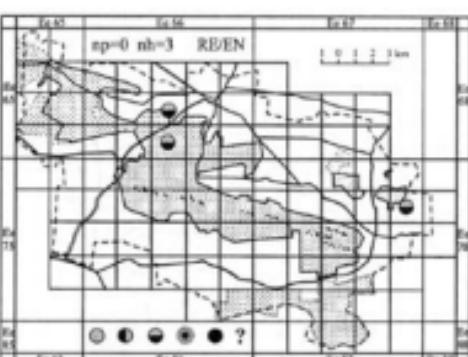
*Hypocenomyce scalaris* (Ach. ex Lilj.) M. Choisy

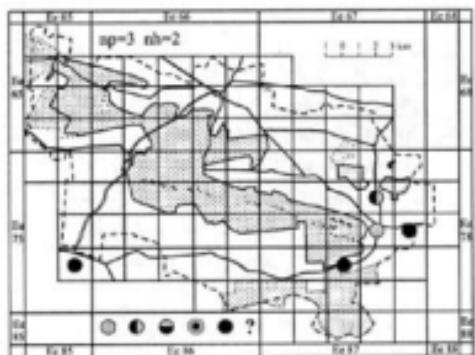
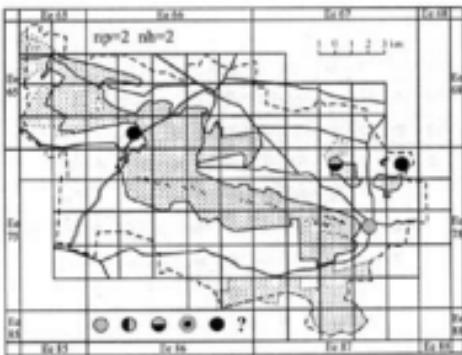
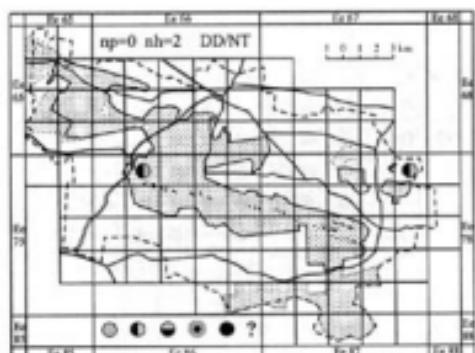
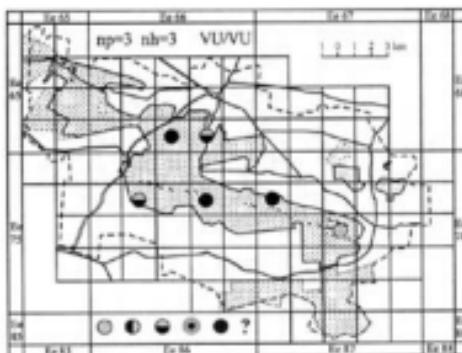
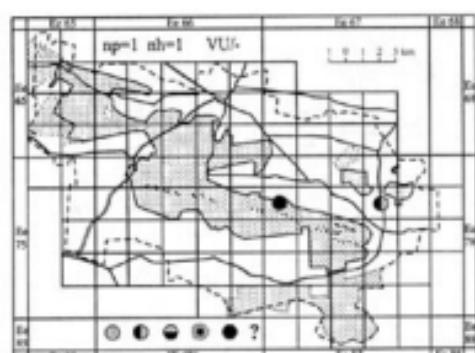
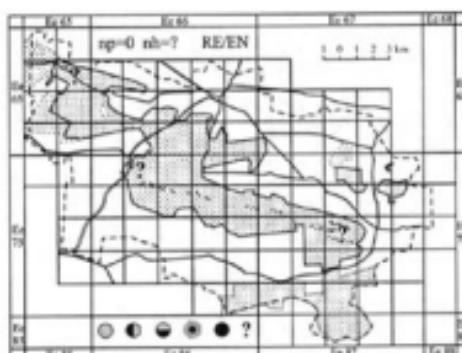


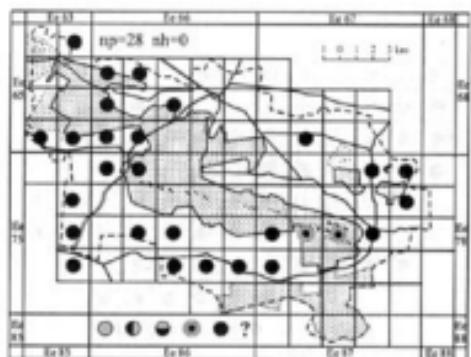
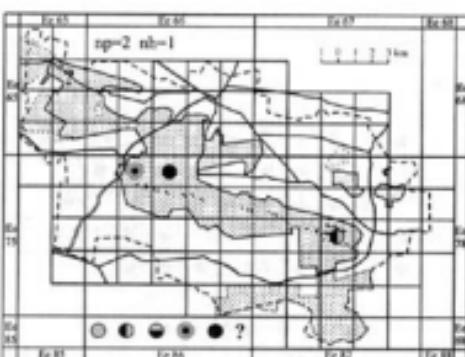
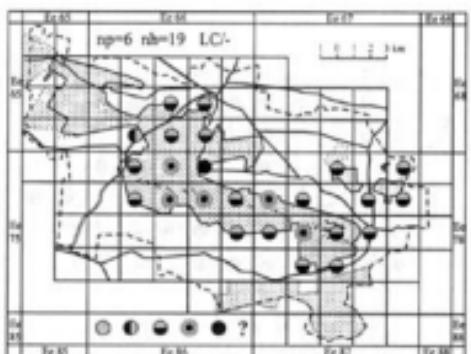
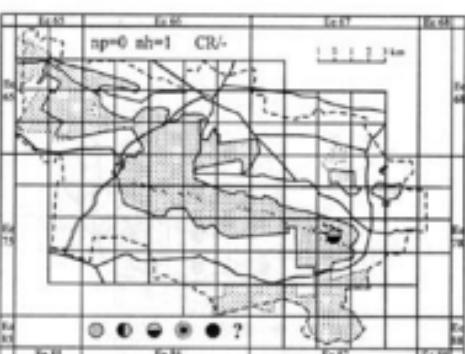
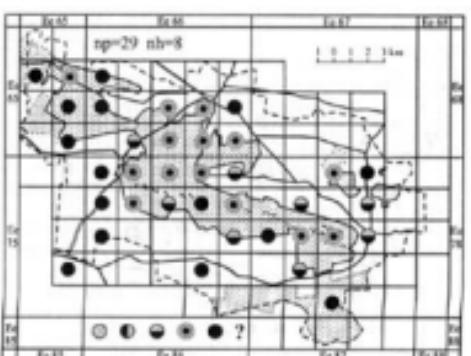
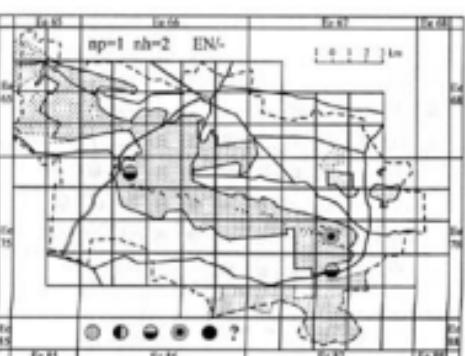
*Hypocenomyce sorophora* (Vain.) P. James  
& Poelt [*Pycnora sorophora* (Vain.) Hafellner]

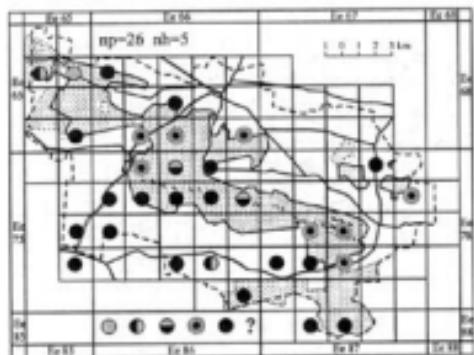
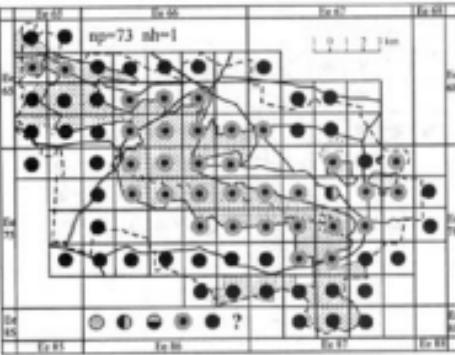
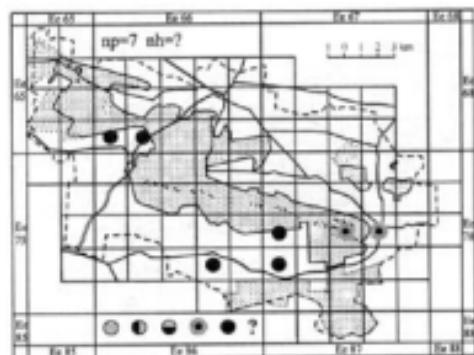
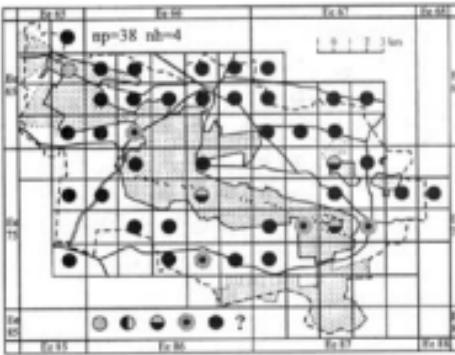
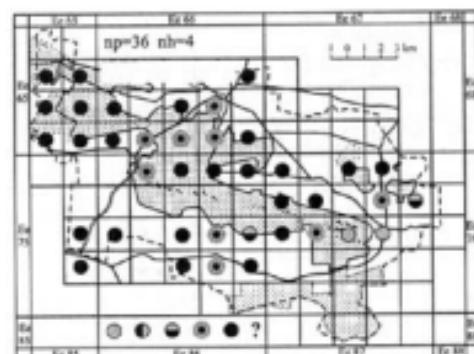
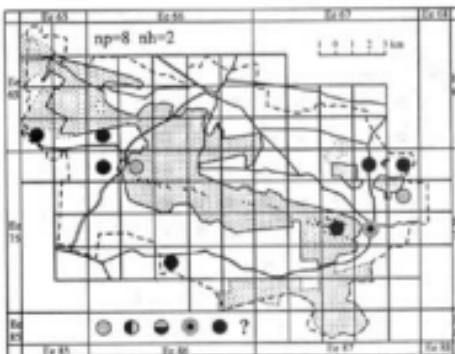


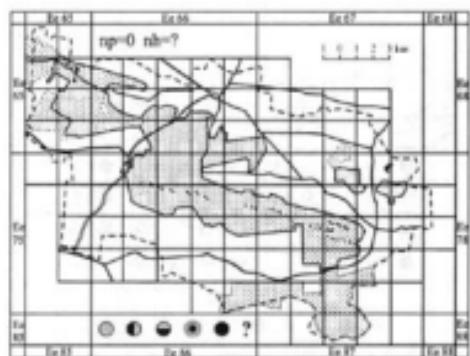
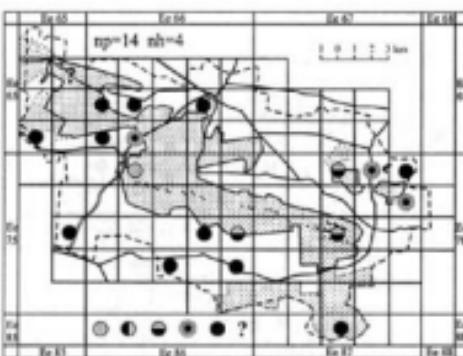
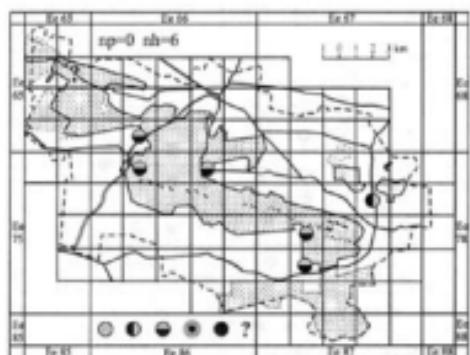
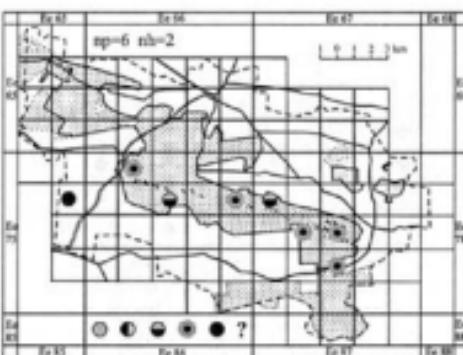
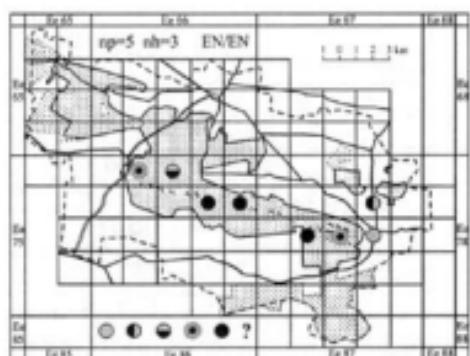
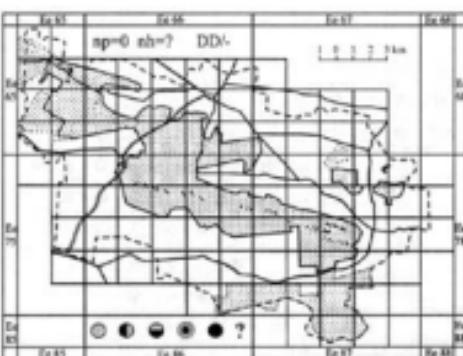
*Hypogymnia farinacea* Zopf

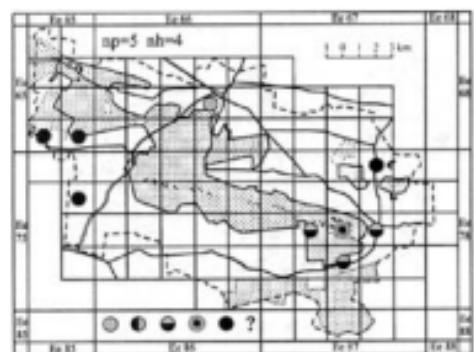
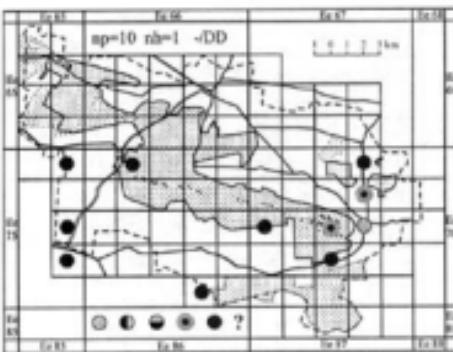
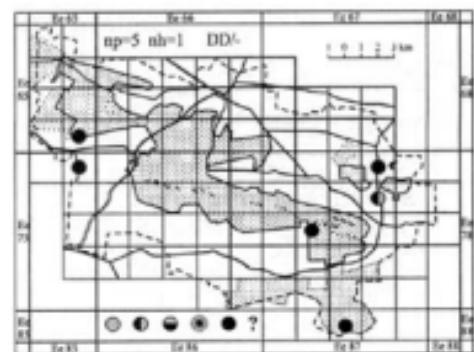
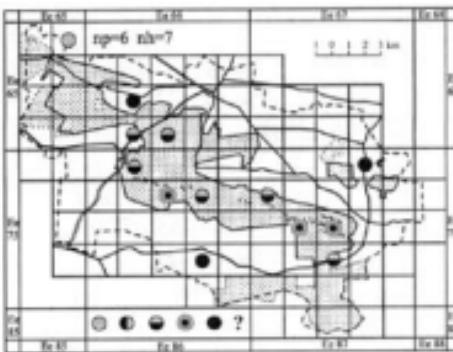
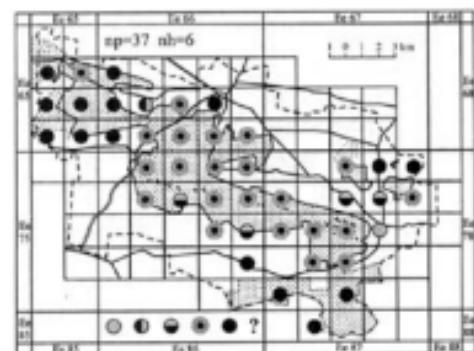
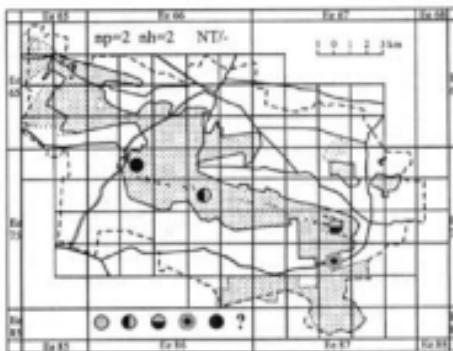
*Hypogymnia physodes* (L.) Nyl.*Hypogymnia tubulosa* (Schaer.) Hav.*Icmadophila ericetorum* (L.) A. Zahlbr.*Imshaugia aleurites* (Ach.) S.L.F. Mey.*Lasallia pustulata* (L.) Mérat*Lecanatis abietina* (Ach.) Körb.

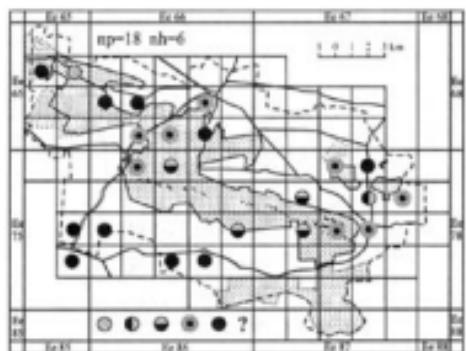
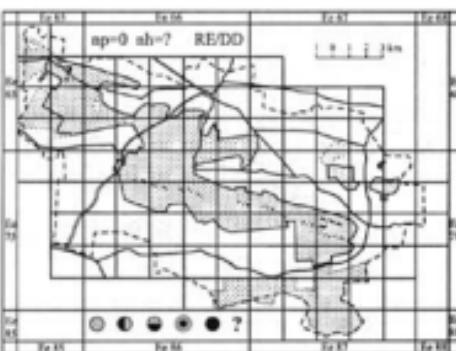
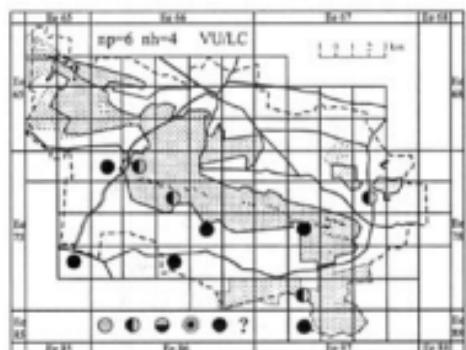
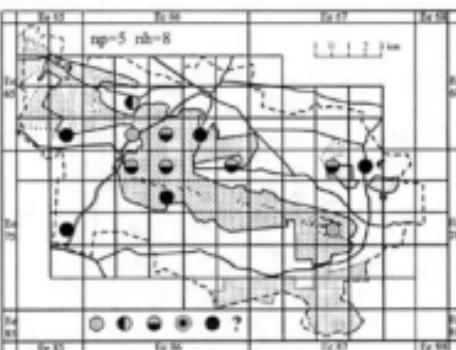
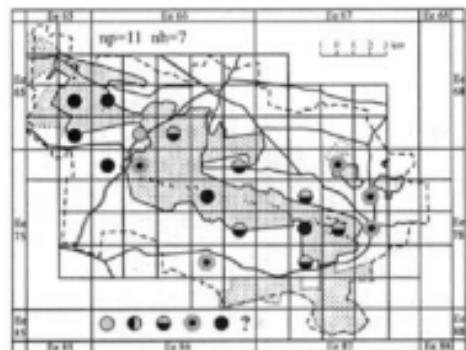
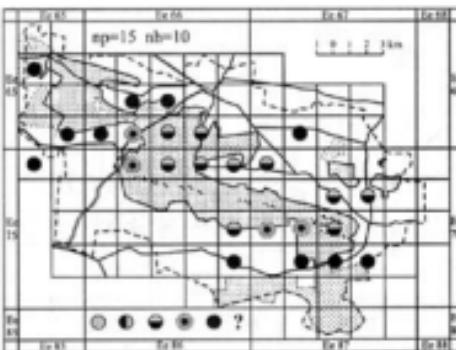
*Lecania cyrtella* (Ach.) Th. Fr.*Lecania erysibe* (Ach.) Mudd*Lecania fuscella* (Schaer.) A. Massal.*Lecania globulosa* (Flörke) P. Boom & Sérus.*Lecania naegelii* (Hepp) Diederich & P. Boom*Lecanora albella* (Pers.) Ach.

