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ORIGINAL RESEARCH PAPER in MORPHO-ANATOMY OF USABLE PLANTS – FUNCTION AND ADAPTATION

Micromorphology and Anatomy of the Flowers in *Clivia* spp. and *Scadoxus multiflorus* (Haemantheae, Amaryllidaceae)

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Abstract

The general morphology, micromorphology, and anatomy of the flowers of Clivia miniata, Clivia nobilis, and Scadoxus multiflorus were studied using light microscopy. The studied species have large syntepalous and trimerous flowers, short floral tubes with adnate stamens, and inferior ovaries that develop baccate fruit. The gynoecium in the studied species consists of synascidiate, symplicate, and hemisymplicate zones. The style is composed of postgenitally fused carpels. The few ovules are located in a locule in the synascidiate and symplicate zones in C. miniata and C. nobilis, whereas in S. multiflorus, solitary ovules occupy the synascidiate zone in each locule. The septal nectaries are located in the hemisymplicate zone and occupy the uppermost 29% to 56% of the ovary height. Septal nectaries are of the nonlabyrinthine lilioid-type, covered with secretory tissue only in its lower portion. Nectary channels are apical or subapical and open near the style base. A common pattern of the venation of the floral parts was observed in all species: Tepal traces and stamen traces were fused in the ovary wall, the style was supplied by dorsal carpellary bundles, and ovules were supplied by ventral carpellary bundles entering the ovary from the bottom. The observed gynoecium inner structure provides adaptations for the development of fleshy fruit, with thickened parenchymous ovary wall, ovary base, and ovary roof, and numerous branched vascular bundles in the ovary wall around locules.

Keywords

Clivia miniata; *Clivia nobilis*; *Scadoxus multiflorus*; inferior ovary; septal nectary; vascular bundle; vertical zonality

1. Introduction

The family Amaryllidaceae s. l. is represented by highly ornamental plants that are commonly found on all continents, except Antarctica (Takhtajan, 2009). Many representatives are found in tropical and subtropical zones and less often in temperate climates. Most genera grow in Central and South America, Southern Africa, and the Mediterranean. Amaryllidaceae are perennial herbaceous plants that can grow up to 2 m in height. The main feature of this family is the presence of subterranean or aerial bulbs. The flowers are gathered in umbels and sometimes cymes. There can sometimes be a reduction in inflorescences and the formation of single flowers (Meerow & Snijman, 1998). The family was established in 1805 and now has approximately 1,600 species and approximately 75 genera, 17 tribes, and three subfamilies: Agapanthoideae Endlicher, Allioideae Herb., and Amaryllidoideae Burnett (The Angiosperm Phylogeny Group, 2016). The genera *Clivia* Lindl. (six species) and *Scadoxus* Raf. (9–12 species) belong to the tribe Haemantheae of the subfamily Amaryllidoideae, which has six genera (Stevens, 2001).

Representatives of the genus *Clivia* have a large number of varieties, the study of which is gaining popularity (Petravich et al., 2015). *Clivia* is also the study subject of frequent fungal diseases in tropical plants (Li et al., 2020; Liu et al., 2020). The toxicity of *C. miniata* fruits and seeds has been studied (Kiepiel & Johnson, 2019), as well as the genes involved in the formation of *Clivia* leaves (Qin-Mei et al., 2018; Wang et al., 2016). Changes in the pollinator community of *Scadoxus cinnabarinus* along its altitude belt on Mount Cameroon have been studied by a group of Czech and African scientists (Mertens et al., 2020).

Using sequences from the plastid region *trn*L-F and internal transcribed spacer (ITS) nrDNA, Meerow and Clayton (2004) investigated the phylogeny of the African tribe Haemantheae on 19 species of all genera of the tribe. ITS and combined matrix parsimony analysis generated a well-maintained tree. Two main clades were detected. One included the monophyletic rhizomatous genera *Clivia* and *Cryptostephanus*. A larger clade combines *Haemanthus* and *Scadoxus* as a sister group to the *Apodolirion-Gethyllis* subclade (Meerow & Clayton, 2004).

Flowering efficiency, increasing the growing degree days required for anthesis time, and the effect of cold storage (12 °C for 4 weeks) before forcing were evaluated in *Scadoxus multiflorus* subsp. *katharinae* (Baker) Friis & Nordal. (Funnell, 2008). The results of butterfly-wing pollination in two subspecies of *S. multiflorus* and a review of the prevalence of this pollination mechanism among other Amaryllidaceae in Southern Africa was recently published (Butler & Johnson, 2020). Karyotypic features include organizations of the 5S and 45S rDNA loci, telomeres of *S. multiflorus* (Monkheang et al., 2016), and tracking biological footprints of climate change using the flowering phenology of the geophytes in *Pancratium tenuifolium* and *S. multiflorus* (Kwembeya, 2021). The effects of light duration on flower development in *S. multiflorus* have been described and evaluated (Vendrame et al., 2004).

Haemantheae is distinct from the other Amaryllidaceae in its rhizomatous habit, baccate fruit, and phytomelan-free seeds (Meerow & Snijman, 1998). Precise examination of the gynoecium with the inferior ovary and septal nectaries in the members of the tribe has not been performed. However, the gynoecium structure in monocots has been actively assessed with regard to the evolution of epigyny, loss of septal nectaries, and pollination mechanisms (Remizova et al., 2010; Rudall, 2002). Amaryllidoideae is the only clade within the higher Asparagales in which the inferior ovary occurs (Rudall, 2002).

