The present study investigated the effect of different soil moisture content levels (60 – 70% SWC (soil water capacity) – control; 30 – 35% SWC – water stress) on yields, gas exchange parameters, seed health, and protein fractions of husked oat grain. The study showed that water deficit resulted in a decrease in grain weight per plant and a reduction in the gas exchange rates, primarily the photosynthesis and transpiration rates. 

Cladosporium cladosporioides was the dominant species on oat kernels in both experimental treatment options and in both years of the study. The presence of Fusarium poae was also found. Higher contents of prolamin, albumin and globulin fractions were found in the oat grain harvested from plants grown under soil water deficit conditions.

Key words: oats, yield, gas exchange rates, seed health, protein fractions

INTRODUCTION

Oat is a cereal grain of high dietary value. In its composition, it contains large amounts of good quality protein, fat rich in unsaturated fatty acids, dietary fibre rich in water-soluble β-glucans, tocols, phenolic compounds with antioxidant properties, and many other substances that may have a beneficial effect on the functioning of the organism (Bartnikowski et al. 2002B). The high nutritional value of protein in oat grain has been used to develop the production technology for protein concentrates and isolates in the manufacture of meat and bakery products (Bartnikowski et al. 2000A). This is why human nutrition experts have suggested for many years that the proportion of the so-called non-bread cereals should be increased in daily dietary intake, particularly oats and barley (Gąsiorowski and Kawka, 1995).

Grain for human consumption should be characterized by high quality and be free from fungal pathogens which may produce mycotoxins dangerous to humans and animals. The species of the genus Fusarium have a major role here, since in cereal grains they most frequently produce trichothecenes from the group of sesquiterpenoids (Nicholson et al. 2003; Nicholson et al. 2004).

Grain yield and quality are dependent on both habitat conditions and agricultural practices (Wróbel and Kijora, 2004). Oats are generally considered to be plants with low thermal requirements and high water requirements, mainly during the period from stem elongation to panicle emergence (Panek, 1992).

Water deficit is one of the principal factors that reduce crop productivity (Grzesiuk et al. 1999; Starck et al. 1995). Plants respond differently to water deficiency in different periods of their growth. The generative phase and the beginning of flowering are most frequently the period of the greatest sensitivity to water deficit (Grzesiuk et al. 1999, Skrabka 1992). Furthermore, water deficit leads to the disturbance of the basic physiological processes, chiefly photosynthesis. On the one hand, this is caused by limited access of CO₂ from the outside air as a result of the decreased permeability of stomata and, on the other hand, by the reduced activity of the enzymes participating in photosynthesis (Skrabka, 1992).

If oat grain is intended for human consumption, not only its health is of essential significance, but also its technological value, in particular the composition of proteins. In quantitative terms, globulins make up the
largest percentage of proteins in oat kernels, from 50 to 80%, while prolamin and glutenin fractions account for the remaining part (more than 20% of total protein) (Gasiorek, 1995; Bartnikowska et al. 2000A). But there is no gluten in oat kernels, which excludes oat grain as raw material used for bread baking (Sulek et al. 2005). In connection with the above, oat grain may and should be used in the diet of people suffering from coeliac disease.

The aim of the present study was to determine plant productivity, seed health, and protein fractions of oats grown under different soil moisture conditions.

**MATERIALS AND METHODS**

In the period 2004–2005, a single-factor pot experiment was carried out in two experimental series, in quadruplicate, in a greenhouse of the University of Warmia and Mazury in Olsztyn. The experimental factor was varying soil moisture content: 60 – 70% SWC (soil water capacity) – control; 30 – 35% SWC – water stress; such conditions were maintained from the stage of grain development (BBCH 75). Oats were grown in Kick-Brauckmann pots. Before sowing, the pots were filled with typical brown soil with the following nutrient availability: P2O5 – 6.41 mg ‰, K2O – 1.30 mg ‰, Mg – 190 mg ‰ kg-1, and Ca – 218 TPPS, 5 mol/L NaCl + 0.067 mol/L HKNaPO4 with a pH of 7.6.