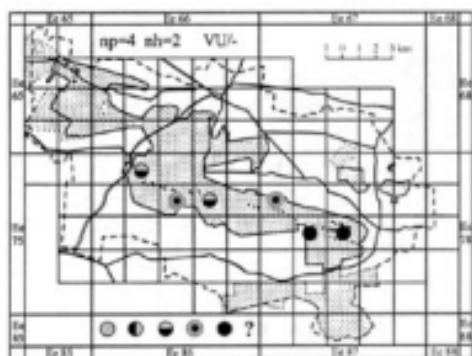
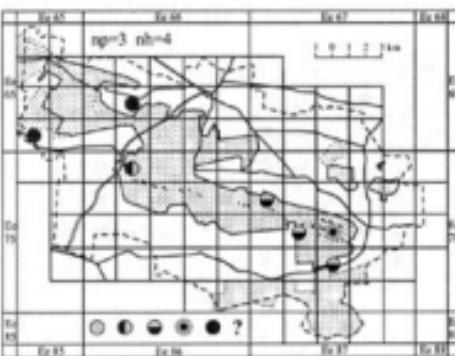
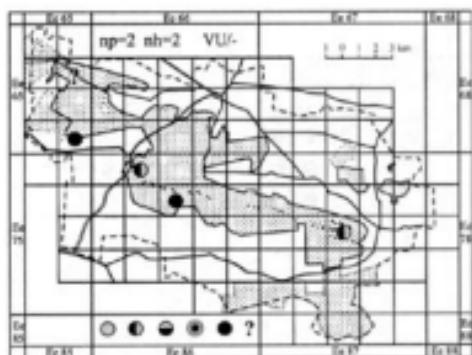
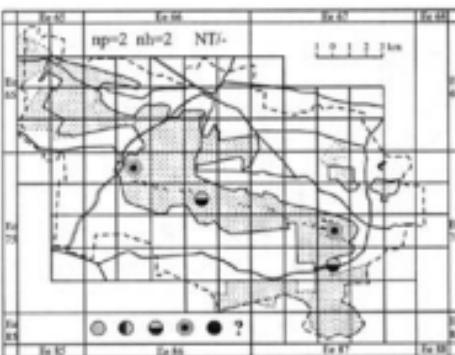
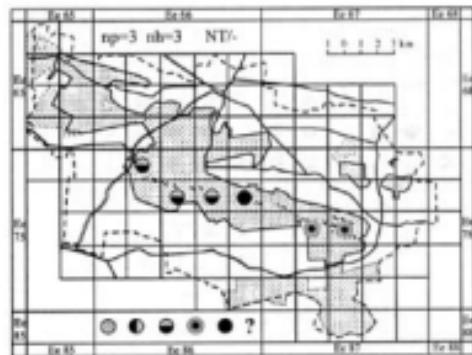
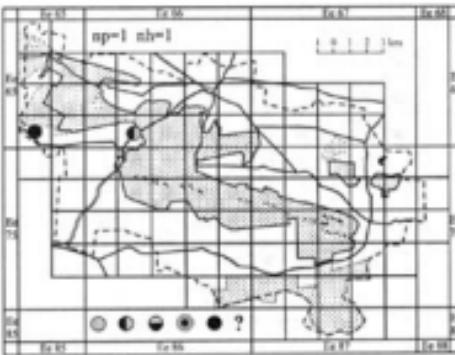
*Lecanora albescens* (Hoffm.) Branth & Rostr.*Lecanora allophana* Nyl.*Lecanora argentata* (Ach.) Malme*Lecanora campestris* (Schaer.) Hue*Lecanora carpinea* (L.) Vain.*Lecanora cenisia* Ach.

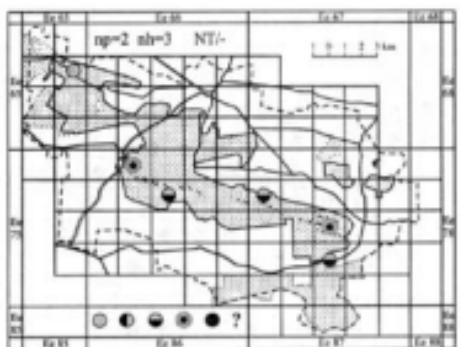
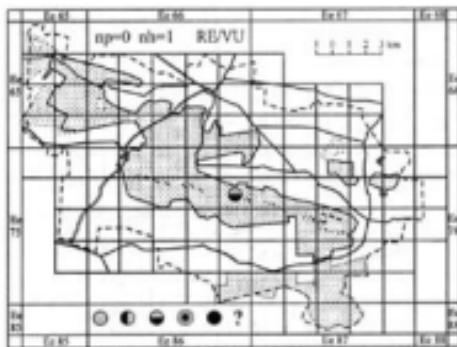
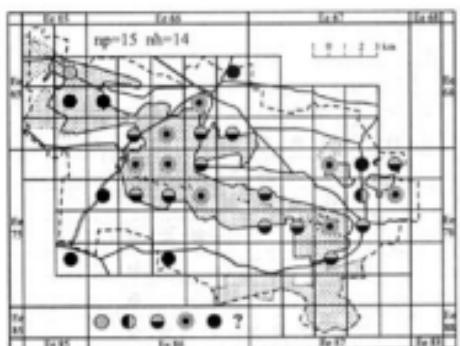
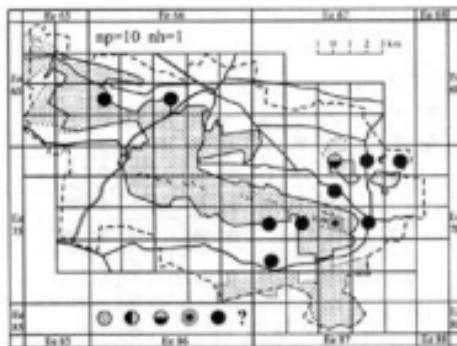
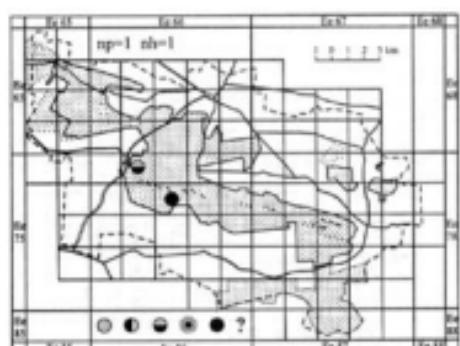
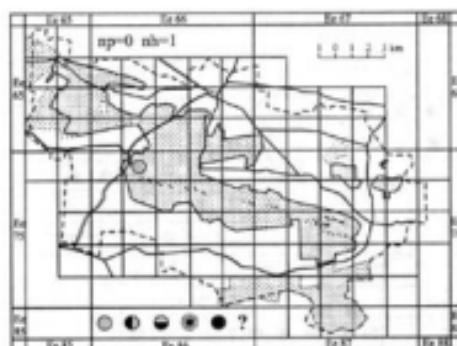
*Lecanora chlarotera* Nyl.*Lecanora conizaeoides* Nyl. ex Crombie*Lecanora crenulata* Hook.*Lecanora dispersa* (Pers.) Sommerf.*Lecanora expallens* Ach.*Lecanora flotowiana* Spreng.

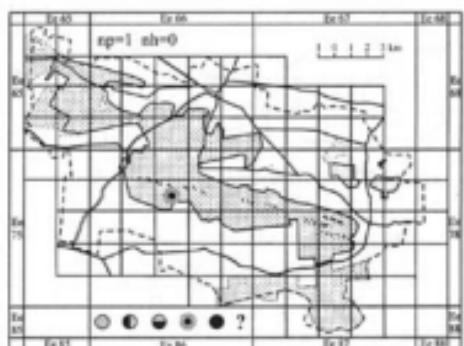
*Lecanora glabrata* (Ach.) Malme*Lecanora hagenii* (Ach.) Ach.*Lecanora impudens* Degel.*Lecanora intricata* (Ach.) Ach.*Lecanora intumescens* (Rebent.) Rebenn.*Lecanora leptyrodes* (Nyl.) Degel.

*Lecanora muralis* (Schreb.) Rebent.*Lecanora persimilis* (Th. Fr.) Nyl.*Lecanora piniperda* Körb.*Lecanora polytropa* (Ehrh. ex Hoffm.) Rabenh.*Lecanora pulicaris* (Pers.) Ach.*Lecanora rupicola* (L.) Zahlbr.

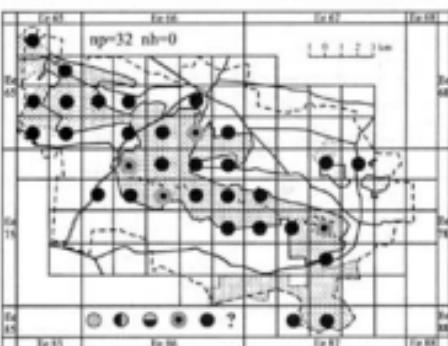
*Lecanora saligna* (Schrad.) Zahlbr.*Lecanora sambuci* (Pers.) Nyl.*Lecanora subrugosa* Nyl.*Lecanora symmicta* (Ach.) Ach.*Lecanora umbrina* (Ach.) A. Massal.*Lecanora varia* (Hoffm.) Ach.

*Lecidea confluens* (Weber) Ach.*Lecidea fuscoatra* (L.) Ach.*Lecidea lapticida* (Ach.) Ach. var. *lapticida**Lecidea lapticida* (Ach.) Ach. Var. *pantherina* Ach.*Lecidea lithophila* (Ach.) Ach.*Lecidea nylanderi* (Anzi) Th. Fr.

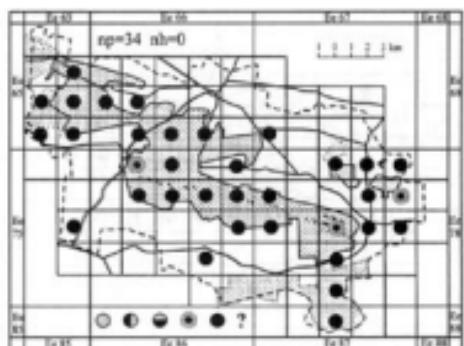
*Leucidea plana* (J. Lahm) Nyl.*Leucidea turgidula* Fr.*Lecidella elaeochroma* (Ach.) M. Choisy*Lecidella stigmatea* (Ach.) Hertel & Leuckert*Lepraria borealis* Lohtander & Tønsberg*Lepraria cacuminum* (A. Massal.) Kümmerling & Leuckert



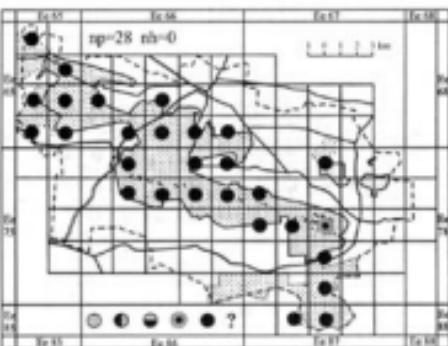
*Lepraria* cf. *eburnea* J.R. Laundon



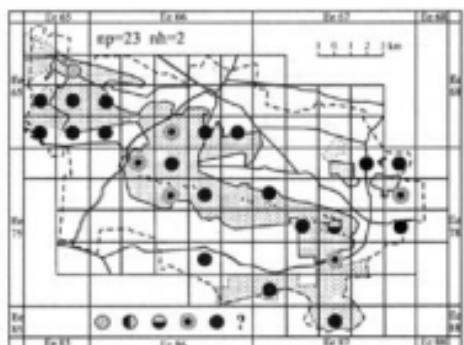
*Lepraria elobata* Tønsberg



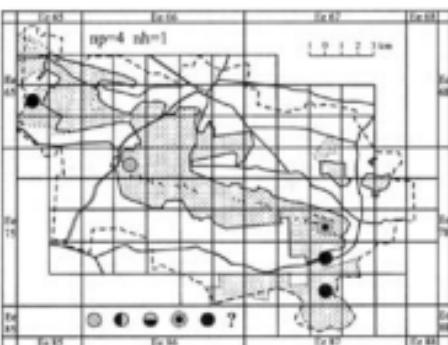
Lepraria incana (L.) Ach.



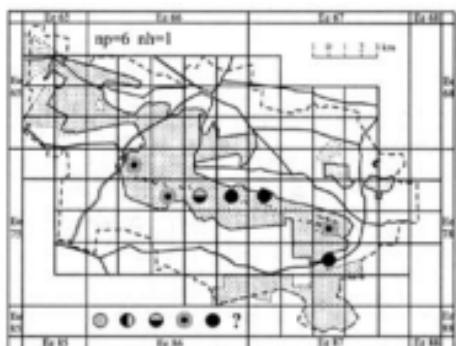
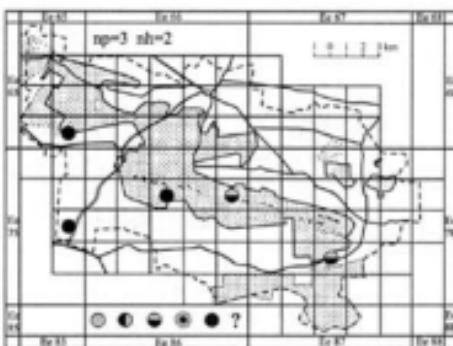
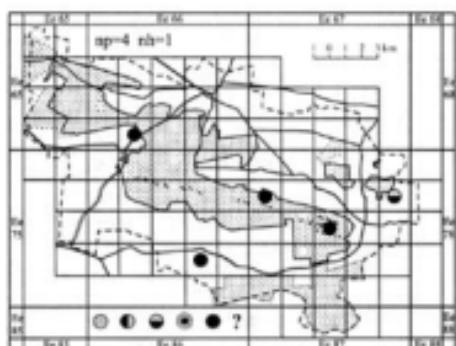
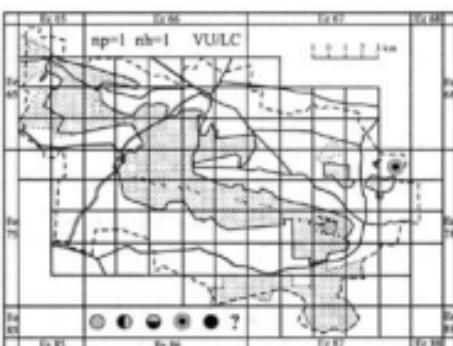
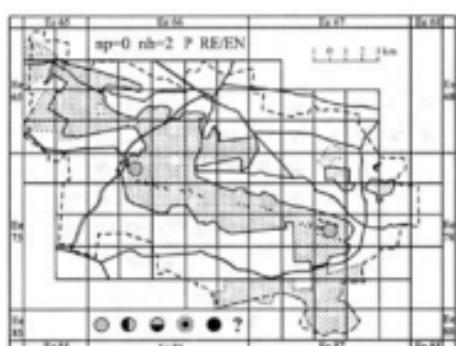
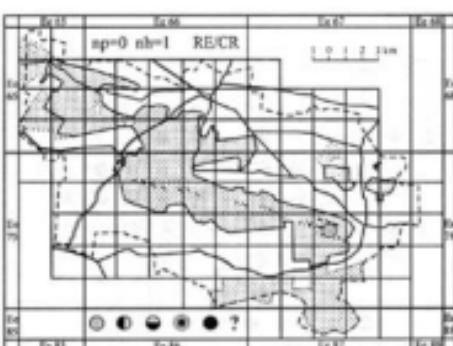
*Lepraria jackii* Tønsberg

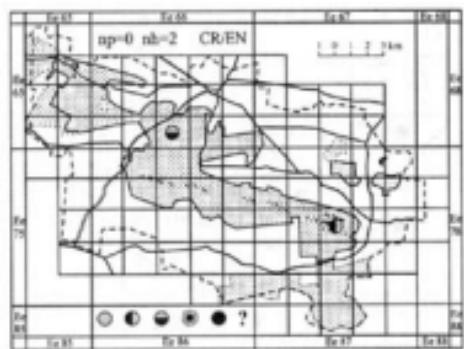


*Lepraria lobifrons* Nyl.

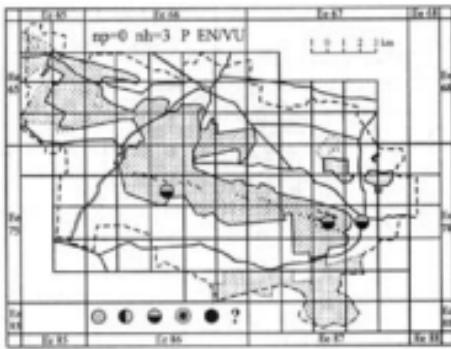


*Lepraria membranacea* (Dicks.) Vain.

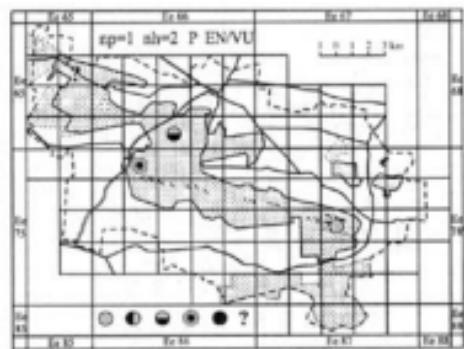
*Lepraria neglecta* (Nyl.) Lettau*Lepraria rigidula* (de Lesd.) Tønsberg*Lepraria vrouauxii* (Hue) R.C. Harris*Leptogium lichenoides* (L.) Zahlbr.*Lobaria pulmonaria* (L.) Hoffm.*Lopodium pezizoideum* (Ach.) Körb.



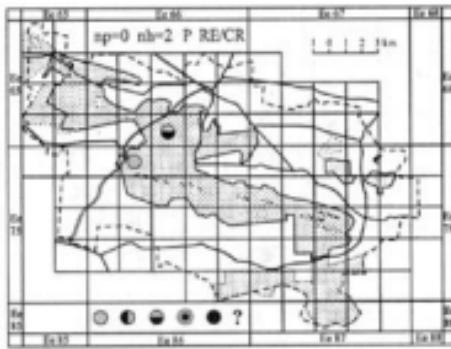
*Laxospora elatina* (Ach.) A. Massal.



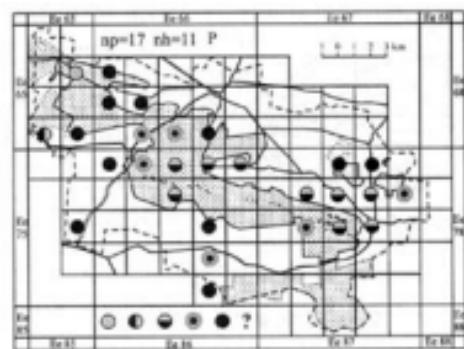
*Melanella disjuncta* (Erichsen) Essl.



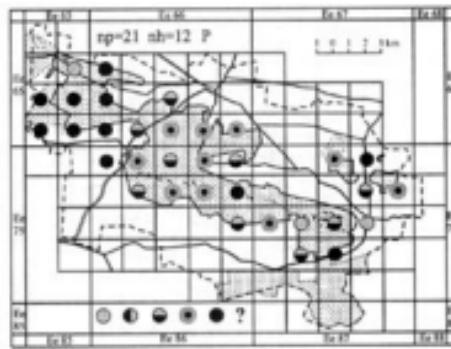
*Melanella elegans* (Zahlbr.) Essl.



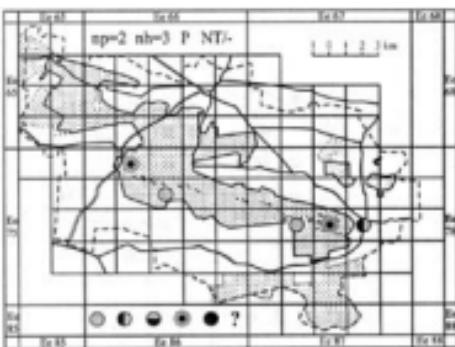
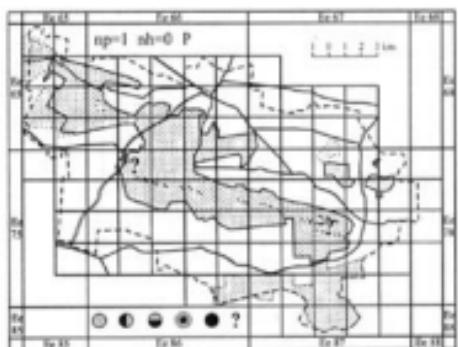
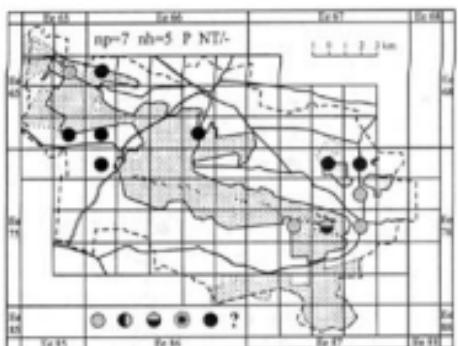
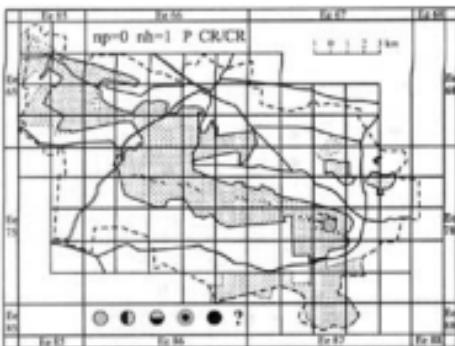
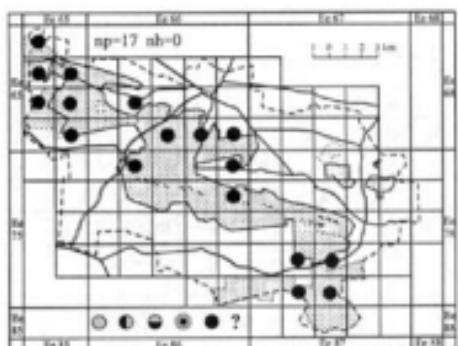
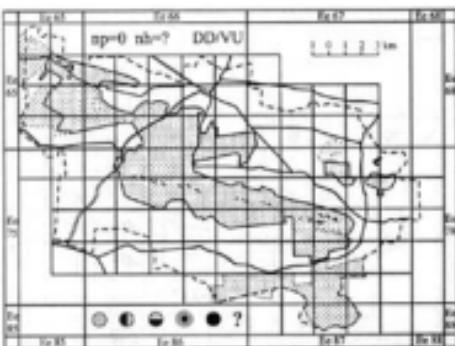
*Melanelia exasperata* (De Not.) Essl.