Species of *Clivia* are rhizomatous, evergreen plants that naturally grow in Southern Africa. Representatives of the genus *Clivia* have actinomorphic or weakly zygomorphic flowers, narrowly to widely funnelform, have a pendulous spread, and a short floral tube. The filaments are free, and the stigma is tricuspidate. There are four-to-five ovules per locule. The berry is subglobose and red. The seeds are turgid and ivory-colored (Meerow & Snijman, 1998). *Clivia miniata* has an erect, subterranean, or shortly aerial rhizome, with thick fleshy roots. The umbel has 16 to 30 flowers raised above the leaves. The stamens are weakly to strongly declinate. The fruit is a baccate capsule with few seeds growing from the inside of the capsule (Duncan, 2017). *Clivia nobilis* has a pendent umbel of 20 to 50 flowers raised below the leaf tips (Duncan, 2017).

The genus *Scadoxus* is characterized by actinomorphic flowers, funnelform to salverform, with perigonium segments equal to or longer than the tube. The stamens are free, and the stigma is undivided. There are just a few ovules per locule. The berry is ovoid to globose and red. The seeds are ovoid and ivory-colored (Meerow & Snijman, 1998). *Scadoxus multiflorus* is a deciduous, summer-growing plant with rhizomatous bulbs, which grows naturally in sub-Saharan Africa, Arabian Peninsula, and Seychelles (Duncan, 2017). Umbels are many-flowered and spherical, and leaf blades are ovate, erect, and shiny, with two or three reduced, sheathing basal leaves.

The aim of our study was to conduct a comparative morphological analysis using micromorphological and anatomical characteristics of the flower in representatives of the tribe Haemantheae with regard to gynoecium-specific features.

2. Material and Methods

Three species were selected for this study: *Clivia miniata* (Lindl.) Bosse, *Clivia nobilis* Lindl., and *Scadoxus multiflorus* (Martyn) Raf. (Figure 1). Buds and flowers were



Figure 1 Flowering scapes of *Clivia miniata* (Lindl.) Bosse. (A), *Clivia nobilis* Lindl. (B), and *Scadoxus multiflorus* (Martyn) Raf. (C).

sampled in the A. V. Fomin Botanical Garden of Taras Shevchenko National University of Kyiv. The samples were fixed in 70% alcohol in April 2020. Five flower buds of each species that were 1.0 to 1.7 cm in length were dehydrated in an increasing series of t-butanol (20%, 30%, 50%, 70%, 100%; 2 hr each for 20%–70%, 24 hr for 100%). Each sample was stored in a 1:1 preparation of 100% t-butanol and Paraplast. Infiltration with Paraplast (Merck; New York, NY, USA) was performed as previously described (Baryikina et al., 2004). Transverse sections 20 µm in thickness were obtained with an MPS-2 manual rotary microtome (Tochmedpribor; Kharkiv, USSR) using an existing staining protocol (Soukup & Tylová, 2019) with safranin (Sigma-Aldrich; St. Louis, MO, USA) and Astra Blue (Merck). Slides were mounted in Eukitt (Sigma-Aldrich). Light microscopy was used, and images with 3,584 × 2,748 resolution saved as .jpg files were obtained with a 10MP digital camera attached to a modelT490B-10M microscope (AmScope; https://amscope.com/). Photopea software (https://www.photopea.com/) was used for image processing.

For morphological analysis, measurements were performed on at least 15 fresh flowers. To analyze the internal structure of the gynoecium, we used Leinfellner's (1950) concept of vertical zonation. According to this concept, the congenital multilocular synascidiate, unilocular symplicate, transitional hemisymplicate, and asymplicate (apocarpous) zones can be formed in the syncarpous gynoecium. In the conditions of incomplete fusion of carpels only in their outer part, a hemisyncarpous gynoecium with hemisynascidiate, hemisymplicate, and asymplicate zones arises. This concept was subsequently developed for monocots with epigynous and hypogynous flowers regarding the septal nectary structure (Novikoff & Odintsova, 2008; Odintsova, 2013). The height of the gynoecium zones was measured according to the number of cross-sections.

3. Results

3.1. Clivia miniata

The flowers of C. miniata were 8.3 cm long (Figure 2B). The scape was 35–40 cm in diameter, 1.5 cm in diameter at the base, and 1.3 cm above. There were 10–13 flowers in an inflorescence. Bracts were cone-shaped, 3.2-3.6 cm long, 1.7-2.0 cm wide, leathery, and light brown. The pedicel was 3.2 cm long and 0.4 cm in diameter. The perigonium was slightly zygomorphic and bright orange. The tepals were fused into a short floral tube, 0.7 cm long and 0.7 cm in diameter. The outer tepals were 6.0-6.2 cm long and 1.9-2.1 cm wide. The tepals of the inner whorl were 6.4-6.5 cm long and 2.7-2.9 cm wide (Figure 2A). The stamens were adnate to the floral tube at the base (Figure 2A). The length of the stamen filaments of the outer and inner whorl was 4.1-4.2 cm and 4.6 cm, respectively. The filaments became narrower at the top. Anthers were introse linear, 0.4 cm long and 0.1 cm wide, and were connected to filament below the middle (Figure 2D). The style was slightly zygomorphic. The inferior ovary was obovate, 1.4 cm high, and 0.7 cm in diameter (Figure 3A). The lower part of the ovary was green, and the upper part was yellow (Figure 2C). The style was located in the center of the ovary. The style was filamentous, curved, 5.8 cm long, 0.2 cm in diameter at the base, and yellow. The stigma was light yellow and three-lobed with short lobes.

