Development and morphology of Micromycopsis mirabilis Canter

JOANNA ZOFIA KADŁUBOWSKA

In 1949 Canter described Micromycopsis mirabilis on the basis of material from the water bodies in Great British. He found M. mirabilis parasitizing in the cells of Closterium Isunula, C. costatum, C. disc., C. kitzingii and Closterium ap.; however he could not investigate the whole cycle of development but only the prosorus and sorus stages of this fungus.

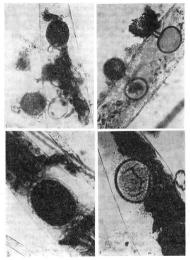
In the samples taken on 13 May, 1966 from the peet bog Przerpeise near Lubies (Belchatów district in the Łość region) parasitis Micromycopsis mirabilis Canter were noticed multiplying in a number of cells of Closterium lumula (Mill.) Nitzech. in the samples taken from the same site on 12 June, 1967, only resting spores in one cell of Closterium, so, were found. Material gathered from Toporwoe Stawki (Tatra Mas,) on 22 September, 1963 revealed one resting spore in the cell of Closterium, Insula.

Morphological features

1. Samples from the peat bog Przerębiec

Prosori (Figs. 1, 2) ellipsoidal, surrounding cell wall with concentric short spines. Prosorus length 30−3π µ (mean length 40 µ, confidence interval 28—38 µ). Diameter of prosorus discharge tube about 5 n length of discharge tube dependent on the distance of the prosorus from the host cell wall. Sori spherical (Figs. 1, 2), surrounded with brownish cell wall bearing short spines. The sorus consists of several tens of sporangia about 5 µ in diameter. Sorus diameter 30—50 µ (mean diameter 38 µ, confidence interval 31—41 µ). Resting spores (Fig. 3) surrounded with a three-layer cell wall: endosporium dark-brown thin, mesosporium also thin colourless, exosporium colourless, thick, bearing numerous verruese.

In the sample of 13 May, 1966 only two resting spores were found, $62 \times 68 \,\mu$ and 68×64 in size.



Figs 1—4. Micromycopsis mirabilis: 1— two prosori in Closterium lumula cell; on the right: one prosorus with developed discharge tube, the other with a sorus outside of the heat sporangium; 2— two prosori with discharge tubes and sorl. Figs 3 and 4— resting spore in Closterium lumula cell; 3— thick verucous exposurum is clearly seen: 4— sents an other resting spore cell wall are well visible

In the sample of 12 June 1967 the stage of resting spores was solely noticed. In the middle of the Clusterium sp. cell three ellipsoidal resting spores occurred. Because of the deformation of Closterium thromatophores the species could not be identified. Resting spores were surrounded with a dark-brown thin layer of endosporium, with colourless thin layer of mesosporium and a thick colourless layer of exosporium bearing numerous verrucae. Resting spores $40 \times 42 \, \mu$, $45 \times 50 \, \mu$ and $50 \times 63 \, \mu$ in Size.

2. Sample from Toporowe Stawki

The preserved material revealed only one resting spore in a Closerium cell. On the spore cell wall some septa could be seen (Fig. 4). It was not clear, however, whether the septa penetrated into the spore or only divided the cell wall into several segments. Resting spore size was 60×672.5

Development cycle

The sample taken from the peat bog Præcrebiec on 13 May 1986 was placed in a glass vessel at room temperature. On the first day of observation brownish proson; were noticed in the cells of Closterium Inuula. The chromatophores were slightly damaged and pushed towards the cell wall. In the host cells generally two proson; locurred. After 7 days the discharging tubes began to develop on the proson; (Fig. 1) and perforated the host cell wall. At the tips of discharge tubes just by the Closterium cell wall soni consisting of several tens of spherical spores were formed (Figs. 1, 2). The contents of proson escaped through the discharge tube into the soni; The preson; turned almost colourless and the relief of their cell wall became conspicuous. On the tenth day of observation some sori split (Fig. 2), and discharged spores. After a fortnight in the middle of Closterium Inuula cells resting spores formed in place of the nucleus occurrence (Fig. 3).

DISCUSSION

Micromycopsis mirabilis Canter is a new genus and species for Polish flora. To the authors knowledge the peat bog at Przerębiec and Toporowe Stawki are so far, beside Great Britain, the only places of Micromuconsis mirabilis occurrence in the world.

Micromycopsis mirabilis during its development cycle produces not only prosori and sori described by Canter but also resting spores Figs 3, 4). Resting spores are surrounded by three-layer cell wall: brown, thin, smooth-walled endosporium; colourless, thin, smooth-walled mesosportum, and colourless, thick, verrucous excoporium. Septa divide the cell wall of retting apprex into several asymmets (Fig. 4). The related of the cell wall bears a close resemblance to that of Synchyrium endebiciesm (Schills) Perc (figure in Curtik's paper, 1921). Such a sculpture of the cell wall provides evidence that the development cycle of M. mirabilis includes also the stage of restrict market.

Sparrow (1960) included genus Micromycopsis Scherffel 1926 in the genus Micromyces Dangeard 1839. This does not seem correct since in genus Micromycopsit the prosorus content is discharged into the sorus outside the host, whereas in genus Micromyces the prosorus is deprived of the discharge tube and the sporangia or sorus are formed inside the host. Cejp (1957) and Skirgiello (1954) distinguish both there genera.

The materials are to be found in the Institute of Algology, Department of Taxonomy and Geography of Plants. University of Łódź.

REFERENCES

Cejp K., 1957, Houby, Praha, C. A. V.

Curtis K. M., 1921, The life history and cytology of Synchytrium endobioticum (Schilb.) Perc., the cause of wart disease in potato. Phil. Trans. Roy. Soc. London, Sept. B. 40400, 479.

London, Ser. B, 210:409-478.
Skirgiello A, 1945, Grzyby niższe. Warszawa, PWN.
Sparrow F. K. 1960. Aquatic Phycomicetes. Sec. Rev. Ed. Ann. Arbor. The

University of Michigan Press.

Rozwój i morfologia Micromycopsis mirabilis Canter

Streszczenie

Opisano z okolicy Lodzi Micromycopsis mirabilis Canter, grzyb wodny pasożytujący na przedstawicielach rodzaju Closterium, Grzyb ten znany był dotychczas tylko w Wielkiej Brytanii, obecnie stwierdzono jego występowanie również w Polsce.