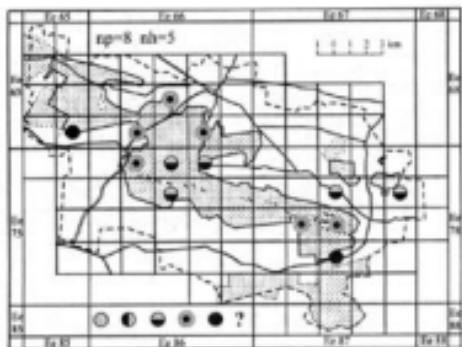
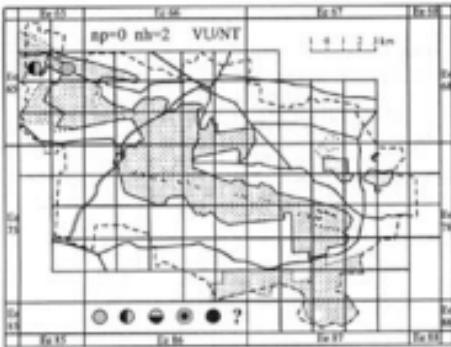
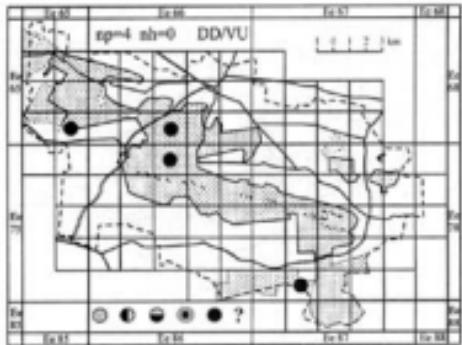
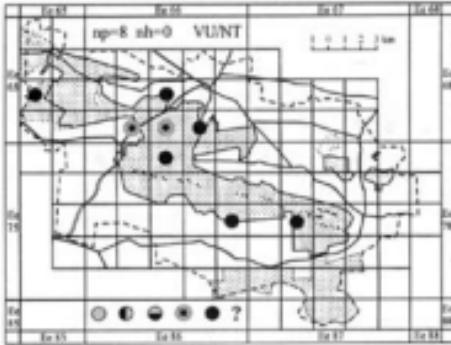
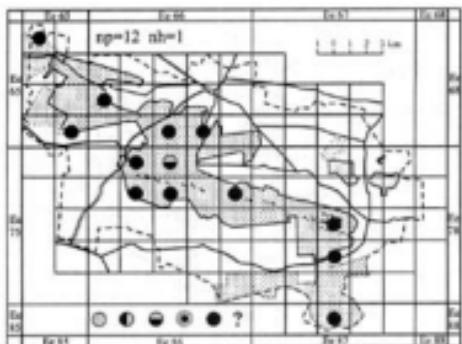
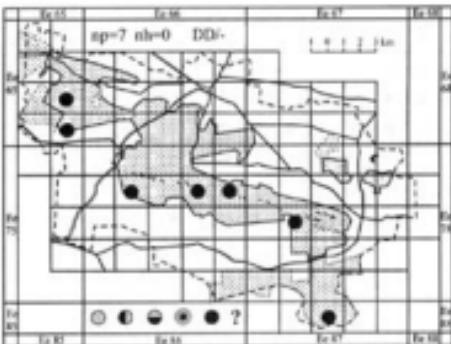


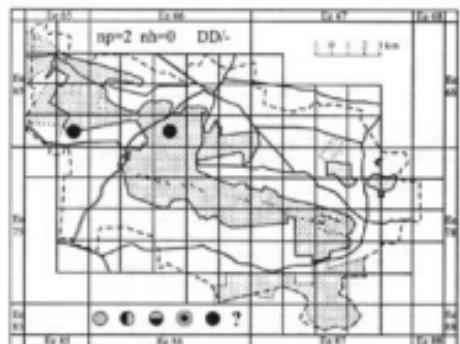
*Melanelia exasperatula* (Nyl.) Essl.



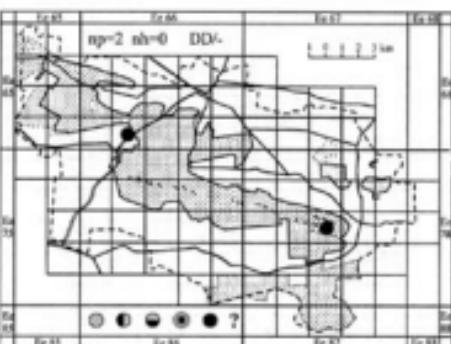
*Melanolia glabratula* (Lamy) Essl.

*Melanelia olivacea* (L.) Essl.*Melanelia stygia* (L.) Essl.*Melanelia subaurifera* (Nyl.) Essl.*Menegazzia terebrata* (Hoffm.) A. Massal.*Micarea botryoides* (Nyl.) Coppins*Micarea cinerea* (Schaer.) Hedi.

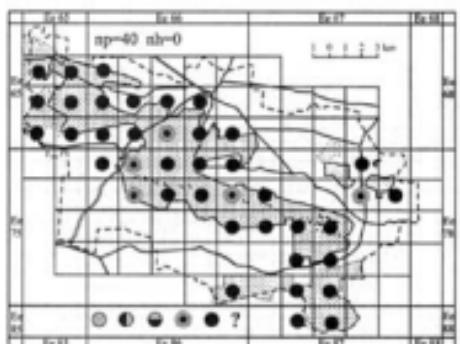
*Micarea denigrata* (Fr.) Hedl.*Micarea erraticata* (Körb.) Hertel, Rambold & Pietschm.*Micarea hendlundii* Coppins*Micarea melaena* (Nyl.) Hedl.*Micarea misella* (Nyl.) Hedl.*Micarea nigella* Coppins



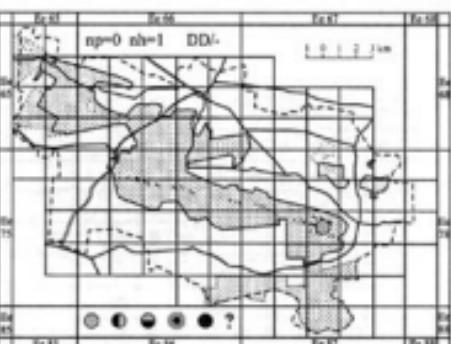
*Micarea nitschkeana* (J. Lahm ex Rabenh.) Harm.



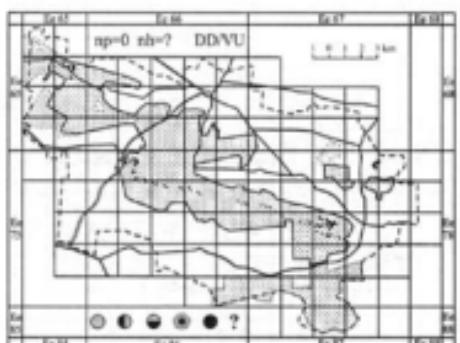
*Micarea peliocarpa* (Anzi) Coppins et R. Sant.



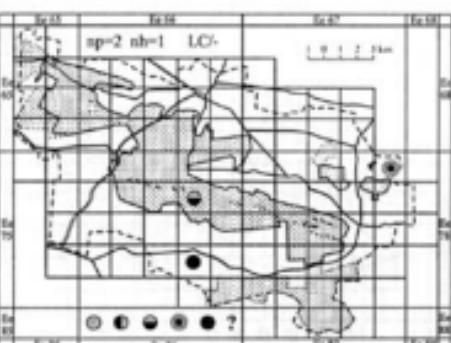
*Micarea prasina* Fr. s.l.



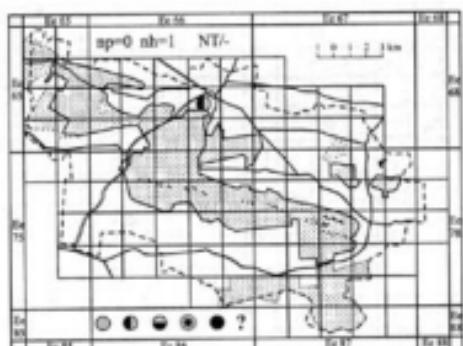
*Micarea sylvicola* (Flot.) Vézda & V. Wirth



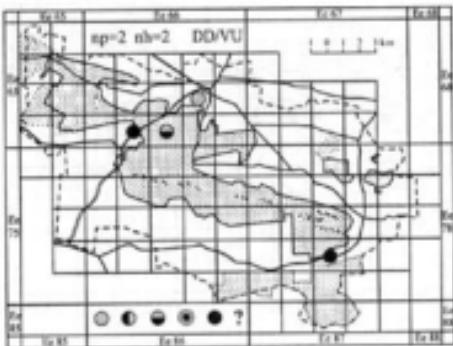
*Miriquidica leucophaea* (Flörke ex Rabenh.) Hertel & Rambold



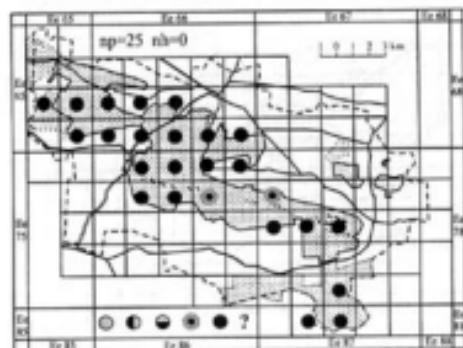
*Mycobilimbia fusca* (A. Massal.) Hafellner & V. Wirth



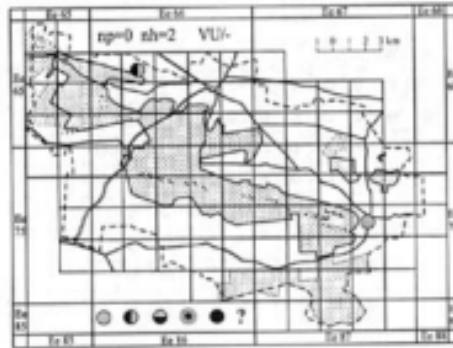
*Mycobilimbia sabuletorum* (Schreb.) Hafellner



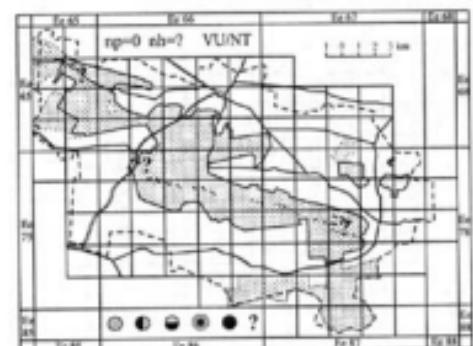
*Mycobilimbia sphaeroides* (J. Dicks.)



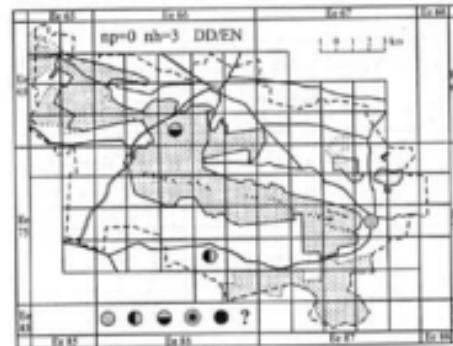
*Mycblastus fucatus* (Stirt.) Zahlbr.



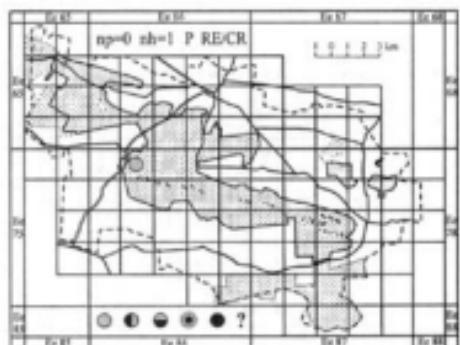
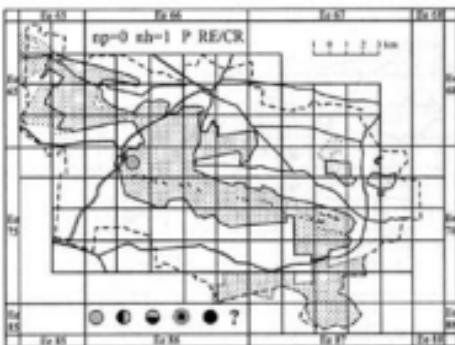
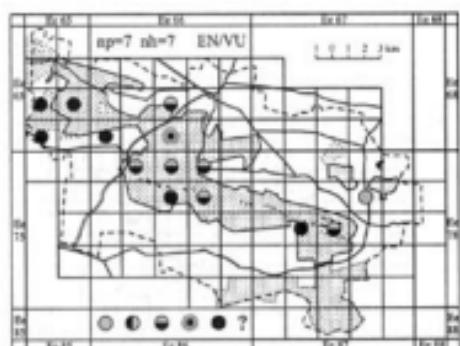
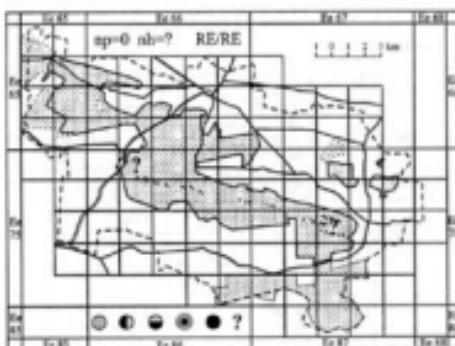
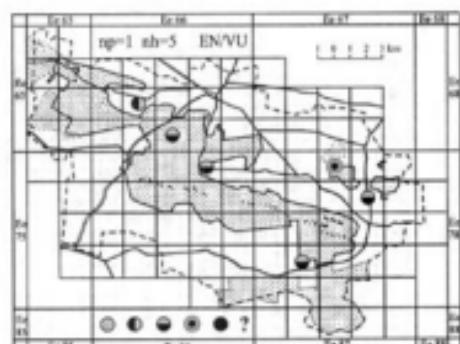
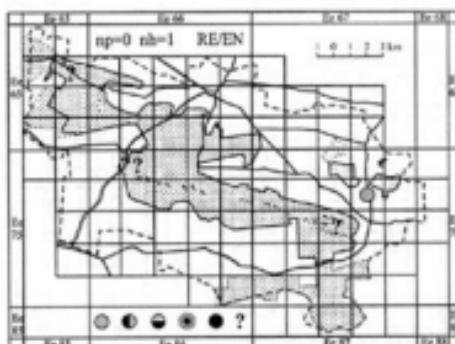
*Neofuscelia loxodes* (Nyl.) Essl.

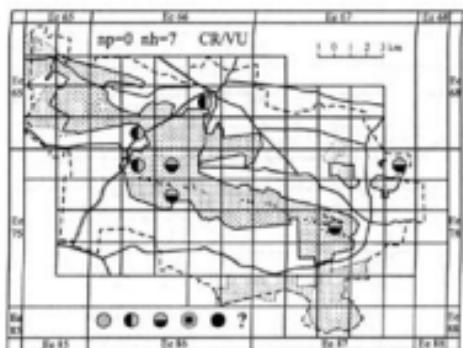
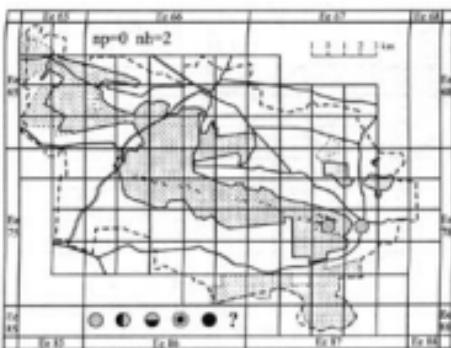
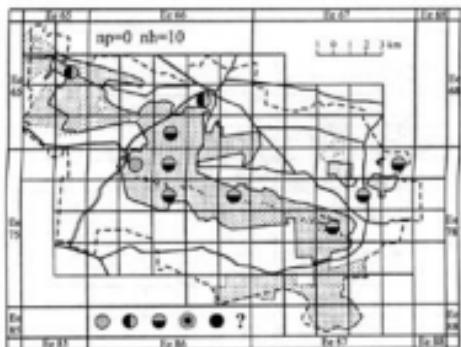
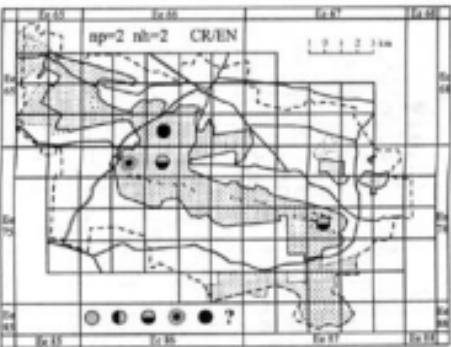
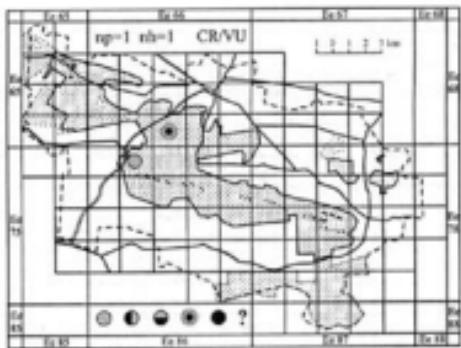
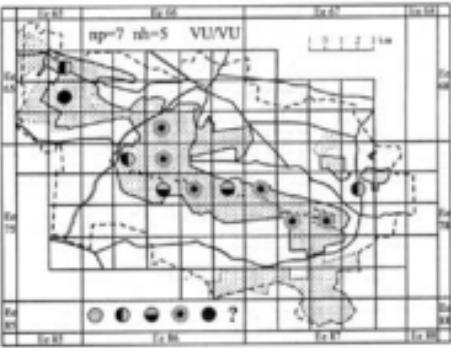


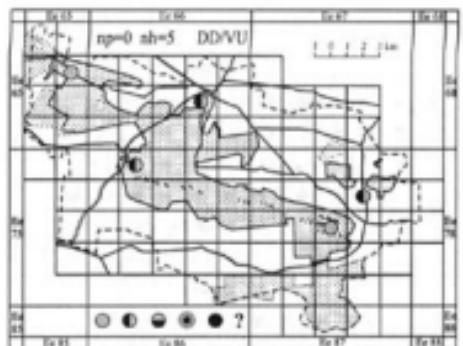
*Neofuscelia pulla* (Ach.) Essl.



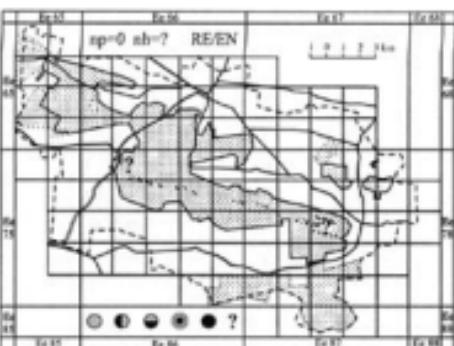
*Neofuscelia verruculifera* (Nyl.) Essl

*Nephroma laevigatum* Ach.*Nephroma resupinatum* (L.) Ach.*Ochrolechia androgyna* (Hoffm.) Arnold*Ochrolechia parella* (L.) A. Massal.*Ochrolechia subviridis* (Hoeg) Erichsen*Opegrapha atra* Pers.

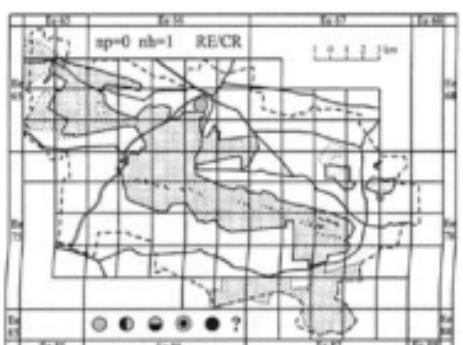
*Opegrapha rufescens* Pers.*Opegrapha rupestris* Pers.*Opegrapha varia* Pers. var. *varia**Opegrapha vermicellifera* (Kunze) J.R. Laundon*Opegrapha viridis* (Pers. ex Ach.) Behlen & Desberger*Opegrapha vulgata* Ach. var. *subsiderella* Nyl.