To determine the content of particular protein fractions, a 3 g grain sample was ground in an IKA A10 laboratory mill (Labortechnik) in such a manner so that all particles could be sieved through a 400 µm mesh sieve (ether particles smaller than 250 µm accounted for 90%). The samples were degreased with petroleum ether in Soxhlet extractors (16 hours). After evaporation of the solvent, 100 mg portions of powder were weighed out and placed in Eppendorf tubes, and then three protein fractions were extracted according to Wieser et al. (1998).

1) albumins + globulins – triple extraction of 1 cm³ of the mixture (0.4 mol L-1 NaCl + 0.067 mol L-1 NaCl + 0.067 mol L-1 NaCl + 0.006 mol L-1 HKNaPO4 with a pH of 7.6
2) prolamins – triple extraction of 1 cm³ of the mixture (60% ethanol)
3) glutenins – double extraction of 1 cm³ of the mixture (50% propanol + 2 mol/L urea 0.05 mol L-1 with a pH of 7.5) + 1% DTE under nitrogen.

The first two protein fractions were extracted at room temperature using an Eppendorf thermomixer (10-minute extraction). Glutelins were extracted at a temperature of 60°C in the thermomixer. After each extraction, the mixture was centrifuged at 11000 x g. The collected fractions were lyophilized and then dissolved in 2 cm³ of the respective phase (1-3), cleaned through a Spartan – 3NY filter with a 0.45 µm mesh and transferred to glass vials. The determinations were made using a Hewlett Packard Series 1050 system with the following parameters: column RP-18 Vydac 218TPP54, 5 µm, 250 x 4.6 mm, pre-column Zorbax...
300SB-C18 4.6x12.5 mm, column temperature 45°C, mobile phase flow rate 1ml × min, injection size 20 μl. The separation was performed using a two-component gradient. The proportion of component A: 0 min 75%, 5 min 65%, 10 min 50%, 17 min 25%, 18 min 15%, 19 min 75%. The first gradient (A) was water with an addition of 0.1% TFA, while the second gradient (B) was ACN with an addition of 0.1% TFA. The detection was carried out using a detector manufactured by the same company, and the reading was done at a wavelength of 210 nm.

The results were analysed using HPLC 3D Chem Station software (Hewlett Packard).

The assays of protein fractions were carried out at the Department of Processing and Chemistry of Plant Raw Materials, Faculty of Food Sciences, University of Warmia and Mazury in Olsztyn.

Statistical calculations were performed using the STATISTICA software package (data analysis software system, version 6, StatSoft, Inc. 2003) based on the analysis of variance. Differences between means were determined at a significance level of p = 0.01. The mean values for grain weight per plant, biometric features, rates of photosynthesis and transpiration, intercellular-space CO₂ concentration, and stomatal conductance were classified into homogeneous groups using Fisher’s test of significance.

**RESULTS AND DISCUSSION**

The present study showed that a reduction in soil moisture content from 60 – 70% SWC down to 30 – 35% SWC starting from the stage with 50% of panicle emerged up to the milk stage of grain development resulted in a decrease in grain weight per plant (Table 1). Podolska as well as Holubowicz-Kliza (2006A and 2006B) also demonstrated that cereal plants were characterized by the greatest sensitivity to water deficit during the heading, flowering and grain development stages.

### Table 1

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Soil water capacity (%)</th>
<th>Plant height (cm)</th>
<th>Number of panicles per plant</th>
<th>Number of grains per panicle</th>
<th>Thousand grain weight (g)</th>
<th>Grain weight per plant (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Flâmingsstern</td>
<td>60 – 70 %</td>
<td>57.05 a</td>
<td>5.21 b</td>
<td>24.31a</td>
<td>23.98 a</td>
<td>2.57 b</td>
</tr>
<tr>
<td></td>
<td>30 – 35 %</td>
<td>58.68 a</td>
<td>2.70 a</td>
<td>21.96 a</td>
<td>20.45 a</td>
<td>1.09 a</td>
</tr>
</tbody>
</table>