The ovary wall was thick (approximately 0.2 cm), as well as the ovary base and ovary roof, with five-to-six ovules in a locule (Figure 3A). In the *C. miniata* ovary, the following structural zones could be discerned (Table 1): a fertile synascidiate structural zone, fertile symplicate structural zone, and hemisymplicate zone (Figure 4). In the subapical portion, the locules became gradually rounded in cross section. They merged into a single channel within the ovary roof, which entered the center of the style as a style channel (Figure 5F). Septal nectaries appeared at the level of the hemisymplicate zone (Figure 5E, Figure 6B) and opened with slits that had no secretory tissue at the base of the style (Figure 6D). The cavities of the septal nectaries were oval in cross section and characterized by the absence of secretory tissue in the upper part. The asymplicate zone formed a style above the opening of the septal nectary (Figure 6D).



Figure 2 Anthetic flowers and flower details in members of the tribe Haemantheae.
(A-D) *Clivia miniata*. (A) Flower, general view; (B) pistil; (C) series of tepals with connected stamens. (D) Preparated perigonium and androecium. (E,F) *Clivia nobilis*.
(E) Flower; (F) pistil. (G-I) *Scadoxus multiflorus*. (G) Flower in the longitudinal section at late anthesis. (H) Fragment of the floral tube with stamens. (I) Pistil. Scale bars: 10 mm.

The anatomical characteristics of the studied species are presented in Table 1. All floral parts displayed a parenchymatous, homogenous histological structure, with the presence of idioblasts with raphides in the peduncle and ovary wall (Table 1). The inner epidermis of the ovary comprised small cells (Figure 5D).

The peduncle of *C. miniata* contained six large vascular bundles and six small vascular bundles (Figure 5A), from which three septal veins and three dorsal veins formed at the ovary base (Figure 5B). The other vascular bundles were concentrated closer to the center of the ovary. Every dorsal (Figure 5D) and septal vein (Figure 6A) became three-bundled and produced small horizontal branches in the ovary wall. In the upper part of the ovary, the small bundles branched from the dorsal vascular bundles, which extended into the inner layer of the ovary wall (Figure 5E).





In the central column of the ovary, on the septum radius, a massive vascular bundle comprised the roots of the ventral complex (Figure 5C). Distally, each root of the ventral complex is divided into two bundles (ventral bundles of the carpel) that extended to the ovules. The small branches that supply the septal nectaries formed from the ventral bundles. Above the locules, the ventral bundles of the carpel, as well as the septal bundles and other small veins, fused with the dorsal bundles and entered the style. In the floral tube, the dorsal and septal veins were divided, and their branches entered the stamen and tepal traces (Figure 5F). Tepal traces branched higher in the floral tube, initially into three bundles and, higher, into a larger number of bundles (Table 1). The stamen traces were two-bundled.

3.2. Clivia nobilis

The flowers of *C. nobilis* were 6.9 cm long and 1.0–1.1 cm in diameter (Figure 1B). The scape was 36–38 cm, 0.9 cm at the base, with 15–17 flowers in the inflorescence. The pedicel was 0.9–1.0 cm long and 0.2 cm in diameter. The perigonium was slightly zygomorphic and bright red-yellow with green tips of the tepals (Figure 2F). The tepals fused into a short floral tube (Figure 7F,G) that was 1.3 cm long and 0.8 cm in diameter. The outer tepals were 4.1–4.6 cm long and 0.9 cm wide. The inner tepals were 4.4 cm long and 1.4–1.5 cm wide. The stamens were adnate to the floral tube. The length of the stamen filaments of the outer whorl was 3.3–3.5 cm, and the stamen filaments of the inner whorl were 3.7–3.8 cm. The filaments were 0.1 cm in diameter in the outer stamens and 0.15 cm in the inner stamens and narrowed towards the top. The anthers were linear, 0.5 cm long, and 0.1 cm wide and connected to a filament below the middle (Figure 7H).

The gynoecium was slightly zygomorphic (Figure 2F). The ovary was obovate, 0.6 cm in diameter, and 0.8–1.0 cm in height (Figure 3B). The lower part of the ovary was green, and the upper part was yellow. The style was located in the center of the ovary. The style was filamentous, yellow, curved in the middle, 5.6 cm long, 0.1 cm in diameter at the base. The stigma was light yellow and three-lobed. There were six-to-eight ovules per locule (Figure 3B, Figure 7C, Table 1). The *C. nobilis* gynoecium contained the same structural zones as in *C. miniata*. They were the same height, except for the symplicate zone (Figure 4), which is 3.5 times shorter (Table 1). The septal nectary (Figure 7D, Figure 8B) was similarly located in the hemisymplicate region of the ovary (Figure 4). The style channels formed apically