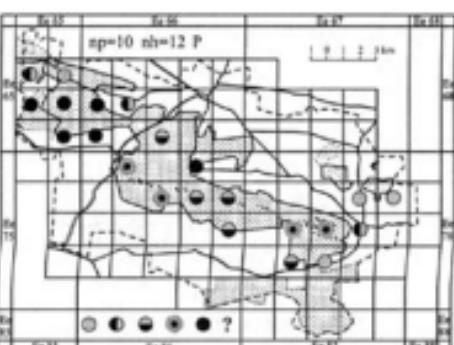
*Opegrapha vulgata* Ach. var. *vulgata*



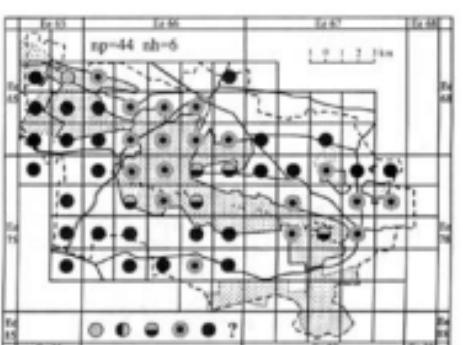
*Orpiniospora mosigii* (Körb.) Hertel & Rambold



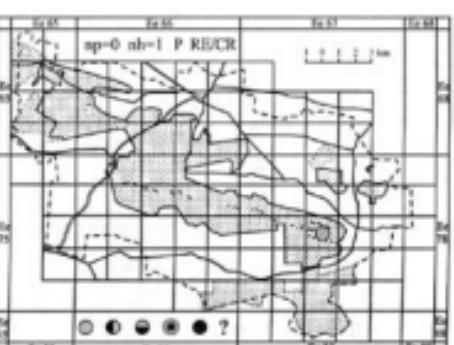
*Pannaria pezizoides* (Weber) Trevisan



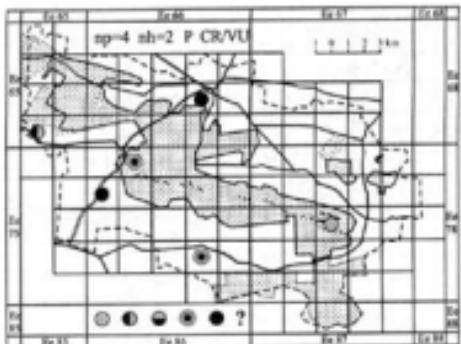
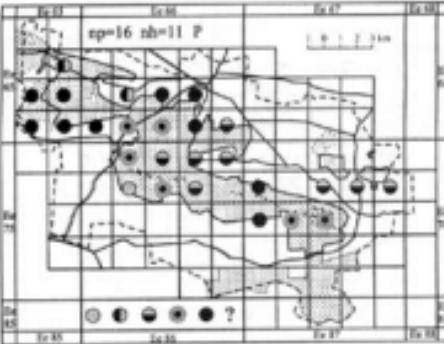
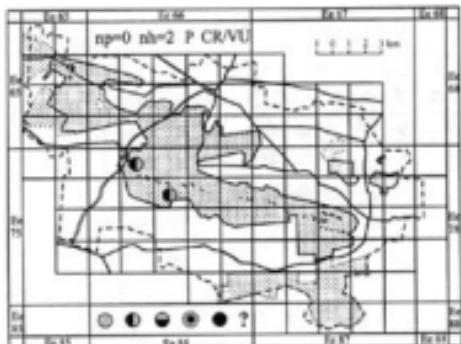
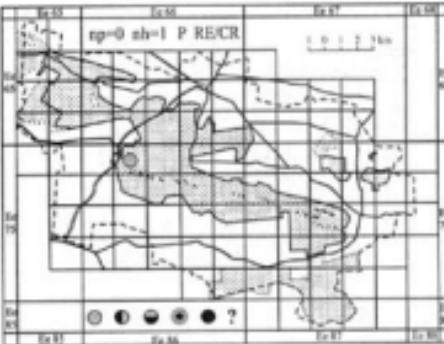
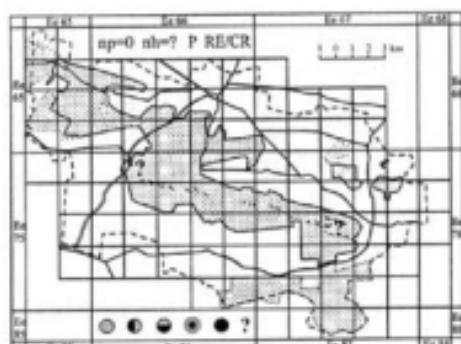
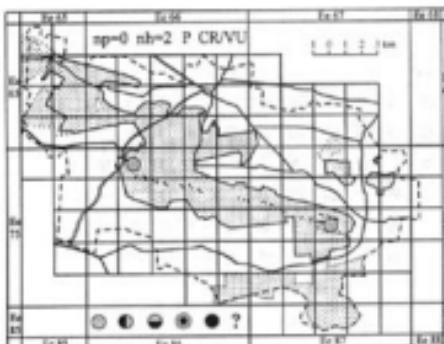
*Parmelia saxatilis* (L.) Ach.

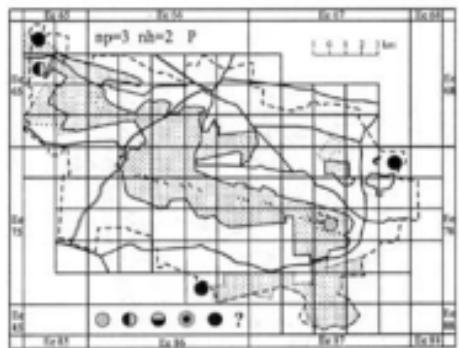


#### *Parmelia sulcata* Tuck.

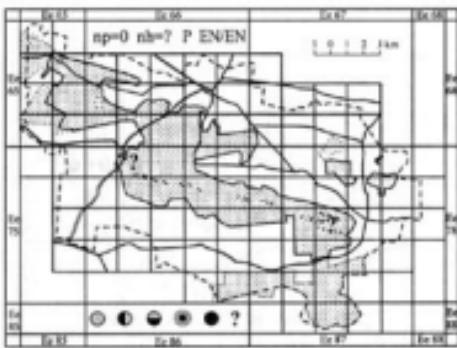


#### *Parmelina austriaca* (Savill) Hale

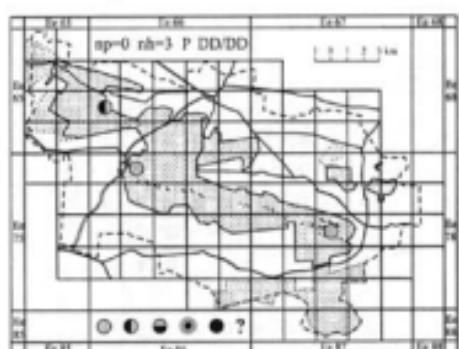
*Parmelina tiliacea* (Hoffm.) Hale*Parmeliopsis ambigua* (Wulfen) Nyl.*Parmeliopsis hyperopta* (Ach.) Arnold*Parmotrema chinense* (Osbeck) Hale & Ahti*Peltigera aphthosa* (L.) Willd.*Peltigera canina* (L.) Willd.



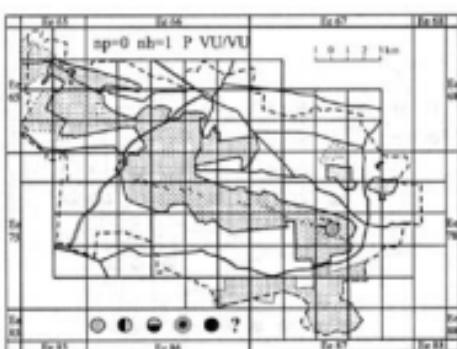
*Peltigera didactyla* (With.) J.R. Laundon  
var. *didactyla*



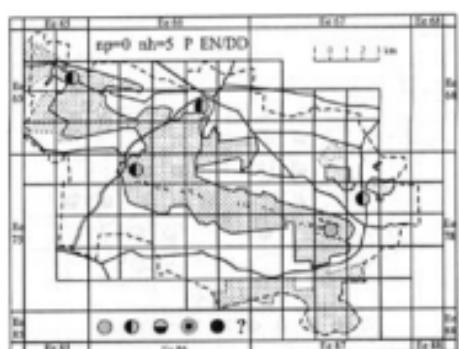
*Peltigera horizontalis* (Huds.) Baumg.



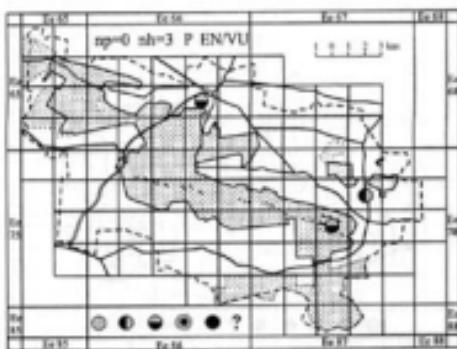
*Peltigera hymenina* (Ach.) Delise



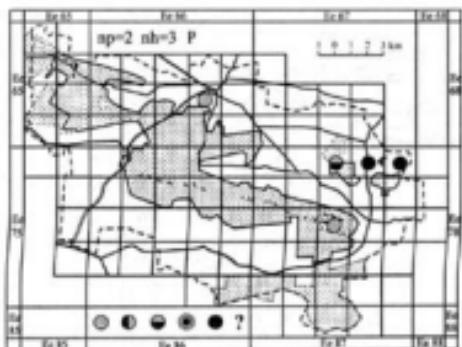
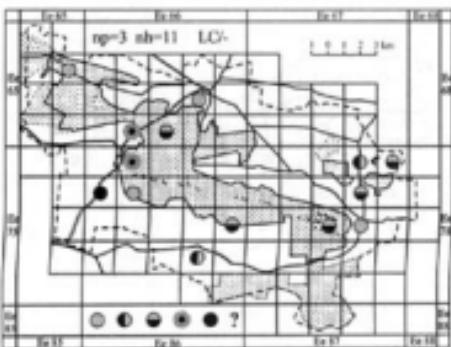
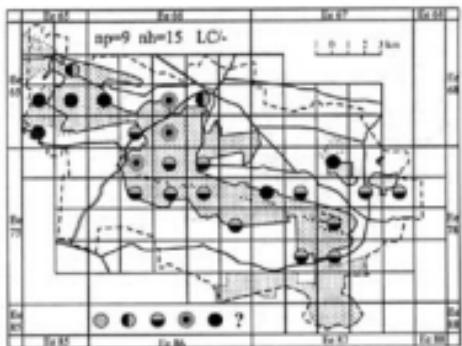
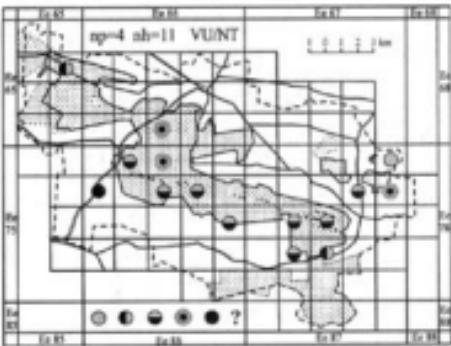
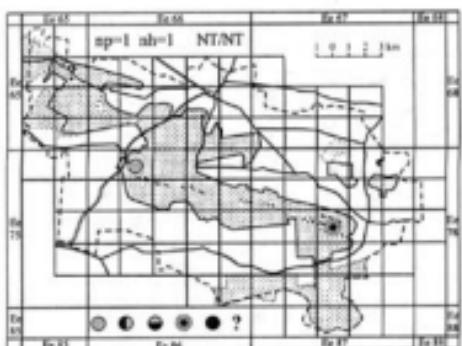
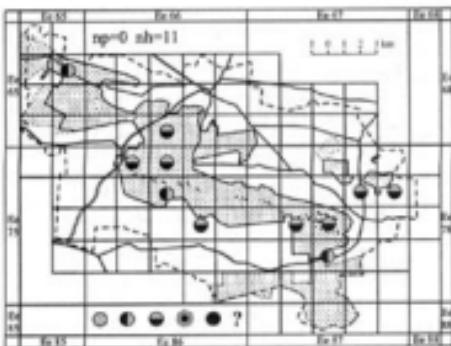
*Peltigera malacea* (Ach.) Funck

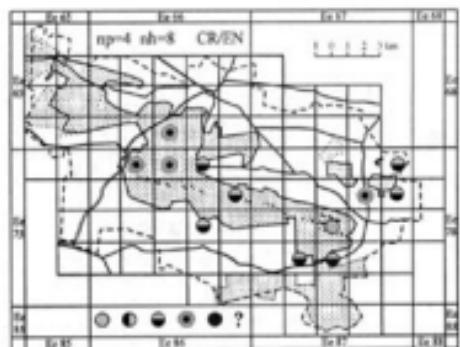
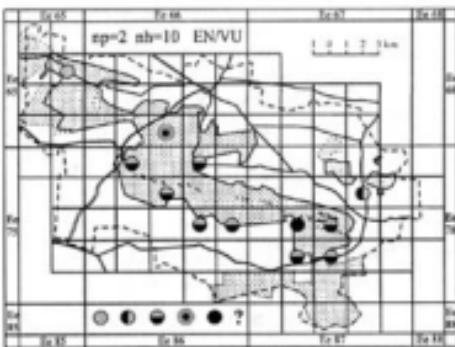
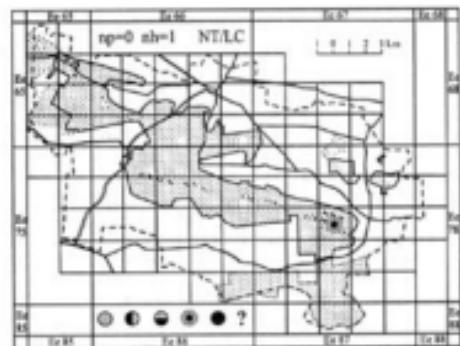
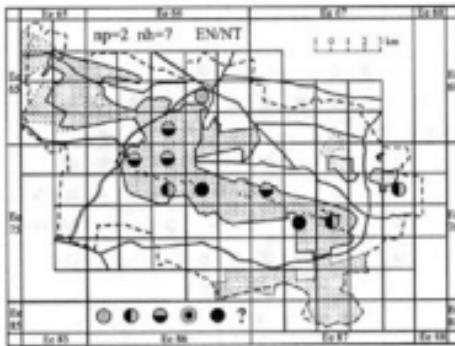
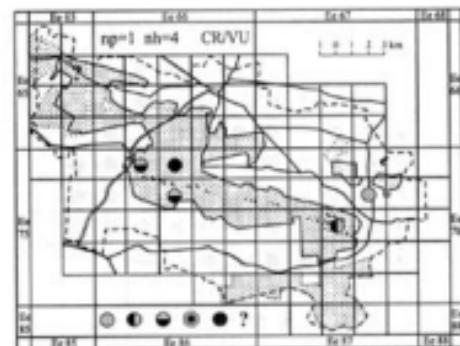
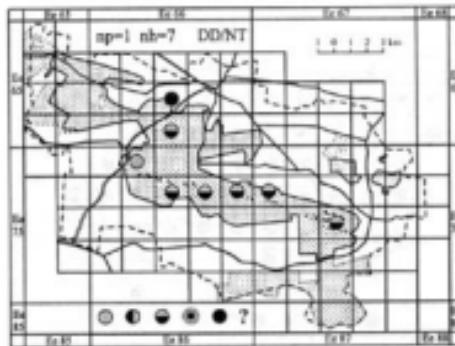


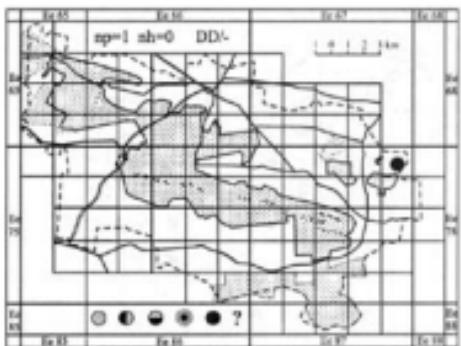
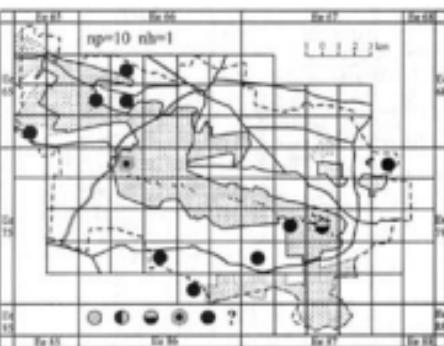
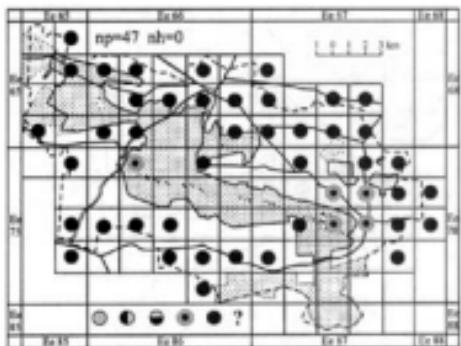
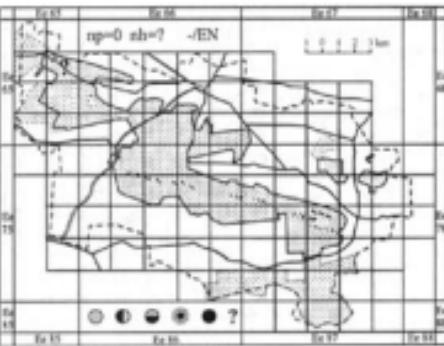
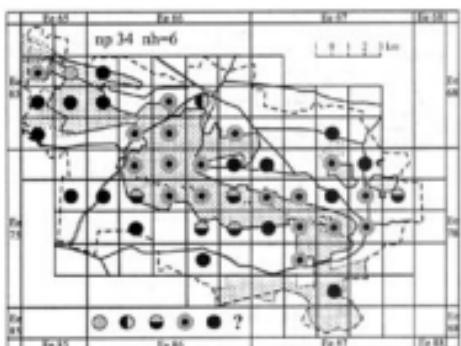
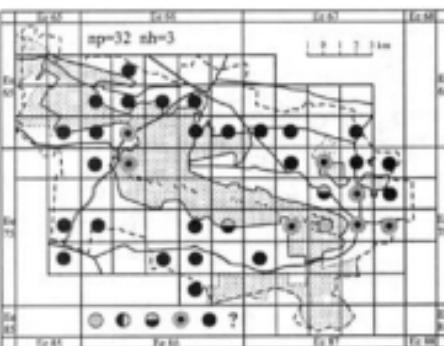
*Peltigera polydactylon* (Neck.) Hoffm.

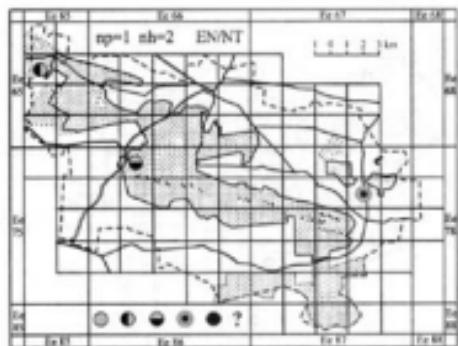
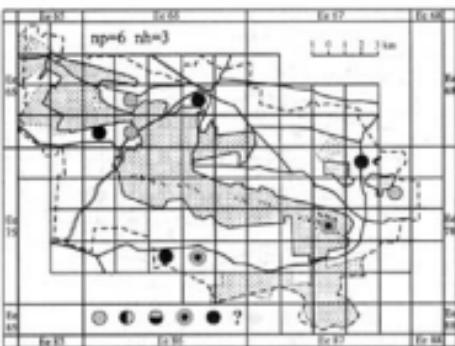
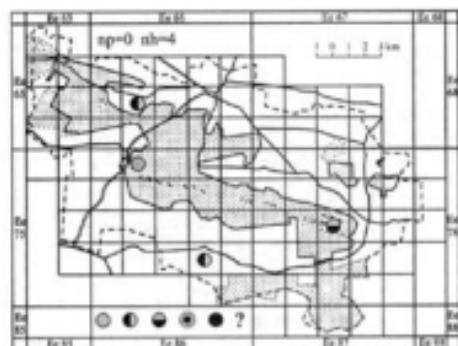
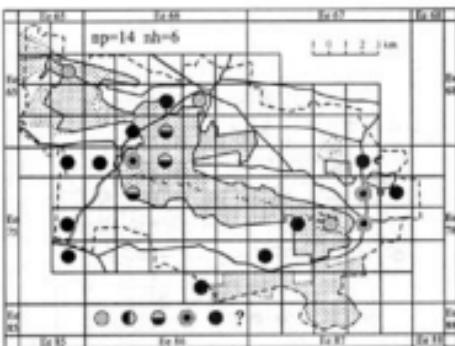
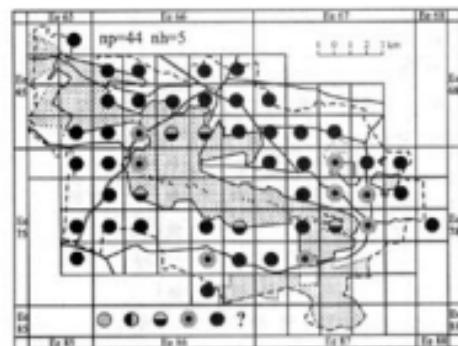
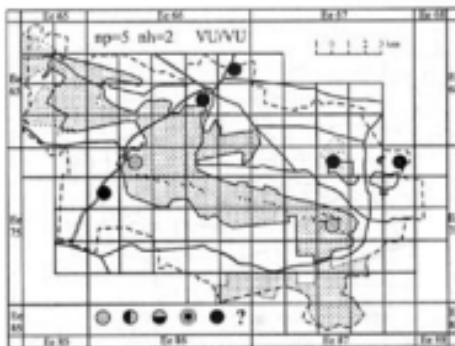


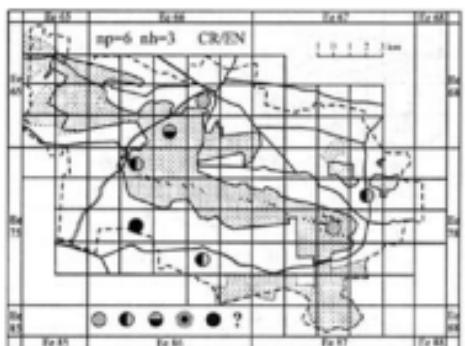
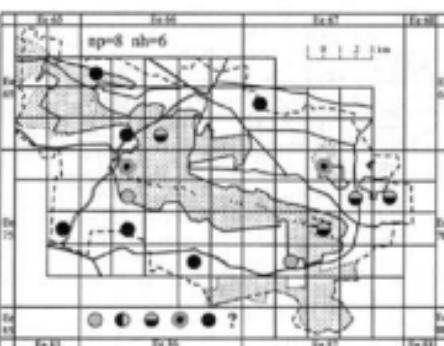
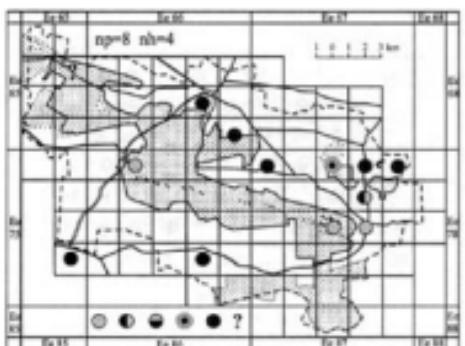
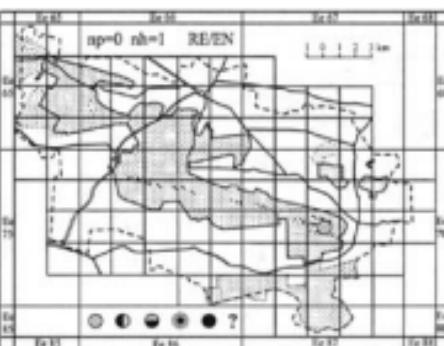
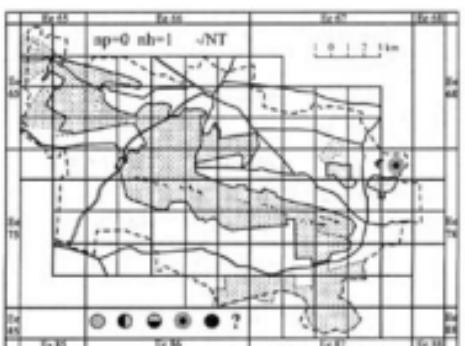
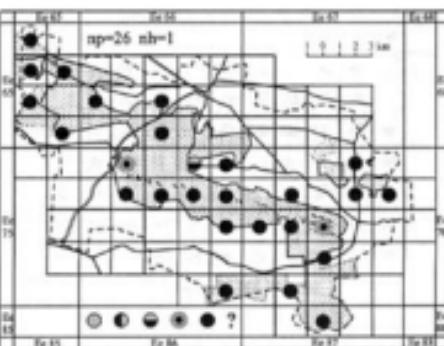
*Peltigera praetextata* (Flürke ex Sommerf.) Zopf

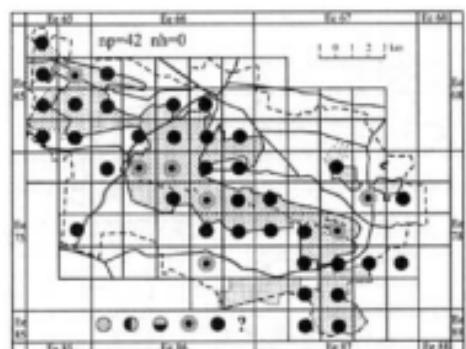
*Peltigera rufescens* (Weiss) Humb.*Pertusaria albescens* (Huds.) M. Choisy & Werner*Pertusaria amara* (Ach.) Nyl.*Pertusaria coccodes* (Ach.) Nyl.*Pertusaria corallina* (L.) Arnold*Pertusaria coronata* (Ach.) Th. Fr.

*Pertusaria flavidula* (DC.) J.R. Laundon*Pertusaria hemisphaerica* (Flörke) Erichsen*Pertusaria lactea* (L.) Arnold*Pertusaria leioplaca* DC.*Pertusaria pertusa* (Weigel) Tuck.*Pertusaria pupillaris* (Nyl.) Th. Fr.

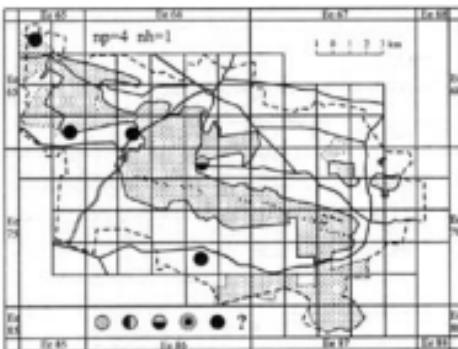
*Phaeophyscia kairamoi* (Vain.) Moberg*Phaeophyscia nigricans* (Flörke) Moberg*Phaeophyscia orbicularis* (Neck.) Moberg*Phlyctis agelaea* (Ach.) Flot.*Phlyctis argena* (Spreng.) Flot.*Physcia adscendens* (Fr.) H. Olivier

*Physcia aipolia* (Ehrh. ex Humb.) Fürnr.*Physcia caesia* (Hoffm.) Fürnr.*Physcia dubia* (Hoffm.) Lettau*Physcia stellaris* (L.) Nyl.*Physcia tenella* (Scop.) DC.*Physconia detersa* (Nyl.) Poelt

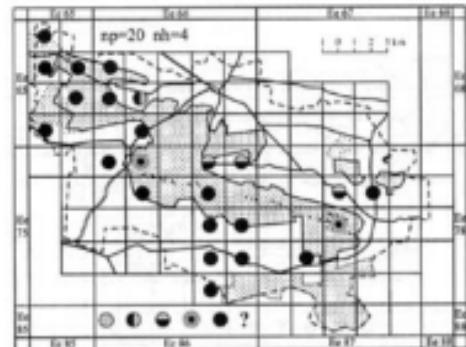
*Physconia distorta* (With.) J.R. Laundon*Physconia enteroxantha* (Nyl.) Poelt*Physconia grisea* (Lam.) Poelt*Placidium rufescens* (Ach.) A. Massal.*Placidium squamulosum* (Ach.) Breuss*Placynthiella dasaea* (Stirt.) Tønsberg



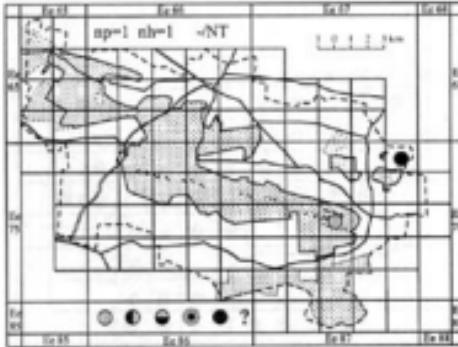
*Placynthiella icmalea* (Ach.) Coppins & P. James



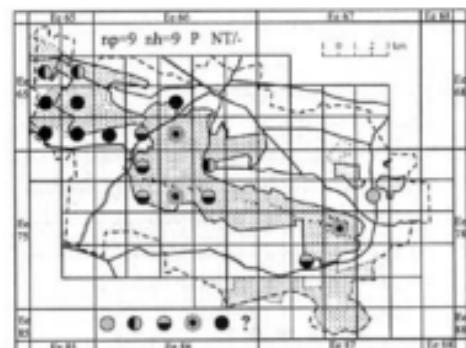
*Placynthiella oligotropha* (J.R. Laundon)  
Coppins & P. James



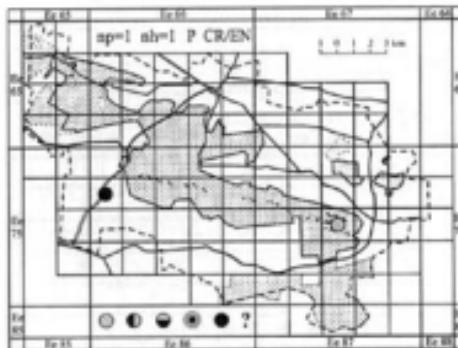
*Placynthiella uliginosa* (Schrad.) Coppins  
& P. James



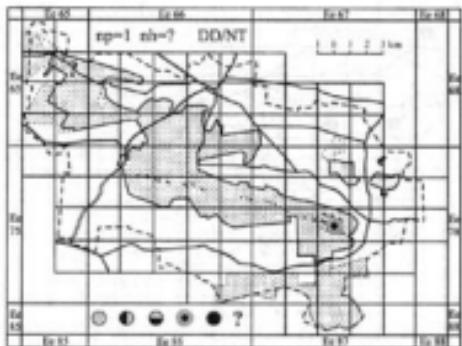
*Placynthium nigrum* (Huds.) S. Gray



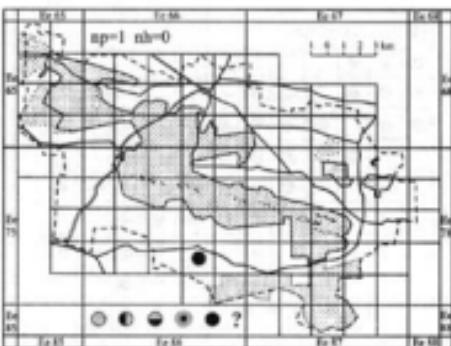
*Platismatia glauca* (L.) W.L. Culb. & C.F. Culb.



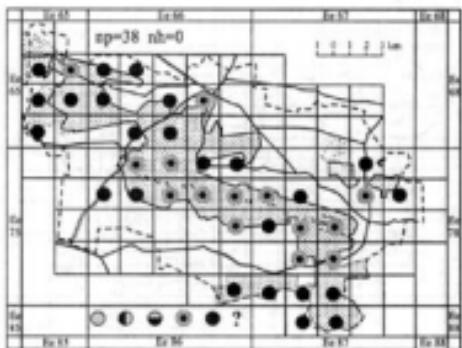
*Pleurostictia acetabulum* (Neck.) Elix  
& Lumbsch



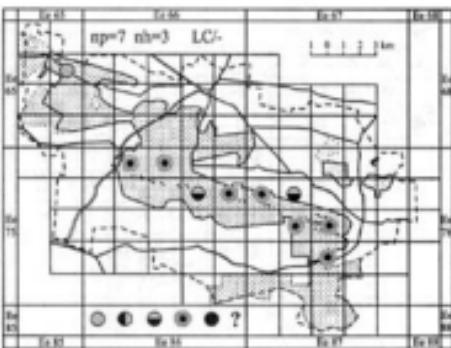
*Polysporina lapponica* (Ach. ex Schaeer.) Degel.



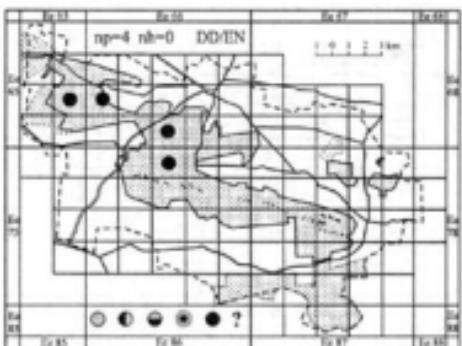
*Polysporina simplex* (Dav.) Vězda



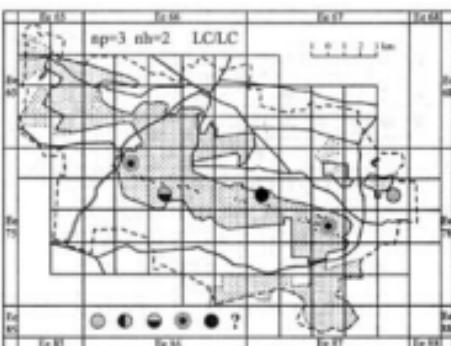
Porina aenea (Wallr.) Zahlbr.



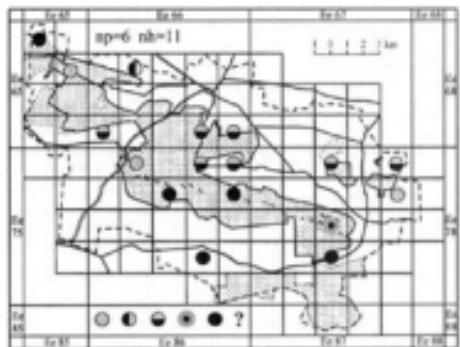
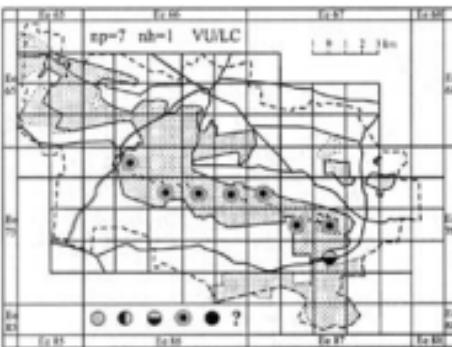
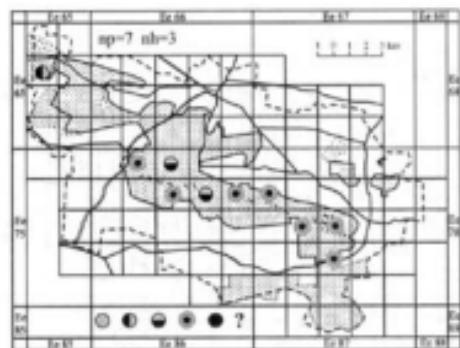
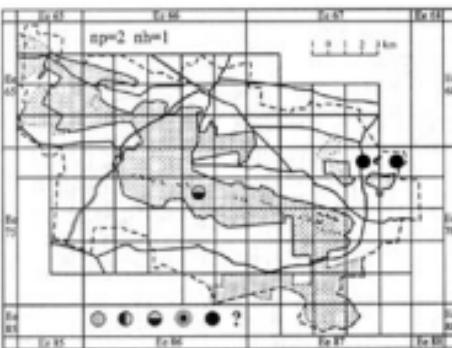
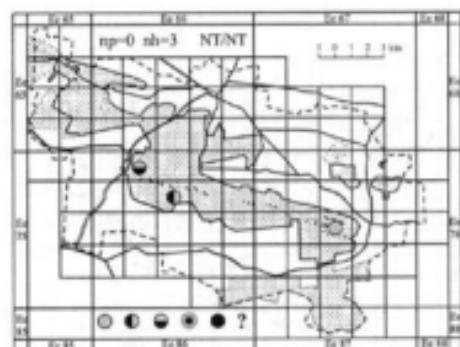
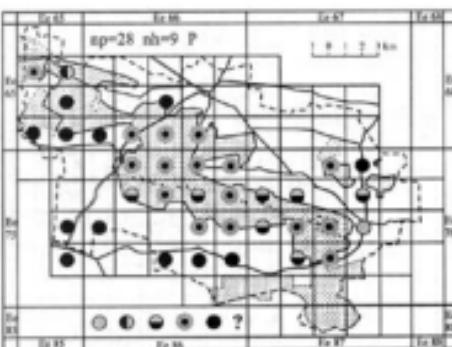
*Porina chlorotica* (Ach.) Müll. Arg.

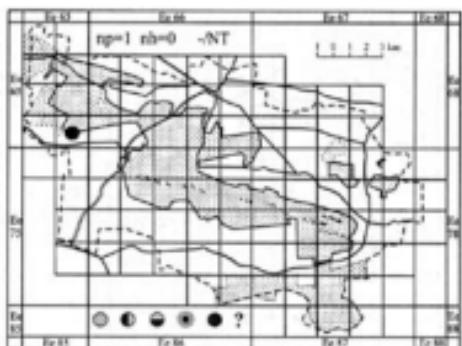
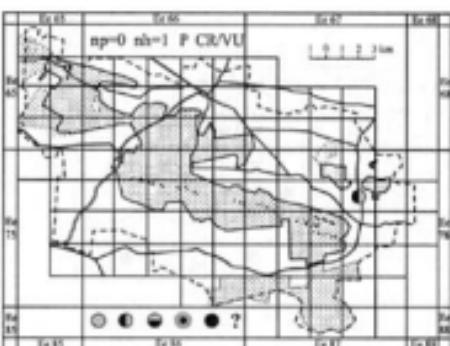
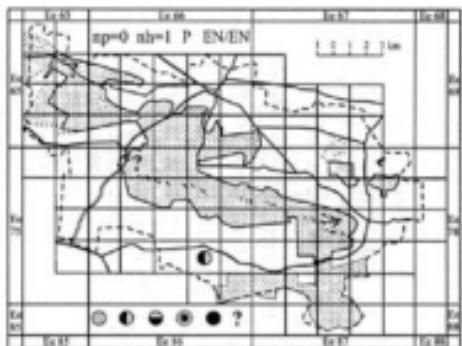
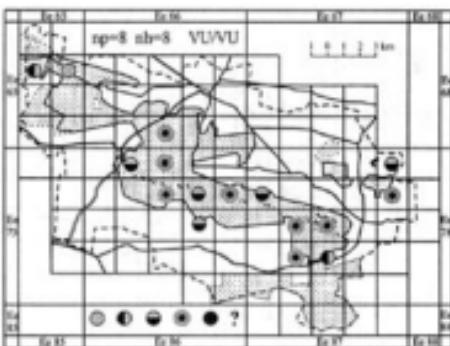
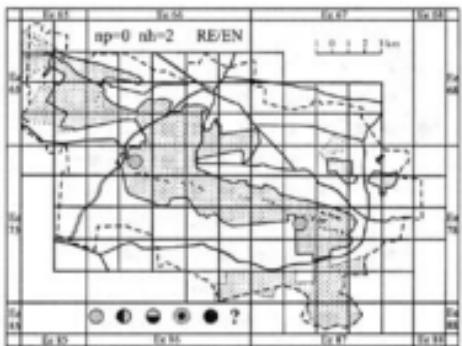
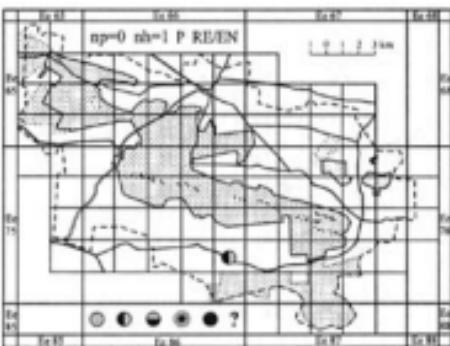


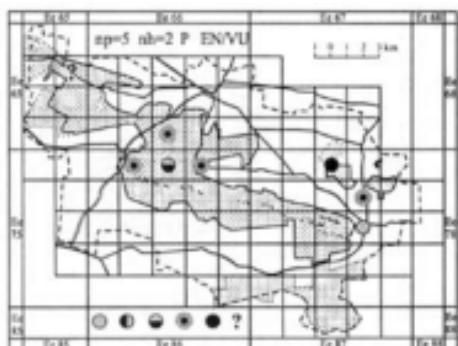
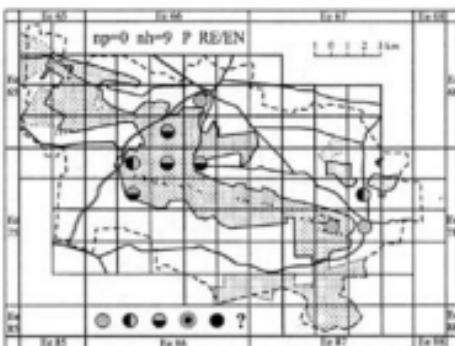
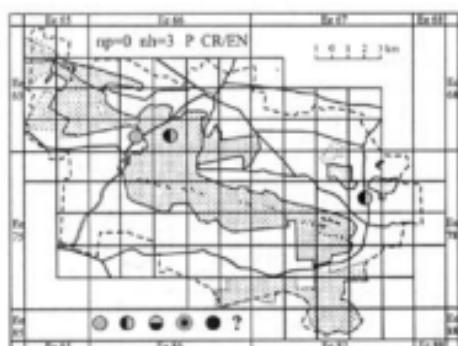
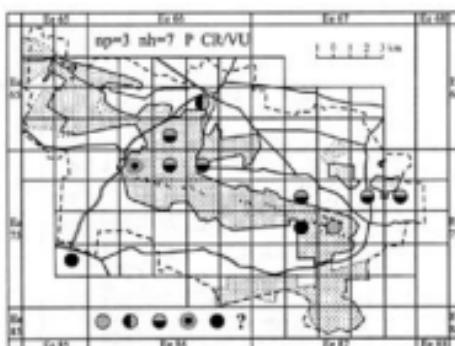
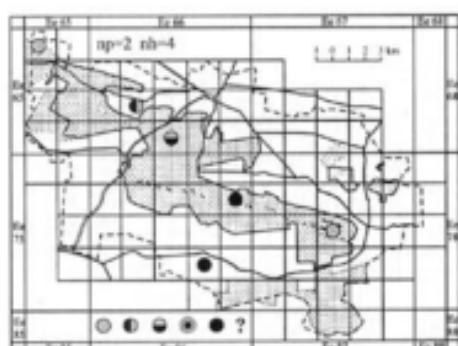
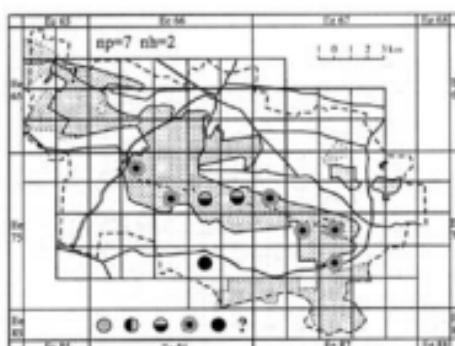
Porina leptalea (Durieu & Mont.) A.L. Sm.