Studied traits	Clivia miniata	Clivia nobilis	Scadoxus multiflorus
Morphological traits of flower			
Flower length (cm)	8.3 ± 0.188	6.9 ± 0.155	4.3 ± 0.112
Floral tube length (cm)	0.7 ± 0.058	1.3 ± 0.079	1 ± 0.081
Morphological traits of anthesis ovary			
Height (cm)	1.4 ± 0.146	0.8 ± 0.169	0.3 ± 0.096
Diameter (cm)	0.7 ± 0.131	0.6 ± 0.119	0.25 ± 0.044
Micromorphological traits of flower buds			
Ovary base length (μm)	640 ± 13.601	620 ± 7.906	240 ± 10.770
Synascidiate zone length (µm)	$1,440 \pm 16.808$	$1,240 \pm 13.342$	260 ± 8.544
Symplicate zone length (µm)	$2,380 \pm 14.983$	680 ± 10.849	80 ± 9.487
Hemisymplicate zone length (µm)	$1,620 \pm 18.668$	$1,700 \pm 17.219$	400 ± 6.978
Ovary roof length (µm)	240 ± 12.021	300 ± 8.944	340 ± 8.860
Septal nectary length (µm)	$1,860 \pm 13.602$	$2,000 \pm 13.342$	740 ± 11.511
Septal nectary proportion in ovary (%)	29	44	56
Ovule number (per locule)	4-6	6–9	1
Ovules insertion in the synascidiate /	+/+	+/+	+/
Anatomical traits			
Raphides	Rare, in the	Rare, in the	Abundant in the peduncle.
	peduncle,	peduncle,	ovary wall, base of floral
	ovary wall	ovary wall	tube, free tepals
Number of cell layers in the ovary wall (approximately)	27	25	12
Outer tepal trace (bundle)	10	9	3
Inner tepal trace (bundle)	8	7	3
Stamen trace (bundle)	2	1	1
Ovule trace (bundle)	1		2

Table 1 Micromorphological and anatomical characters of the studied species of Haemantheae (30 individual plants).

(Figure 7E,F, Figure 8D). There were idioblasts with raphides in the peduncle and ovary wall in *C. nobilis* (Table 1).

The structure of the vascular system of the flower was similar to that of *C. miniata*, with some minor differences (Table 1). The *C. nobilis* peduncle contained 15 vascular bundles (Figure 7A). At the level of the ovary base, septal bundles (Figure 8C) and dorsal bundles departed (Figure 8A). Small vascular bundles were located proximally to them (Figure 7B–D). In the floral tube, the dorsal and septal veins were divided and their branches entered the stamen and tepal traces (Figure 7F). Small bundles located inside the ring of dorsal and septal bundles anastomosed with each other and formed roots of the ventral complex. The bundles of the ventral complex branched into paired ventral bundles of carpels, supplying ovules and septal nectaries.

3.3. Scadoxus multiflorus

The flower of *S. multiflorus* was up to 4.2–4.5 cm long (Figure 1C, Figure 2G,H). The scape was 35–41 cm long and 0.2 cm in diameter. The five bracts were 5.2–5.8 cm long, 1.0–1.6 cm wide, and narrowed at the base. The pedicel was 3.8–4.5 cm long and 0.2 cm in diameter. The perigonium formed a floral tube 0.9–1.1 cm long and 0.3 cm in diameter (Figure 2G). The outer tepals were 2.6 cm long and 0.2 cm wide. The inner tepals were 2.3 cm long and 0.1 cm wide. The floral tubes and tepals were bright red (Figure 2H). The stamens were adnate to the floral tube at the base (Figure 2H). The stamens of the outer whorl were slightly longer than those of the inner whorl. The filaments were 0.2 cm long. On the inner stamens, anthers were 0.15 cm long and 0.05 cm in diameter. The anthers were attached to the filament below the middle (Figure 9H). The ovary was oval, light green, 0.3 cm in





height, and 0.25 cm in diameter, with a single ovule in each locule (Figure 3C). The style was filamentous (Figure 2I), white at the base, red at the top, had a central location, and 3.6 cm high. The stigma was capitate.

The ovary had the same vertical zones, and the septal nectary was located in the hemisymplicate zone. The ovary base and roof were slightly thickened, but the ovary wall was thinner than that in *Clivia* species (Figure 3, Figure 9E,F, Figure 10A,C, Table 1). The septal nectaries were very narrow and barely visible in the cross section (Figure 9G, Figure 10B). The presence of a large number of idioblasts with raphides in all organs of the flower and stomata on the tepals was peculiar to the species (Figure 10C,D, Table 1).

In the peduncle of *S. multiflorus*, only three vascular bundles were available at the base (Figure 9A,B), which branched distally into three bundles each. At the level of the ovary base, nine vascular bundles were present (Figure 9C), from which septal bundles, dorsal bundles, and small vascular bundles were proximal to septal bundles and dorsal bundles departed (Figure 9D). Higher in the ovary in each septum, small vascular bundles were present as the roots of a ventral complex (Figure 9D). They formed paired ventral vascular bundles (Figure 9E,F). Each ovule was supplied by two bundles extending from the ventral bundles of the carpels. The dorsal veins of carpels were two-bundled (Figure 10A), and septal veins were single-bundled.

4. Discussion

The flowers of the studied species have many common features, including a large size, slightly zygomorphic, short-tubular, and red-orange color, with the same set of organs. However, the functional type of flower in *Clivia* species is tubular, broadly tubular to disc-shaped; in *S. multiflorus*, the attractiveness is ensured by narrow, linear tepals, and colored elongated filaments (Leins & Erbar, 2010).