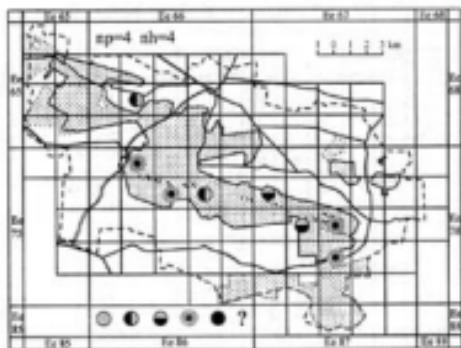
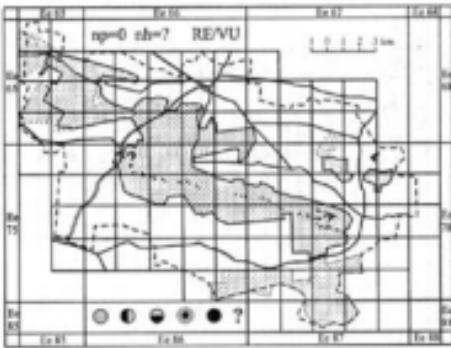
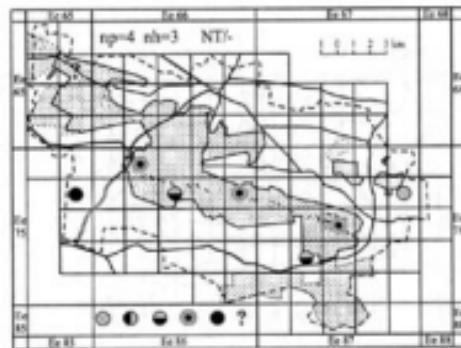
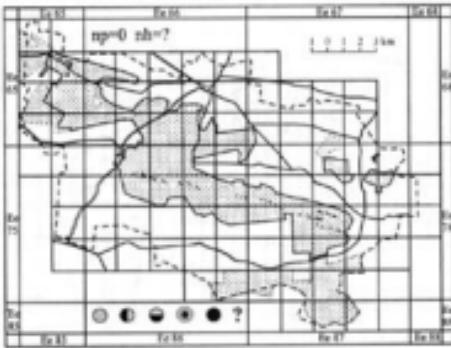
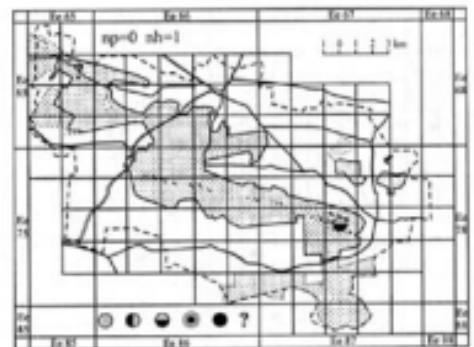
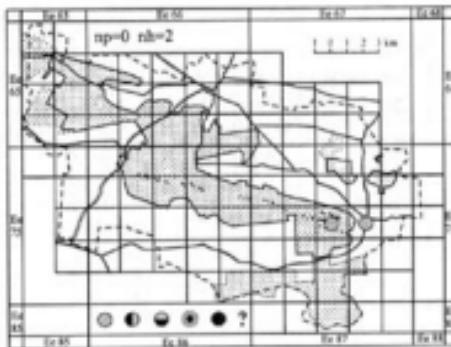


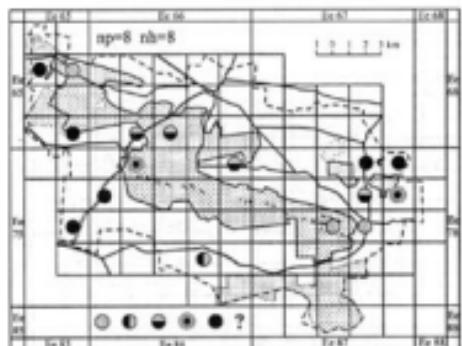
*Porpidia cinereoatra* (Ach.) Hertel & Knopf

*Porpidia crustulata* (Ach.) Hertel & Knoph*Porpidia macrocarpa* (DC.) Hertel & A.J. Schwab*Porpidia tuberculosa* (Sm.) Hertel & Knoph*Protoblastenia rupestris* (Scop.) J. Steiner*Protoparmelia badia* (Hoffm.) Hafellner*Pseudevernia furfuracea* (L.) Zopf.

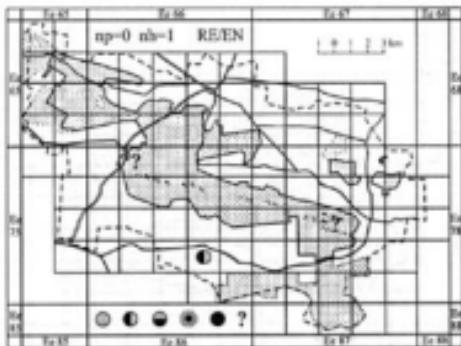
*Psilolechia clavulifera* (Nyl.) Coppins*Punctelia subrudecta* (Nyl.) Krog*Pycnothelia papillaria* Dufour*Pyrenula nitida* (Weigel) Ach.*Pyrenula nitidella* (Flörke ex Scher.) Müll. Arg.*Ramalina baltica* Lettau

*Ramalina farinacea* (L.) Ach.*Ramalina fastigiata* (Pers.) Ach.*Ramalina fraxinea* (L.) Ach.*Ramalina polinaria* (Westr.) Ach.*Rhizocarpon distinctum* Th. Fr.*Rhizocarpon geographicum* (L.) DC.

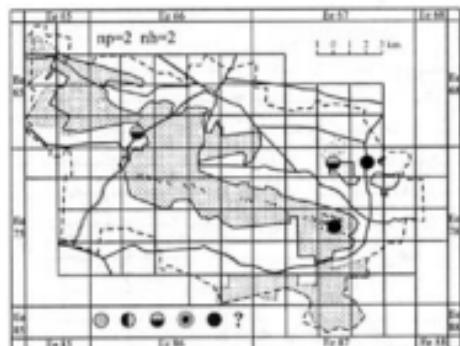
*Rhizocarpon obscuratum* (Ach.) A. Massal.*Rhizocarpon petraeum* (Wulfen) A. Massal.*Rhizocarpon polycarpum* (Hepp) Th. Fr.*Rinodina exigua* (Ach.) S. Gray*Rinodina gennarii* Bagl.*Rinodina immersa* (Körb.) Zahlbr.



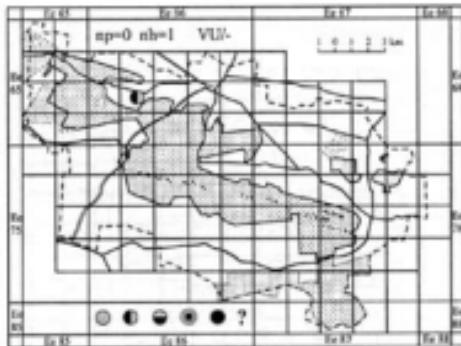
*Rinodina pyrina* (Ach.) Arnold



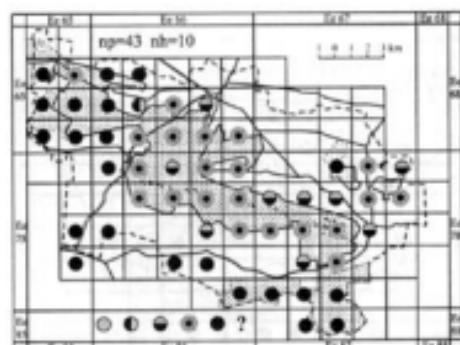
*Rinodina sophodes* (Ach.) A. Massal.



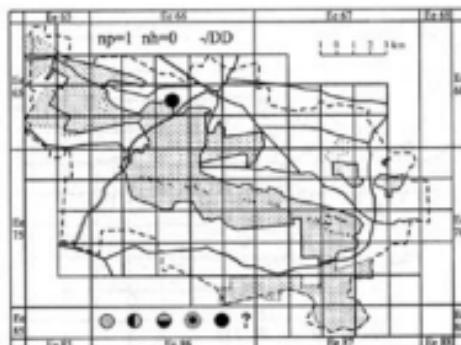
*Sarcogyne regularis* Körb.



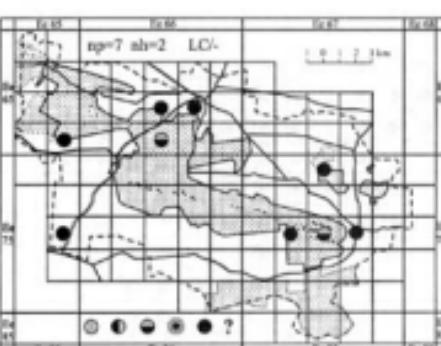
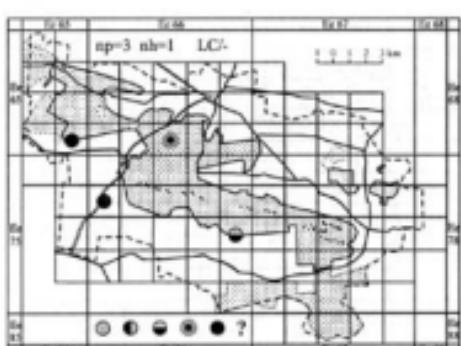
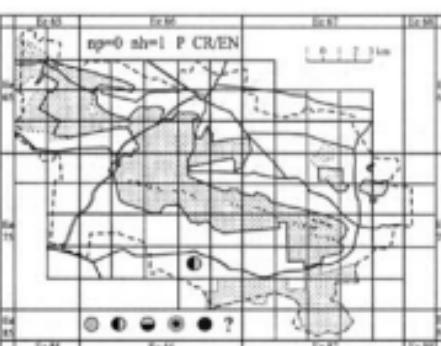
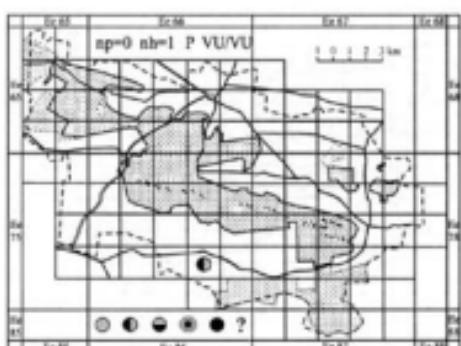
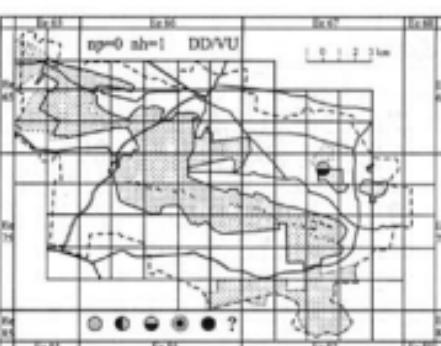
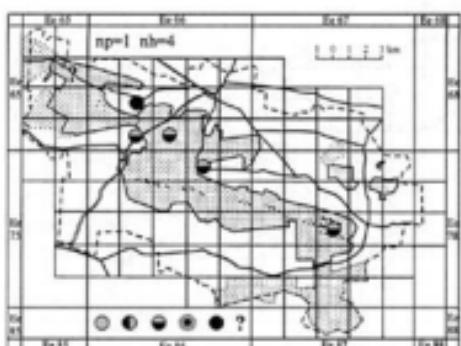
*Sarcosagium campestre* (Fr.) Poetsch & Schiederm.

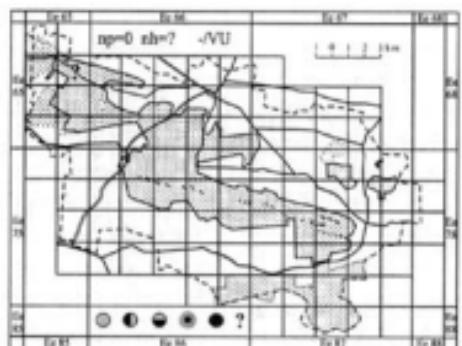
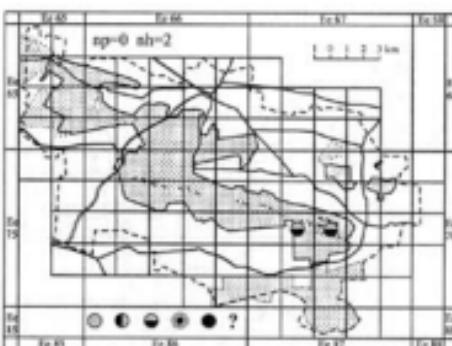
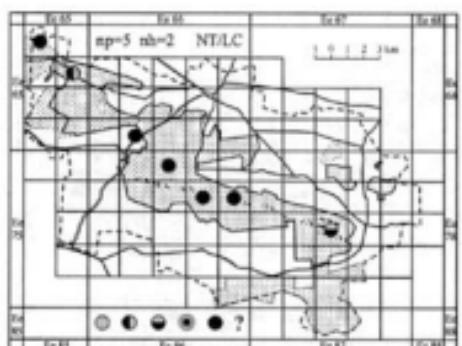
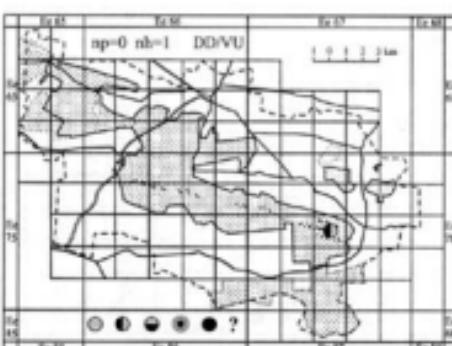
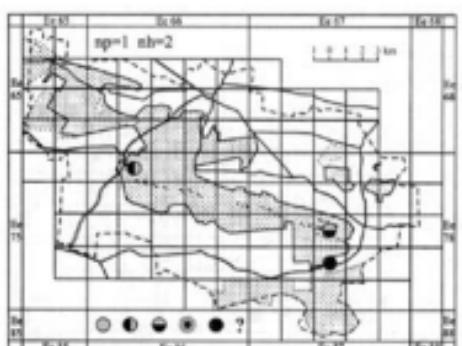
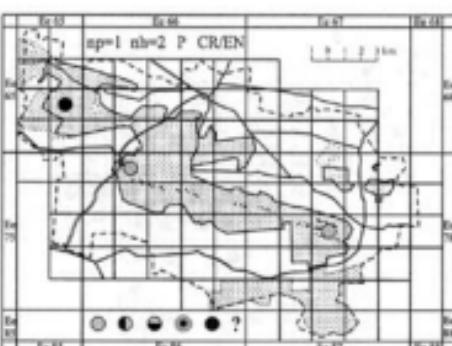


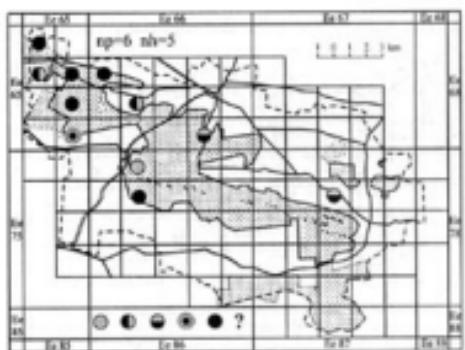
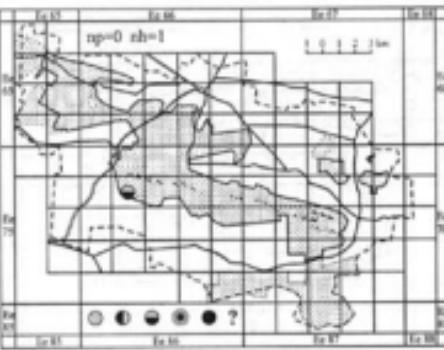
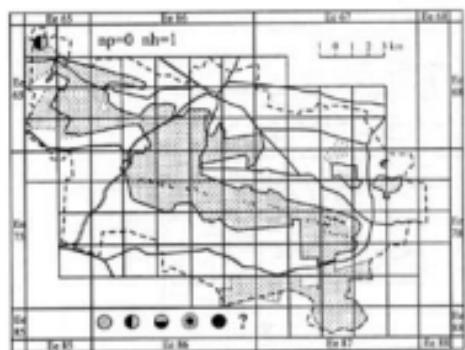
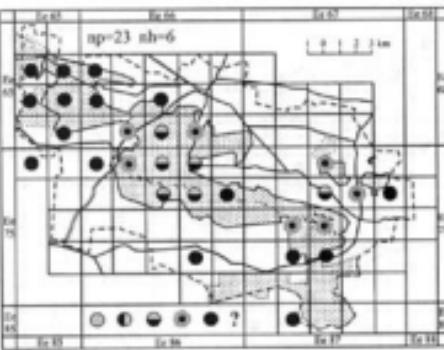
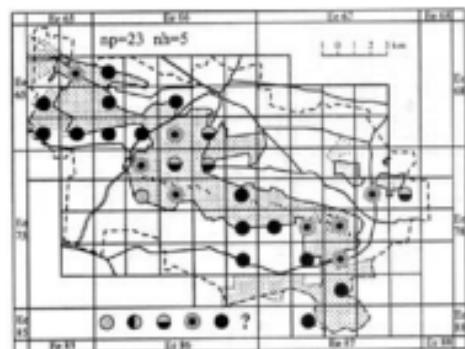
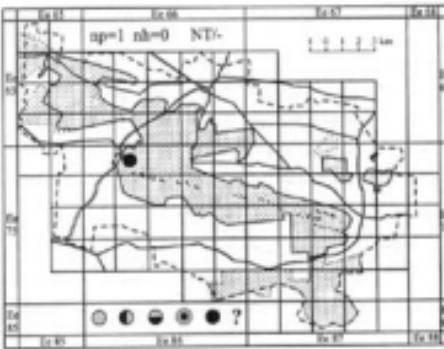
*Scoliciosporum chlorococcum* (Gracwe ex Stenb.) Vězda

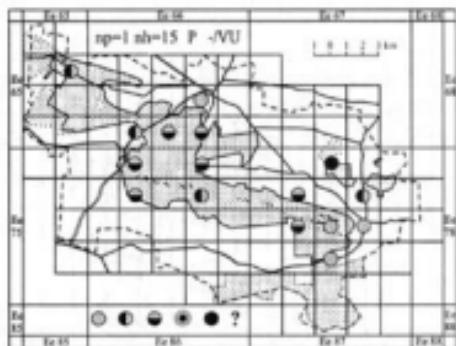


### *Scoliciosporum pruinatum* (P. James) V. zda

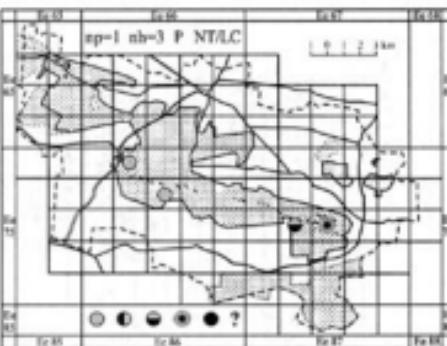


*Strigula stigmatella* (Ach.) R.C. Harris*Tephromela grumosa* (Pers.) Hafellner & Cl. Roux*Thelocarpon epibolum* Nyl.*Thelocarpon intermediellum* Nyl.*Thelocarpon laureri* (Flot.) Nyl.*Thelotrema lepadinum* (Ach.) Ach.

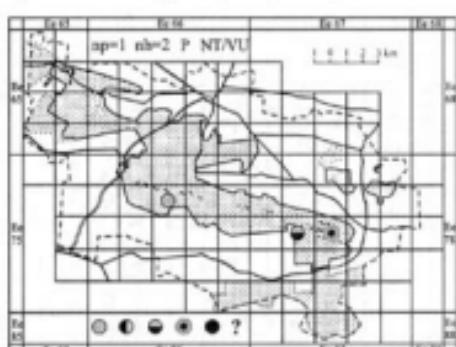
*Trapelia coarcata* (Sm.) M. Choisy*Trapelia involuta* (Taylor) Hertel*Trapelia obtegens* (Th. Fr.) Hertel*Trapeliopsis flexuosa* (Fr.) Coppins & P. James*Trapeliopsis granulosa* (Hoffm.) Lumbsch*Trapeliopsis pseudogranulosa* Coppins & P. James



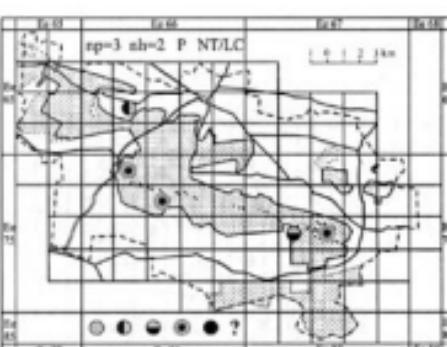
*Tuckermannopsis chlorophylla* (Willd.) Hale  
[*Cetraria chlorophylla* (Willd.) Vain.]



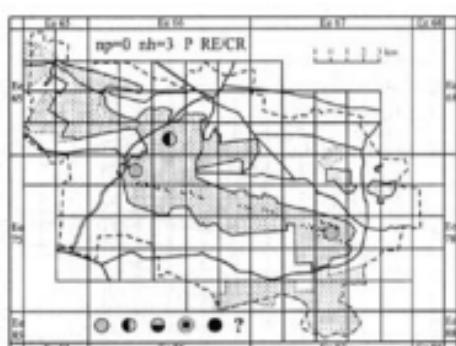
*Umbilicaria densta* (L.) Baumg.



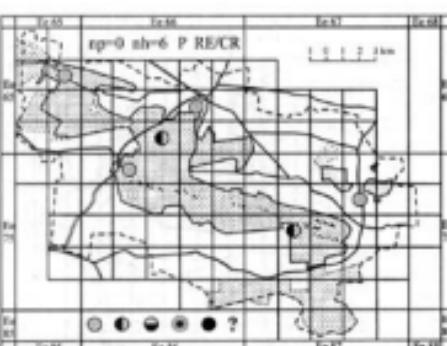
*Umbilicaria hyperborea* (Ach.) Hoffm.