Understanding the evolution of flower morphology requires information about the identity of pollinators and knowledge about specific mechanisms of pollen transfer (e.g., Bożek, 2019). Based on preliminary field observations and flower structure,



Figure 5 Ascending series of transversal sections of the *Clivia miniata* ovary. (**A**,**B**) Peduncle; (**C**–**E**) ovary; (**F**) ovary roof. dv – dorsal vein; lo – ovary locule; ov – ovule; rvc – roots of the ventral complex; sc – style channel; sn – septal nectary; str – stamen trace; sv – septal vein; vb – vascular bundle. (**C**) Synascidiate structural zone. (**D**) Symplicate structural zone. (**E**) Hemisymplicate structural zone. Scale bars: 500 μ m.

Butler and Johnson (2020) hypothesized that pollination mechanisms involving the transfer of pollen on butterfly wings occur in several genera of South African Amaryllidaceae, including *S. multiflorus*.

In the gynoecium of the studied species, we found the same vertical zonality, characteristic of eusyncarpous gynoecium, that is, the presence of synascidiate, symplicate, hemisymplicate, and asymplicate zones. The synascidiate zone was well developed in the studied species, with a central column and placenta. The symplicate zone was longer or shorter and contained the placenta in the *Clivia* species.



Figure 6 Details of the gynoecium structure of *Clivia miniate*. (A) Ovary wall with septa attached; (B) the central part of the ovary with three septal nectaries; (C) ovary wall in the median plane; (D) style base with triradial style channel. dv - dorsal vein; lo – ovary locule; sc – style channel; se – septa; sn – septal nectary; sv – septal vein. Scale bars: 250 μ m.

The hemisymplicate zone in the studied species was sterile, occupying the upper half of the ovary. The main function of this zone was the formation of septal nectaries. In *S. multiflorus*, the size of all zones of the ovary was the smallest, as was the size of the whole ovary. However, in this species, the roof of the ovary was the thickest. Thus, in the studied species, the seed-bearing and nectar-bearing parts of the ovary were separated in space. In *C. miniata* and *S. multiflorus*, the style was slightly longer than that of the stamens, which provides approach herkogamy. In *Clivia*, the style also curved. *Clivia* flowers have nectary guides on the tepals, and the pollinator is rewarded with nectar from the septal nectaries. We have previously studied synascidiate and symplicate zones in *Galanthus nivalis* and *Leucojum vernum* gynoecium (Fishchuk & Odintsova, 2020). We found that in the *Hippeastrum striatum* gynoecium presented synascidiate, symplicate, hemisymplicate, and asymplicate vertical zones (Fishchuk, 2021).

As revealed in this study, the septal nectary opened at the base of the style so that the style formed by the postgenital fusion of the carpels were above the output of the nectary slits. The nectary is represented by three narrow cavities, which connect in the center of the ovary by a common epidermis, although they are postgenitally closed. The septal nectary in the studied species had different lengths. It was shortest in *S. multiflorus* and approximately the same in *C. miniata* and *C. nobilis*. The septal nectary portion in the ovary was the largest in *S. multiflorus*, where it was more than half the height of the ovary. This nectary structure corresponds to the "lilioid" distinct nonlabyrinthine type, according to Schmid (1985). This structure of the septal nectary is characteristic of other species of the order Asparagales, which have an inferior ovary and septal nectary, such as *Gladiolus hybridus* from Iridaceae



Figure 7 Ascending series of transversal sections of the flower details of *Clivia nobilis*. (A) Peduncle; (B–D) ovary; (E,F) ovary roof; (G) floral tube and style; (H) anther in the longitudinal section. an – anther; dv – dorsal vein; fi – filament; ft – floral tube; lo – ovary locule; ov – ovule; sc – style channel; sn – septal nectary; st – style; str – stamen traces; tt – tepal traces; sv – seplal vein; vb – vascular bundle; vv – ventral bundle. Scale bars: 500 μ m.

(Skrypets' & Odintsova, 2013). The nectaries are in the form of a nectary disk located between the stamens and the style of *G. nivalis*. The septal nectary sutures are absent, but in the center of the ovary in the simplicate zone, a triradial cavity forms between the ends of incomplete septa in *Leucojum vernum*. (Fishchuk & Odintsova, 2020). *Hippeastrum striatum* has internal septal nectaries that appear at the ovary base and are continued by nectar fissures (Daumann, 1970). In the genus *Allium* (Amaryllidaceae), nectar is secreted at the top of the ovary, where there is a greater risk of theft and less contact with the fertile part by pollinating visitors. Gynopleural nectaries are also absent from some genera of Amaryllidaceae. In *Sternbergia*





(Amaryllidaceae), tepalar and staminal nectaries occur in addition to gynopleural nectaries (Smets et al., 2000).

According to Daumann (1970), the genera *Clivia* and *Scadoxus* have an internal septal nectary with slits in the ovary septa, characterized by the absence of vascular bundles that supply the nectary. Nectaries of the genus *Clivia* are composed of three septal nectary slits, which mostly extend above the locules, which appear only in the upper two thirds of the ovary height. The cells of the nectary epidermis are significantly elongated (in the form of a palisade). Nectary excretion occurs only in the lower two thirds of the septum, where the cells of the epidermis of the nectary are elongated. Towards the style, they gradually become shorter. At these points, nectary secretion becomes weaker and finally stops completely, so that (although without a sharp limit) approximately the upper one third of the nectary slit provides only the exit channel for nectary excretion, released by the lower parts of the nectary. Prominent groups of vascular bundles pass close to the nectary parenchyma but provide no branches into the nectary tissue (Daumann, 1970).