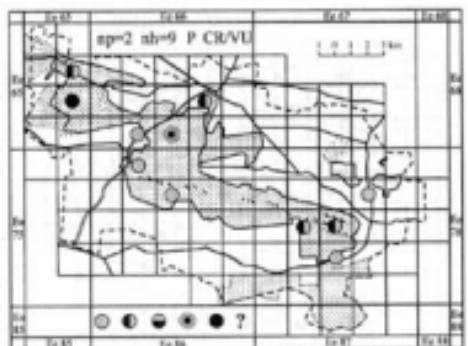
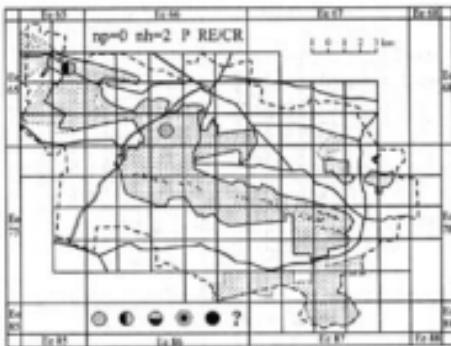
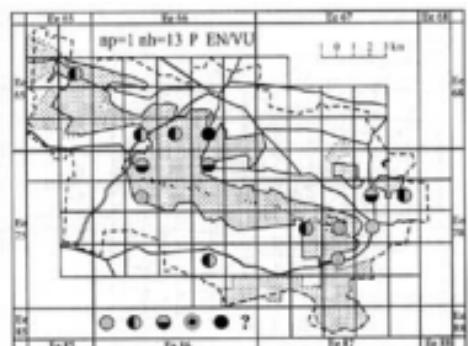
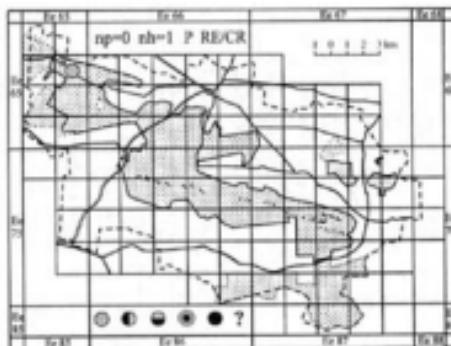
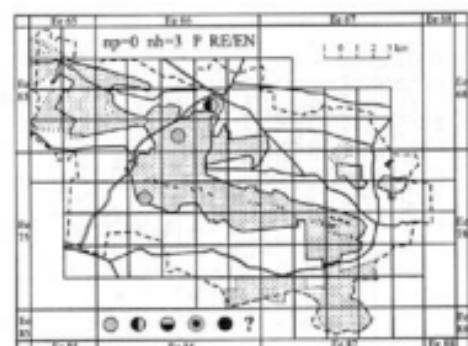
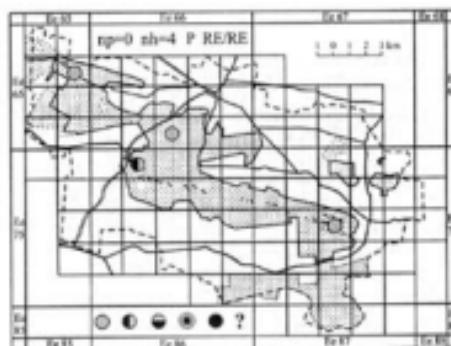
*Umbilicaria polyphylla* (L.) Baumg.

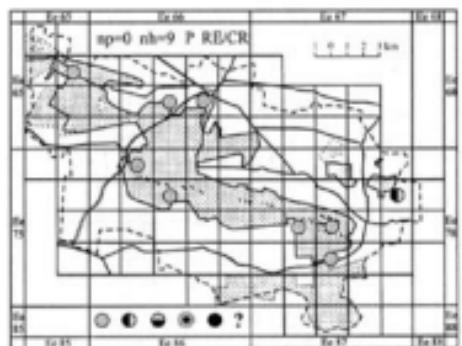
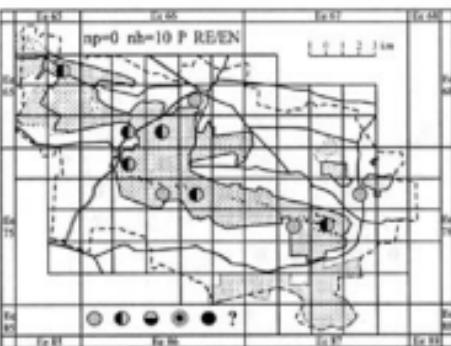
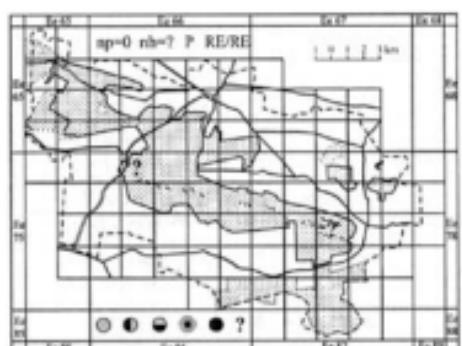
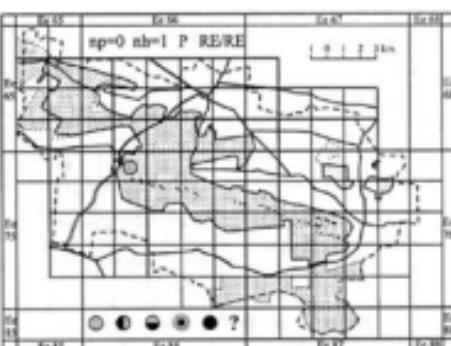
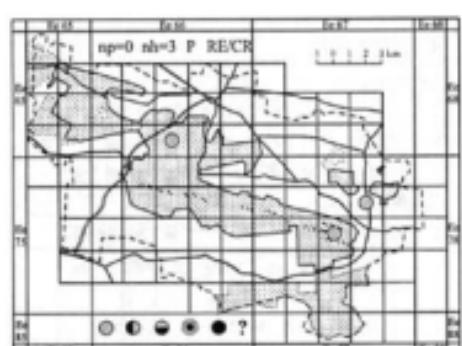
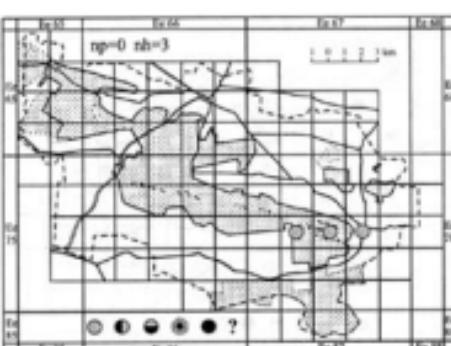


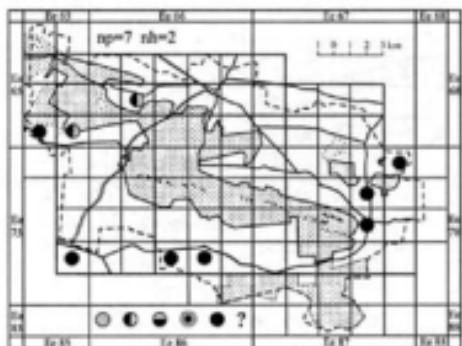
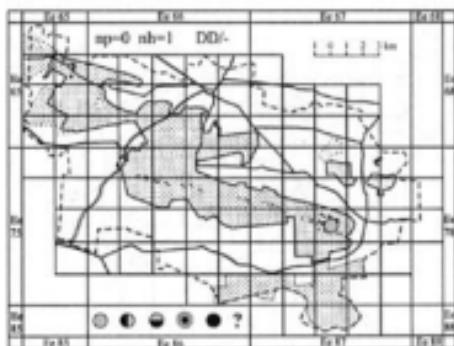
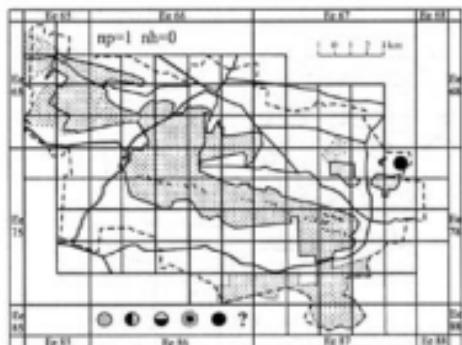
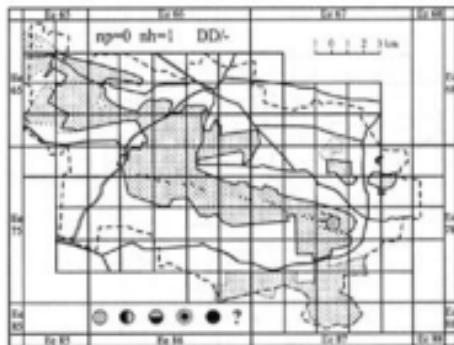
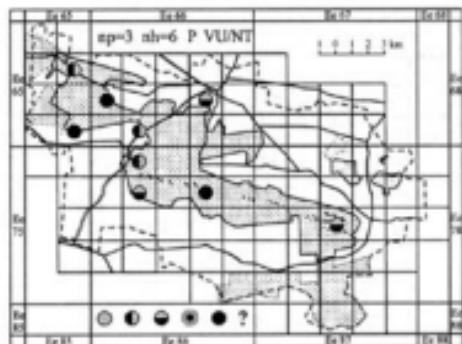
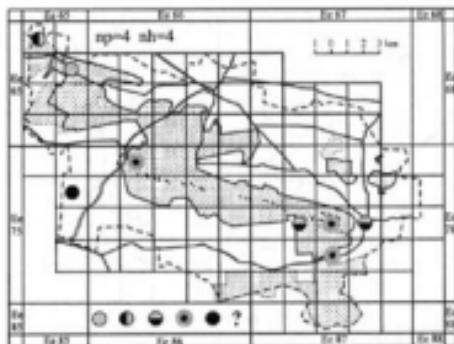
*Usnea ceratina* Ach.

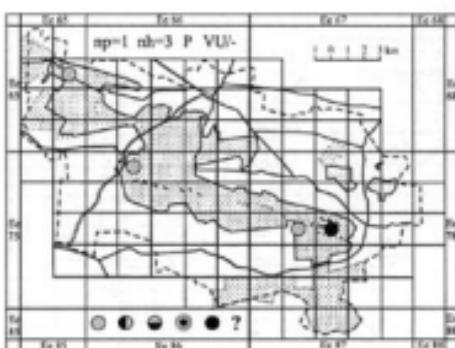
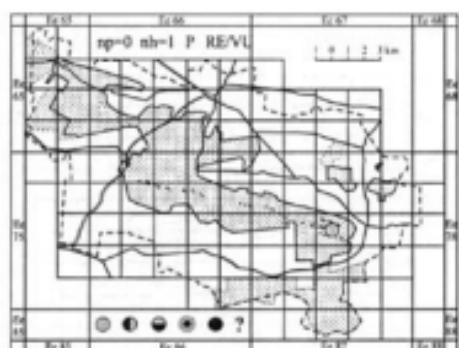


*Usnea faginea* Motsch.

*Usnea filipendula* Stirt.*Usnea fulvovireagens* (Räsänen) Räsänen*Usnea hirta* (L.) Weber F.H. Wigg.*Usnea hirtella* Motska*Usnea laricina* Vain.*Usnea neglecta* Motska

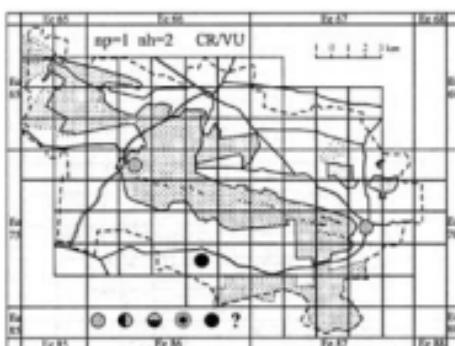
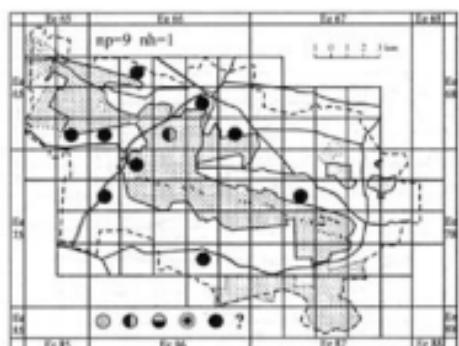
*Usnea rigidula* (Ach.) Motyka s.l.*Usnea subfloridana* Strit.*Usnea sylvatica* Motyka*Usnea uncinulata* Motyka*Usnea wasmuthii* Räsänen*Verrucaria calciseda* DC.

*Verrucaria muralis* Ach.*Verrucaria nigrescens* Pers.*Verrucaria rupestris* Shrad.*Verrucaria velana* (A. Massal.) Zahlbr.*Vulpicidia pinastri* (Scop.) Mattson & M.J. Lai*Xanthoparmelia conspersa* (Ach.) Hale



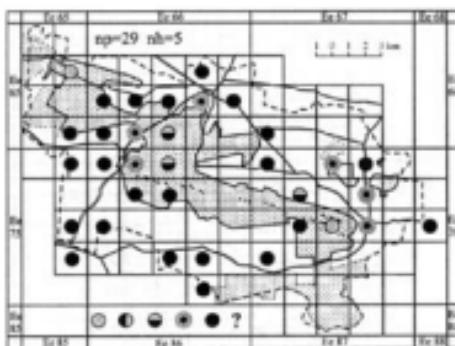
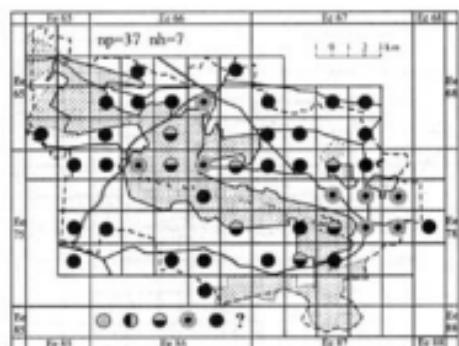
*Xanthoparmelia mougeotii* (Schaer. ex D. Dietr.) Hale

*Xanthoparmelia somloensis* (Gyeln.) Hale



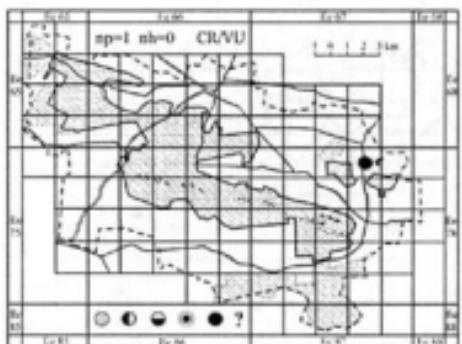
*Xanthoria candelaria* (L.) Th. Fr. s.l.

*Xanthoria fallax* (Hepp) Arnold

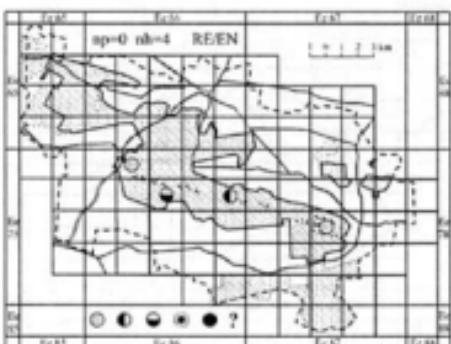


*Xanthoria parietina* (L.) Th. Fr.

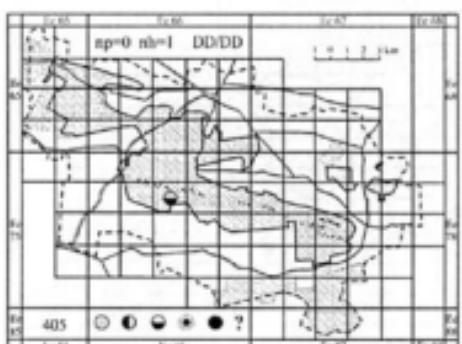
*Xanthoria polycarpa* (Hoffm.) Th. Fr. ex Rieber



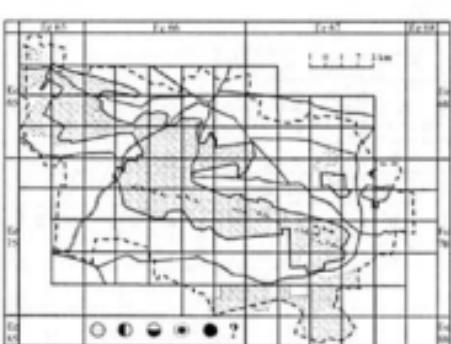
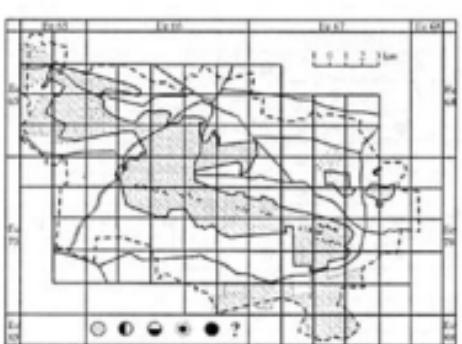
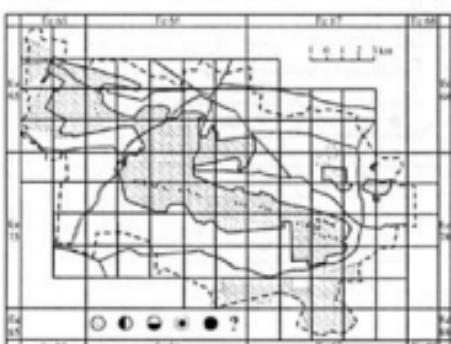
*Xanthoria ulophylloides* Räsänen

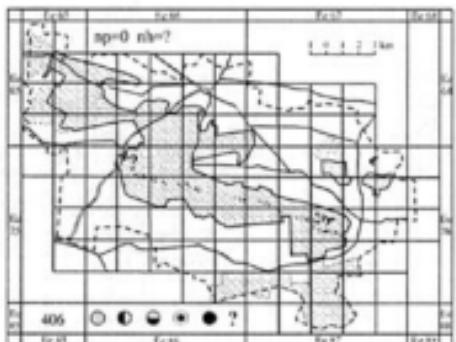


*Xylographa parallela* (Ach.: Fr.) Behren & Desberger

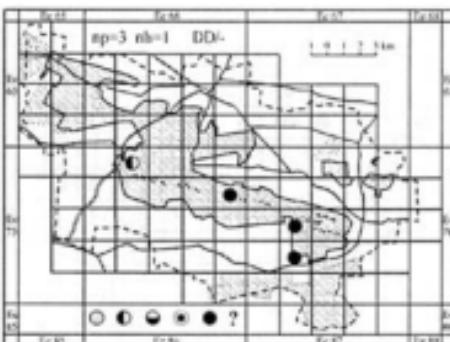


#### Xylagrenha vitiligo (Ach.) J.R. Laundon

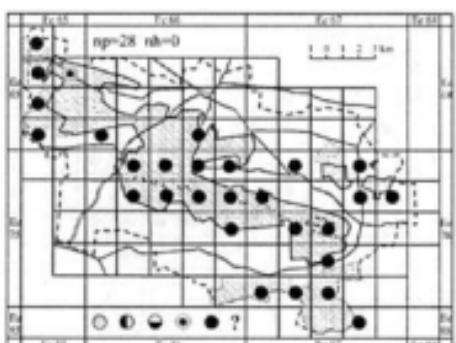




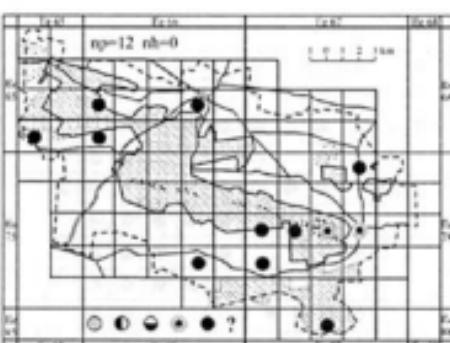
\**Abrothallus parmeilliarum* (Sommerf.) Arnold



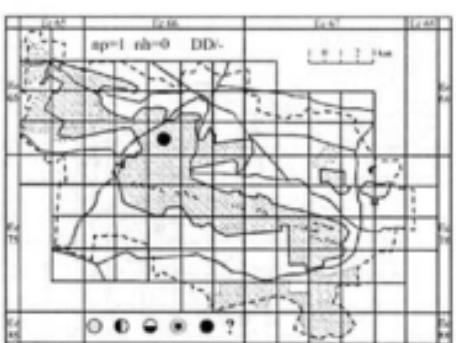
(\**Arthopyrenia lapponina* Anzi



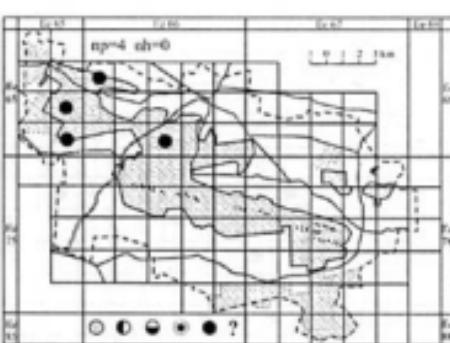
\**Athelia arachnoidea* (Berk.) Jülich



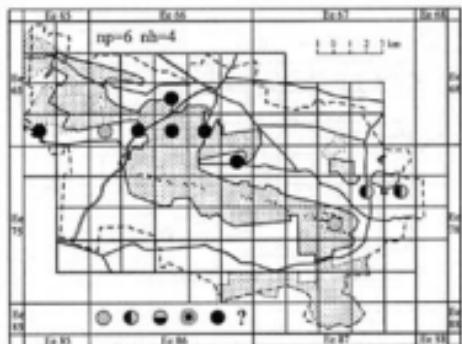
\**Bispora christiansenii* D. Hawksw.