Examining other members of the order Asparagales, we found similar features of thickened ovary base and ovary roof, and the presence of all zones in the gynoecium in the family Asparagaceae s. l. (Fishchuk & Odintsova, 2013, 2014; Odintsova & Fishchuk, 2017). Most of the studied species also have a septal nectary, but they are usually prolonged from the ovary base to the ovary roof.

Despite the obvious adaptations to the attraction of pollinators, a low number of ovules are formed in the ovaries of the studied species, which is characteristic of



Figure 9 Ascending series of transversal sections of *Scadoxus multiflorus* flower. (A–C) Peduncle; (D–F) ovary; (G) ovary roof; (H) floral details at anther level. an – anther; dv – dorsal vein; lo – ovary locule; ov – ovule; sc – style channel; sn – septal nectary; st – style; te – tepal; sv – septal vein; vv – ventral vascular bundles; vb – vascular bundle. Scale bars: 500 μ m.

indehiscent fruits. A solitary ovule occurs in each locule of *S. multiflorus*. In the *H. striatum* ovary, there are 16–20 ovules in each locule (Fishchuk, 2021), in *G. nivalis* there are 14 ovules, and in *L. vernum* there are 12–14 ovules in each locule (Fishchuk & Odintsova, 2020). The other traits, which are distinct in *S. multiflorus* compared with *Clivia* species, include abundant raphides in floral parts, three bundles in tepal traces and two bundles in ovule trace, and fewer cell layers in the ovary wall. Idioblasts with raphides are present in *H. striatum* in the upper part of the peduncle, receptacle, stamen filaments, ovary wall, and free tepals (Figure 4). They are absent in the connective style (Fishchuk, 2021). The upper part





of the pedicel, base of the floral tube, stamen filaments, connectives, the wall of the ovary, and the style contain idioblasts with raphides in *G. nivalis*, and in the free apices of the tepals, they are absent. At the base of the floral tube, the stamen filaments, and the wall of the ovary, free apices of the tepals contain idioblasts with raphides in *L. vernum*. They are absent in the apical part of the pedicel and the style (Fishchuk & Odintsova, 2020). In plant cells, idioblasts with raphides can perform protective, supportive, and spare functions.

Comparing the structure of the flower with representatives of the tribe Galantheae (Amaryllidaceae) – *G. nivalis* and *L. vernum* (Fishchuk & Odintsova, 2020) – several differences were evident. In Haemantheae, anthers are attached to the floral tube and open along their entire length, a septal nectary is present, there is no aerenchyma in the ovary wall and other parts of the flower, and there are few ovules. In the tribe Galantheae, dehiscence of the anthers through the apical pores in *Galanthus* and *Leucojum* could be considered an extremely specialized variant of lengthwise splitting of short sutures in the apical part of the anthers, which is possibly attributed to the pendant position of flowers (Fishchuk & Odintsova, 2020). Common features are thick fleshy ovary walls and septa, multibundle traces of tepals, and the presence of synascidiate and symplicate zones.

The morphological and anatomical features of the flower in Asparagales are being studied to explain the appearance of some features of the flower and fruit (Odintsova

& Fishchuk, 2017; Skrypec & Odintsova, 2020). One such feature is the organization of the flower vascular system. In the studied species, a large number of small vascular bundles were observed in the inferior ovary wall, which is an adaptation to the formation of fleshy fruit. The other trait characteristic of the fleshy fruit is a thickened ovary wall, base, and roof. The flower vascular system is characterized by three-bundled dorsal and septal veins and the presence of branches of the dorsal and septal veins in the ovary in *C. miniata* and *S. multiflorus*. In *S. multiflorus*, the dorsal bundles are doubled, and the septal bundles are single-bundled. In *C. nobilis*, the dorsal and septal bundles are three-bundled but have no branches. The present study of morphological and anatomical features of the flower will be useful in the taxonomical revision of the family Amaryllidaceae.

5. Conclusions

We studied the micromorphology and anatomy of the flower *C. miniata* (Lindl.) Bosse, *C. nobilis* Lindl., and *S. multiflorus* (Martyn) Raf. The examination revealed common features in the gynoecium micromorphology and vascular anatomy of these species. Synascidiate, symplicate, and hemisymplicate zones were found in the inferior ovary, while the style was formed by an asymplicate zone. Ovules were located in the synascidiate and symplicate zones in *C. miniata*, *C. nobilis*, and in the synascidiate zone of *S. multiflorus*. The septal nectary is located in the hemisymplicate zone in the upper part of the ovary. The vascular system in the inferior ovary wall consisted of a circle of ascending bundles on the tepal radii, from which traces of stamens and tepals were formed. In the inner layer of the ovary wall, dorsal and septal veins were anastomosed with each other. Traces of ovules were formed from the ventral bundles of the carpel in the central column of the ovary. The studied species showed great similarity in their micromorphology and vascular anatomy.

References

- The Angiosperm Phylogeny Group. (2016). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Botanical Journal of the Linnean Society*, *181*, 1–20. https://doi.org/10.1111/boj.12385
- Барыкина [Baryikina], Р. П. [R. Р.], Веселова [Veselova], Т. Д. [Т. D.], Девятов [Deviatov], A. Г. [A. G.], Джалилова [Dzhalilova], Х. Х. [Kh. Kh.], Ильина [Il'ina], Г. М. [G. М.], & Чубатова [Chubatova], Н. В. [N. V.]. (2004). Справочник по ботанической микротехнике. Основы и методы [Handbook of the botanical microtechniques]. Издательство Московского университета [Moscow University Publishing House].
- Bożek, M. (2019). Nectar secretion and pollen production in *Hyacinthus orientalis* 'Sky Jacket' (Asparagaceae). *Acta Agrobotanica*, 72(4), Article 1796. https://doi.org/10.5586/aa.1796
- Butler, H. C., & Johnson, S. D. (2020). Butterfly-wing pollination in *Scadoxus* and other South African Amaryllidaceae. *Botanical Journal of the Linnean Society*, 193(3), 363–374. https://doi.org/10.1093/botlinnean/boaa016
- Daumann, E. (1970). Das Blütennektarium der Monocotyledonen unter besonderer Berücksichtigung seiner systematischen und phylogenetischen Bedeutung [The floral nectarium of the monocotyledons with special consideration of its systematic and phylogenetic importance].

Feddes Repertorium, 80(7–8), 463–590. https://doi.org/10.1002/fedr.19700800702 Duncan, G. (2017). *The Amaryllidaceae of Southern Africa*. Tien Wah Press.

- Fishchuk, O. (2021). Comparative flower morphology in *Hippeastrum striatum* (Lam.) H. E. Moore. (Amaryllidaceae). Ukrainian Journal of Ecology, 11(1), 273–278.
- Фіщук [Fishchuk], О. С. [O. S.], & Одінцова [Odintsova], А. В. [A. V.]. (2013). Морфологія та васкулярна анатомія квітки *Sansevieria suffruticosa* N. Е. Br. (Asparagaceae Juss.) [Morphology and vascular anatomy of the flower in *Sansevieria suffruticosa* N. E. Br. (Asparagaceae Juss.)]. *Біологічні студії* [Studia Biologica], 7(1), 139–148. https://doi.org/10.30970/sbi.0701.260
- Фіщук [Fishchuk], O. C. [O. S.], & Одінцова [Odintsova], A. B. [A. V.]. (2014). Вертикальна зональність гінецею у представників роду *Sansevieria* Thunb. та *Dracaena* Vand. ex L. species (Asparagaceae) [Vertical zonality of the gynoecium in *Sansevieria* Thunb. and *Dracaena* Vand. ex L. species (Asparagaceae)]. *Сучасна фітоморфологія* [Modern Phytomorphology], 5, 221–226. https://doi.org/10.5281/zenodo.161028