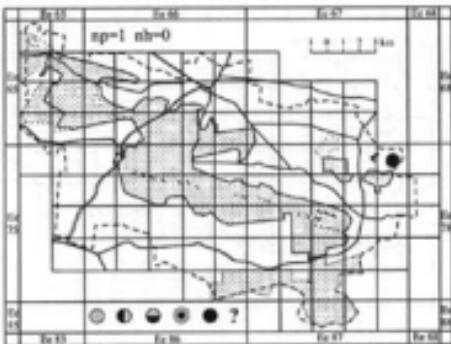
\**Chaenothecopsis nigra* Tibell



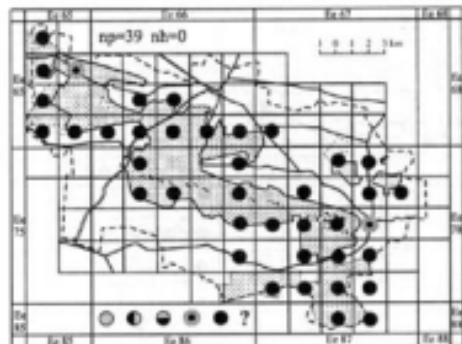
\**Chaenothecopsis pusilla* (Ach.) A.F.W. Schmidt



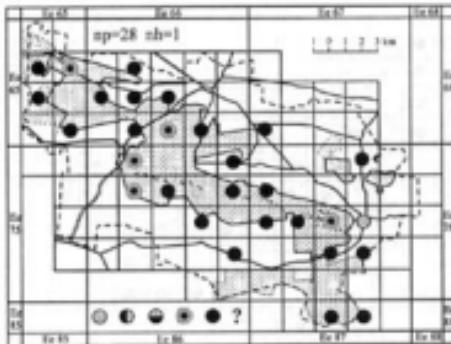
\**Clypeococcum hypocenomyctis* D. Hawksw.



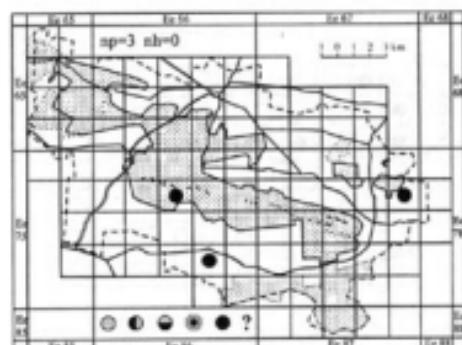
\**Illosporium carneum* Fr.



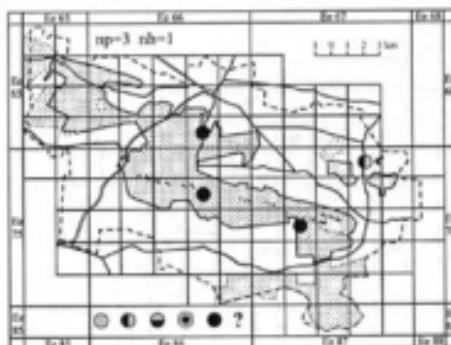
\**Lichenoconium erodens* M.S. Christ.  
& D. Hawksw.



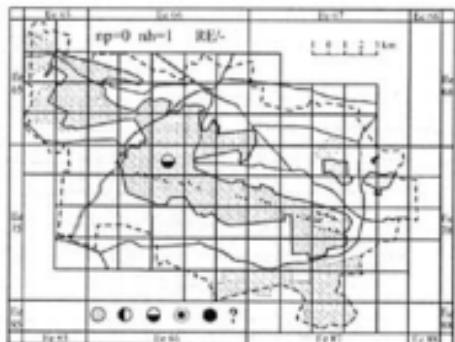
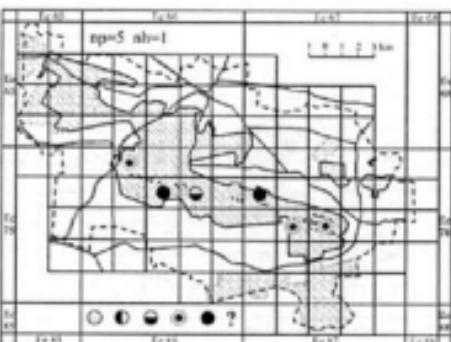
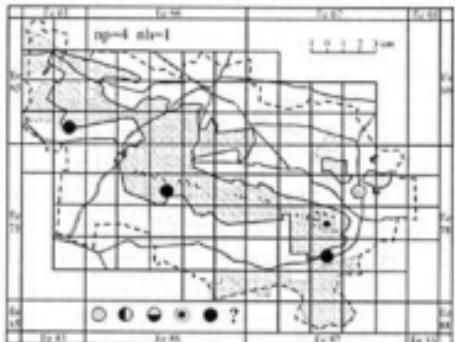
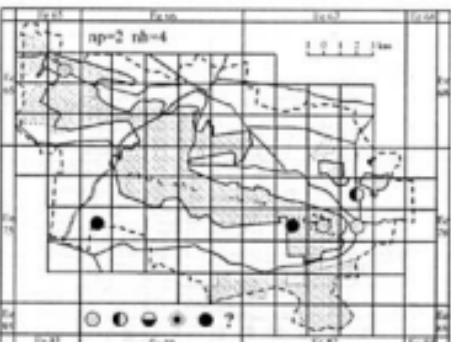
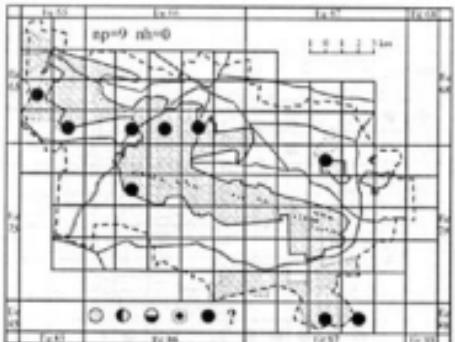
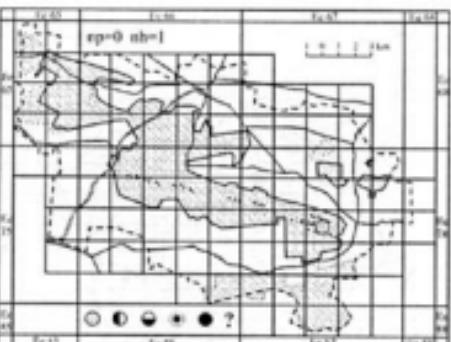
\**Lichenoconium lecanorae* (Jaap) D. Hawksw.

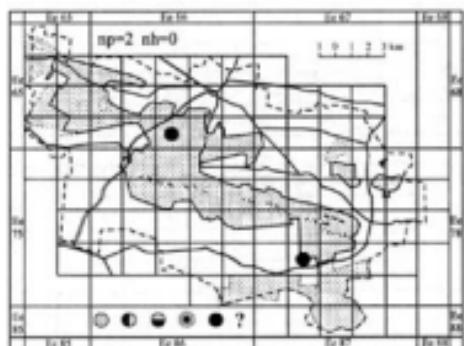
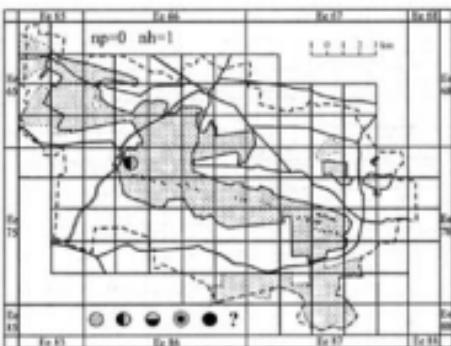
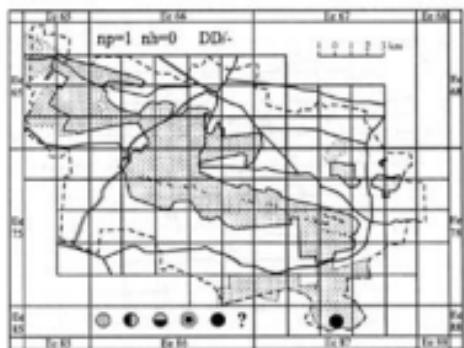
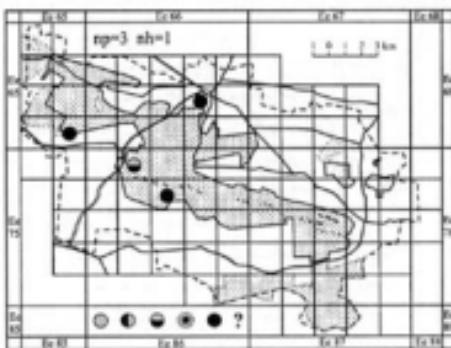
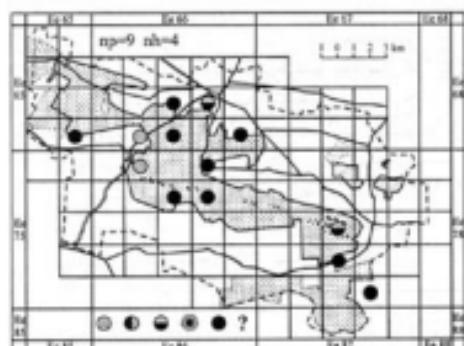
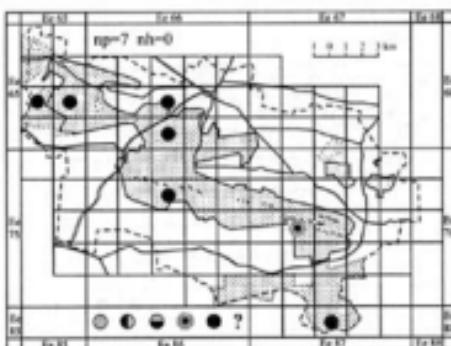


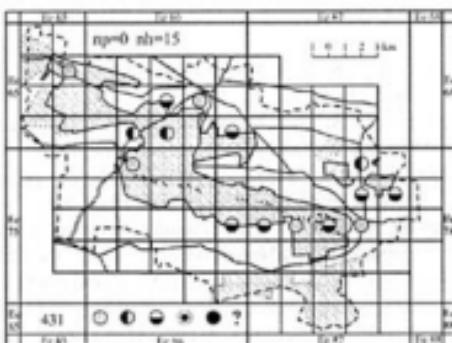
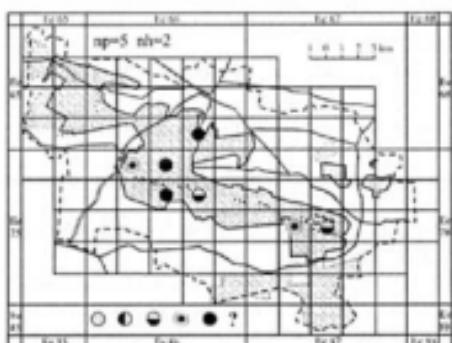
\**Lichenoconium pyxidatae* (Oudem.) Petr. & Syd.



\**Lichenodiplis lecanorae* (Vouaux) Dyko  
& D. Hawksw.

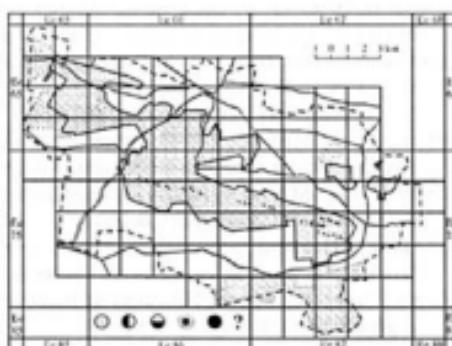
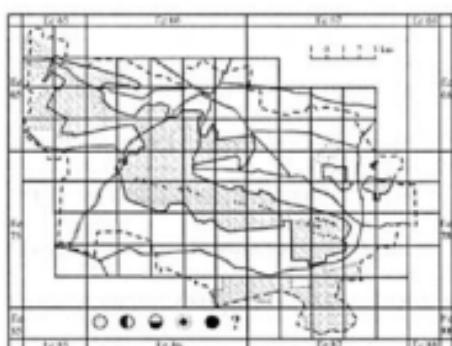
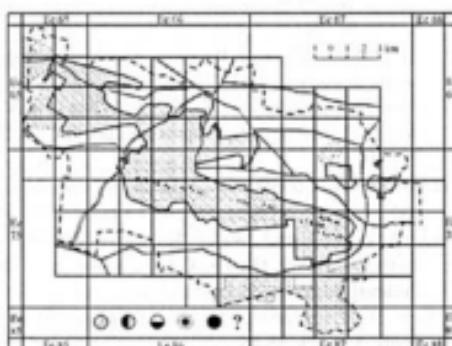
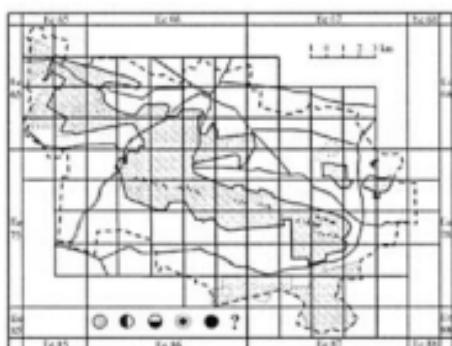
( )*Melaspilea gibberulosa* (Ach.) Zwackh\**Muellerella lichenicola* (Sommerf.; Fr.) D. Hawksw.*Mycocalicium subtile* (Pers.) Szatala( )*Periodothelia fuliginea* (Norman) D. Hawksw.  
[ ]*Mycomicrothelia micula* Köeb.\**Phoma cytopsora* (Vassaux) D. Hawksw.\**Plectocarpon lichenum* (Sommerf.) D. Hawksw.  
in D. Hawksw. & D.J. Galloway

*Stenocybe pullatula* (Ach.) Stein.\**Stigmidium congestum* Triebel in Rambold & Triebel\**Thelocarpon lichenicola* (Fuckel) Poelt & Hafellner\**Tremella cladoniae* Diederich & M.S. Christ.\**Tremella hypogymniae* Diederich & M.S. Christ.\**Tremella lichenicola* Diederich



\* *Vouauxiella lichenicola* (Linds.) Petr. & Syd.

\*\* *Phlyctis erythrosora* Erichsen



## REFERENCES

- Berdau F. 1876. Lišajniki istledovannye do sich por w oblasti Varšavskogo Učebnogo Okruga z ukazaniem do morfologii i fizjologii lišajnikov. Tipogr. K. Kovalevskego. Warszawa, 125 pp.
- Błoński F. 1890. Wyniki poszukiwań florystycznych skrytokwiatowych dokonanych w ciągu lata r. 1889 w obrębie 5 powiatów Królestwa Polskiego. Pam. Fizjogr. 10: 129-190.
- Bystrek J., Cieśliński S. 1976. Gatunki rodzaju *Bryopogon* Link emend. Bystr. na obszarze Górz Świętokrzyskich i ich pobrzeże. Fragm. Flor. Geobot. 22 (4): 553-557.
- Cieśliński S. 1975. Stan oraz wyniki badań nad florą porostów Górz Świętokrzyskich. Chrońmy Przyr. Ojcz. 31 (4): 18-23.
- Cieśliński S. 1981. Nowe i bardziej interesujące gatunki porostów naziemnych na Wyżynie Kielecko-Sandomierskiej i jej pobrzeżach. Fragm. Flor. Geobot. 27 (3): 527-539.
- Cieśliński S. 1985. Zmiany we florze porostów epifitycznych i epiksylicznych na obszarze Świętokrzyskiego Parku Narodowego. Roczn. Świętokrzyski 12: 125-142.
- Cieśliński S. 1991. Stan aktualny oraz zmiany we florze porostów naskalnych i naziemnych w Świętokrzyskim Parku Narodowym. Parki nar. Rez. przyr. 10 (3-4): 125-136.
- Cieśliński S. 2000. Porosty. (In:) S. Cieśliński and A. Kowalkowski (eds) Monografia Świętokrzyskiego Parku Narodowego. Świętokrzyski Park Narodowy, Bodzentyn-Kraków: 259-265.
- Cieśliński S., Bystrek J. 1982. Gatunki rodzaju *Usnea* Wigg. emend. Mot. na obszarze Górz Świętokrzyskich i ich wymieranie. Roczn. Świętokrzyski 10: 101-118.
- Cieśliński S., Fałtynowicz W. 1993. Note from editors. (In:) S. Cieśliński and W. Fałtynowicz (eds) Atlas of the geographical distribution of lichens in Poland I. W. Szafer Institute of Botany of Polish Academy of Sciences, Kraków: 7-8.
- Cieśliński S., Halicz B. 1971. Studia nad zespołami porostów Górz Świętokrzyskich. Bull. Soc. Sci. Łódz. Prace Wydz. III, Nauk Mat.-Przyr., 111.
- Cieśliński S., Lubek A. 2003. Czerwona lista porostów zagrożonych w Górzach Świętokrzyskich. (In:) K. Czyżewska (ed.) Zagrożenie porostów w Polsce [The threat to lichens in Poland]. Monogr. Bot. 91: 143-158.
- Cieśliński S., Taborowicz K. 1989. Nowe i bardziej interesujące gatunki porostów na Wyżynie Kielecko-Sandomierskiej. Fragm. Flor. Geobot. 34 (1-2): 173-184.
- Cieśliński S., Czyżewska K., Fabiszewski J. 2003. Czerwona lista porostów wymarłych i zagrożonych w Polsce. (In:) K. Czyżewska (ed.) Zagrożenie porostów w Polsce [The threat to lichens in Poland]. Monogr. Bot. 91: 13-49.
- Halicz B., Cieśliński S. 1967. Flora porostów modrzewia polskiego (*Larix polonica* Racib.). Zesz. Nauk. UŁ, 23: 35-45.
- Halicz B., Kuziel S. 1965. Research on the distribution and ecology of lichens occurring in the Świętokrzyskie Mountains. Bull. Soc. Sci. Lett. Łódź, Cl. Sci. Math.-Nat., 16 (9): 1-20.
- Halicz B., Kuziel S. 1966. Some data concerning rock and terrestrial lichens occurring in the Świętokrzyskie Mountain. Bull. Soc. Sci. Lett. Łódź, Cl. Sci. Math.-Nat., 17 (22): 1-11.
- Hawksworth D.L. 1983. Key to fungi occurring on lichens in the British Isles. Lichenologist 15: 3-44.
- Kobenda R. 1939. Goloborza i ich stosunek do lasu w Górzach Świętokrzyskich. Inst. Bad. Lasów Państw. Rozprawy i sprawozdania, Ser. A, 43.
- Kobenda R., Motyka J. 1928. Führer durch die „Goloborza“ - Blockhalden des Lysogóry - Höhenzuges. Guide des excursion en Pologne, Kraków 13: 1-8.
- Kobenda R., Motyka J. 1929. La végétation des éboulis des Monts de S-te Croix. Bull. Acad. Pol. Sci. Lett., Cl. Sci. Math.-Nat., B, Sci. Nat. 1: 175-207.
- Lubek A. 2004. Antropogeniczne przemiany porostów (grzybów naporostowych) w Świętokrzyskim Parku Narodowym. Doctoral thesis, University of Łódź.
- Scholz P. 2000. Katalog der Flechten und flechtenbewohnenden Pilze Deutschlands. Schriftenreihe für Vegetationskunde, Heft 31, 298 pp.
- Tobolewski Z. 1962. Materiały rozmieszczenia rodziny Caliciaceae (Lichenes) w Polsce. Materiały do flory porostów Tatr. PTPN, Wydz. Mat.-Przyr., Prace Komisji Biol. 24 (2): 1-28.
- Taborowicz K. 1977. Nowe stanowiska gatunków rodzaju *Thelecarpon* Nyl. w Polsce. Fragm. Flor. Geobot. 23 (2): 253-256.
- Věžda A., Liška J. 1999. Katalog lišejníků České Republiky. Institute of Botany Academy of Sciences of the Czech Republic, Práhovice, 283 pp.

- Wróblewski T. 2000. Charakterystyka orograficzna i toponimia. (In:) S. Cieśliński and A. Kowalkowski (eds) Monografia Świętokrzyskiego Parku Narodowego. Świętokrzyski Park Narodowy, Bodzentyn-Kraków, 46-50.
- Dz.U. Nr 168, poz. 1765, Rozporządzenie Ministra Środowiska z dnia 9 lipca 2004 r. w sprawie gatunków dziko występujących grzybów objętych ochroną.

## Rozmieszczenia porostów i grzybów naporostowych w Świętokrzyskim Parku Narodowym

### Streszczenie

W pracy przedstawiono rozmieszczenie 405 gatunków, podgatunków i odmian porostów oraz 26 gatunków grzybów naporostowych.

Materiały badawcze pochodzą z trzech źródeł: z literatury publikowanej i niepublikowanej, z historycznych zbiorów zielnikowych oraz z własnych badań terenowych przeprowadzonych w latach 1999-2002.

Rozmieszczenie przestrzenne porostów i grzybów naporostowych przedstawiono na kartogramach. Pojedynczy kartogram – sieć kwadratów o bokach 2 km dostosowana do ogólnopolskiej sieci ATLICHEN – wchodzi w skład serii kartogramów i odpowiada jednemu taksonowi porostu lub grzybowi naporostowemu. Kartowaniu poddano również historyczne materiały publikowane, niepublikowane i zielnikowe. Na pojedynczej mapie zawarto następujące dane: nazwa gatunku, obecna i historyczna liczba stanowisk, ochrona ścisła lub częściowa, status zagrożenia w Górzach Świętokrzyskich/Polsce, kolejne przedziały czasowe, w których stwierdzony został gatunek, sygnowane różnymi znakami.