- Fishchuk, O. S., & Odintsova, A. V. (2020). Micromorphology and anatomy of the flowers of Galanthus nivalis and Leucojum vernum (Amaryllidaceae). Regulatory Mechanisms in Biosystems, 11(3), 463–468. https://doi.org/10.15421/022071
- Funnell, K. A. (2008). Growing degree-day requirements for scheduling flowering of Scadoxus multiflorus subsp. katharinae (Baker) Friis & Nordal. HortScience, 43(1), 166–169. https://doi.org/10.21273/HORTSCI.43.1.166
- Kiepiel, I., & Johnson, S. D. (2019). Spit it out: Monkeys disperse the unorthodox and toxic seeds of *Clivia miniata* (Amaryllidaceae). *Biotropica*, 51, 619–625. https://doi.org/10.1111/btp.12698
- Kwembeya, E. G. (2021). Tracking biological footprints of climate change using flowering phenology of the geophytes: *Pancratium tenuifolium* and *Scadoxus multiflorus*. *International Journal of Biometeorology*, 65, 577–586. https://doi.org/10.1007/s00484-020-02052-2
- Leinfellner, W. (1950). Der Bauplan des synkarpen Gynözeums [The structure of the syncarpous gynoecium]. Österreichische Botanische Zeitschrift, 97, 403–436. https://doi.org/10.1007/BF01763317
- Leins, P., & Erbar, C. (2010). Flower and fruit. Morphology, onthogeny, phylogeny, function and ecology. Schweizerbart'Sche Verlagsbuchhandlung.
- Li, Y. L., Yan, Z. B., Wang, Y. H., & Zhou, Z. (2020). First report of *Fusarium* proliferatum causing leaf sheath rot on *Clivia miniata* in Henan Province, China. *Plant Disease*, 104(5), Article 1552. https://doi.org/10.1094/PDIS-12-18-2276-PDN
- Liu, L., Li, X., Jiang, J., Nie, Q., Zheng, T., Wan, W., Hsiang, T., Sun, Z., & Zhou, Y. (2020). First report of *Fusarium oxysporum* causing basal stem rot of *Clivia miniata* in China. *Plant Disease*, 104(5), Article 1561. https://doi.org/10.1094/PDIS-10-19-2099-PDN
- Meerow, A. W., & Clayton, J. (2004). Generic relationships among the baccate-fruited Amaryllidaceae (tribe Haemantheae) inferred from plastid and nuclear non-coding DNA sequences. *Plant Systematics and Evolution*, 244, 141–155. https://doi.org/10.1007/s00606-003-0085-z
- Meerow, A. W., Francisco-Ortega, J., & Schnell, R. J. (2006). Phylogenetic relationships and biogeography within the Eurasian clade of Amaryllidaceae based on plastid *ndhF* and nrDNA ITS sequences: Lineage sorting in a reticulate area? *Systematic Botany*, *31*(1), 42–60. https://doi.org/10.1600/036364406775971787
- Meerow, A. W., & Snijman, D. A. (1998). Amaryllidaceae. In K. Kubitzki (Ed.), Flowering plants: Monocotyledons (pp. 83–110). Springer. https://doi.org/10.1007/978-3-662-03533-7_11
- Mertens, J. E. J., Janeček, Š., Dzekashu, F. F., Chmelová, E., Fokam, E. B., & Tropek, R. (2020). Changes in pollinator community of *Scadoxus cinnabarinus* (Amaryllidaceae) along its elevational range on Mount Cameroon. *Arthropod–Plant Interactions*, 14, 215–226. https://doi.org/10.1007/s11829-020-09741-0
- Monkheang, P., Chaveerach, A., Sudmoon, R., & Tanee, T. (2016). Karyotypic features including organizations of the 5S, 45S rDNA loci and telomeres of *Scadoxus multiflorus* (Amaryllidaceae). *Comparative Cytogenetics*, 10(4), 637–646. https://doi.org/10.3897/CompCytogen.v10i4.9958
- Novikoff, A., & Odintsova, A. (2008). Some aspects of gynoecium morphology in three bromeliad species. Wulfenia, 15, 13–24.
- Одінцова [Odintsova], А. В. [A. V.]. (2013). Два основних типи септальних нектарників однодольних [Two main types of septal nectaries in monocotyledons]. Вісник Львівського університету, Серія біологічнаю [Visnyk of the Lviv University, Series Biology], 61, 41–50.
- Odintsova, A., & Fishchuk, O. (2017). The flower morphology in three Convallariaceae species with various attractive traits. *Acta Agrobotanica*, 70(1), Article 1705. https://doi.org/10.5586/aa.1705
- Petravich, A., Harbage, J. F., & Taylor, M. (2015). Clivia miniata 'Longwood Debutante,' Clivia miniata 'Longwood Fireworks,' and Clivia miniata 'Longwood Sunrise'. HortScience, 50(7), 1092–1095. https://doi.org/10.21273/HORTSCI.50.7.1092
- Qin-Mei, W., Jianguo, C., Hongyan, D., Yongbin, Z., Na, L., & Zhihong, Z. (2018). Comparative transcriptome profiling of genes and pathways involved in leaf-patterning of *Clivia miniata* var. *variegata*. *Gene*, 677, 280–288. https://doi.org/10.1016/j.gene.2018.07.075
- Remizova, M. V., Sokoloff, D. D., & Rudall, P. J. (2010). Evolutionary history of the monocot flower. Annals of the Missouri Botanical Garden, 97, 617–645. https://doi.org/10.3417/2009142
- Rudall, P. J. (2002). Homologies of inferior ovaries and septal nectaries in monocotyledons. International Journal of Plant Sciences, 163, 261–276. https://doi.org/10.1086/338323

- Schmid, R. (1985). Functional interpretations of the morphology and anatomy of septal nectaries. Acta Botanica Neerlandica, 4(1), 125–128. https://doi.org/10.1111/j.1438-8677.1985.tb01862.x
- Smets, E. F., Ronse de Craene, L. P., Caris, P., & Rudall, P. J. (2000). Floral nectaries in monocotyledons: distribution and evolution. In K. L. Wilson & D. A. Morrison (Eds.), *Monocots: Systematics and evolution* (pp. 230–240). CSIRO.
- Skrypec, K., & Odintsova, A. (2020). Morphogenesis of fruits in *Gladiolus imbricatus* and *Iris sibirica* (Iridaceae). Ukrainian Botanical Journal, 77(3), 210–224. https://doi.org/10.15407/ukrbotj77.03.210
- Скрипець [Skrypets'], Х. І. [С. І.], & Одінцова [Odintsova], А. В. [А. V.]. (2013). Морфологія та васкулярна анатомія гінецея *Gladiolus* ×*hybridus* С. Morren hort. (Iridaceae Juss.) [Gynoecium morphology and vascular anatomy in *Gladiolus* ×*hybridus* С. Morren hort. (Iridaceae Juss.)]. *Сучасна фітоморфологія* [Modern Phytomorphology], 3, 241–244. https://doi.org/10.5281/zenodo.162026
- Soukup, A., & Tylová, E. (2019). Essential methods of plant sample preparation for light microscopy. In F. Cvrčková & V. Žárský (Eds.), *Plant cell morphogenesis* (pp. 1–26). Humana. https://doi.org/10.1007/978-1-4939-9469-4_1
- Stevens, P. F. (2001). Angiosperm Phylogeny Website, version 14, July 2017 (and more or less continuously updated since). http://www.mobot.org/MOBOT/research/APweb/
- Takhtajan, A. (2009). *Flowering plants*. Springer. https://doi.org/10.1007/978-1-4020-9609-9
- Vendrame, W. A., Garofalo, J. F., & Meerow, A. W. (2004). Effects of light duration on flower development in blood lily. *Proceedings of the Florida State Horticultural Society*, 117, 341–345.
- Wang, Q. M., Wang, L., Zhou, Y., Cui, J., Wang, Y., & Zhao, C. (2016). Leaf patterning of *Clivia miniata* var. *variegata* is associated with differential DNA methylation. *Plant Cell Reports*, 35(1), 167–184. https://doi.org/10.1007/s00299-015-1877-7