Homogeneous groups a, ab, b, according Fisher’s LSD test

### Table 2

Gas exchange rates for oats under different soil moisture conditions in 2004

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Soil water capacity (%)</th>
<th>Photosynthesis (μmolCO₂ m⁻² s⁻¹)</th>
<th>Transpiration (mmolH₂O m⁻² s⁻¹)</th>
<th>Intercellular-space CO₂ concentration (μmolCO₂ mol⁻¹)</th>
<th>Stomatal conductance (molH₂O m⁻² s⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>I II III</td>
<td>I II III</td>
<td>I II III</td>
<td>I II III</td>
<td>I II III</td>
</tr>
<tr>
<td>Flâmingsstern</td>
<td>60 – 70%</td>
<td>4.5b 3.5b 5.2b</td>
<td>1.2a 0.9a 1.6a</td>
<td>321b 312b 327ab</td>
<td>0.52b 0.40b 0.57b</td>
</tr>
<tr>
<td></td>
<td>30-35%</td>
<td>4.4a 5.0a 4.2a</td>
<td>1.2a 0.9a 1.6a</td>
<td>266a 239a 264a</td>
<td>0.07a 0.06a 0.10a</td>
</tr>
</tbody>
</table>

I – Measurement of gas exchange rates at panicle emergence; II – Measurement of gas exchange rates at full flowering; III – Measurement of gas exchange rates at grain formation Homogeneous groups a, ab, b, according Fisher’s LSD test
Table 3
Gas exchange rates for oats under different soil moisture conditions in 2005

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Soil water capacity (%)</th>
<th>Photosynthesis (μmolCO₂m⁻²s⁻¹)</th>
<th>Transpiration (mmolH₂Om⁻²s⁻¹)</th>
<th>Intercellular-space CO₂ concentration (μmolCO₂mol⁻¹)</th>
<th>Stomatal conductance (molH₂Omm⁻²s⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>I  II  III</td>
<td>I  II  III</td>
<td>I  II  III</td>
<td>I  II  III</td>
<td>I  II  III</td>
</tr>
<tr>
<td>Flämingsstern</td>
<td>60 – 70%</td>
<td>16.9a 15.2b 15.0b</td>
<td>3.9b 3.6b 3.2b</td>
<td>295b 177ab 141a</td>
<td>0.33b 0.05a 0.22b</td>
</tr>
<tr>
<td></td>
<td>30 – 35%</td>
<td>12.7a 6.2a 7.2a</td>
<td>2.6a 1.5a 0.8a</td>
<td>223a 130a 114a</td>
<td>0.09a 0.02a 0.03a</td>
</tr>
</tbody>
</table>

I – Measurement of gas exchange rates at panicle emergence; II – Measurement of gas exchange rates at full flowering; III – Measurement of gas exchange rates at grain formation. Homogeneous groups a, ab, b, according Fisher’s LSD test.

Table 4
Number of fungal isolates in the grain of oats cv. Flämingsstern under different soil moisture conditions in 2004

<table>
<thead>
<tr>
<th>Fungal species</th>
<th>Control 60 – 70% soil water capacity</th>
<th>Water stress 30 – 35% soil water capacity</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Acremoniella atra (Corda) Sacc.</td>
<td>–</td>
<td>4</td>
</tr>
<tr>
<td>2. Alternaria alternata Keissler Nees</td>
<td>3</td>
<td>6</td>
</tr>
<tr>
<td>3. Cladosporium cladosporoides (Fr.) de Wries</td>
<td>36</td>
<td>25</td>
</tr>
<tr>
<td>4. Fusarium poae (Peck) Wollenw.</td>
<td>15</td>
<td>6</td>
</tr>
<tr>
<td>5. Penicillium spp.</td>
<td>12</td>
<td>3</td>
</tr>
<tr>
<td>6. Sclerotinia sclerotiorum (Lib.) de Bary</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>7. Stemphylium botryosum Wallroth</td>
<td>–</td>
<td>1</td>
</tr>
<tr>
<td>8. Yeast-like mycelia</td>
<td>–</td>
<td>4</td>
</tr>
<tr>
<td>Total</td>
<td>67</td>
<td>49</td>
</tr>
</tbody>
</table>

SWC – soil water capacity; 60 – 70% SWC – control; 30 – 35% SWC – water stress

Table 5
Number of fungal isolates in the grain of oats cv. Flämingsstern under different soil moisture conditions in 2005

<table>
<thead>
<tr>
<th>Fungal species</th>
<th>Control 60 – 70% soil water capacity</th>
<th>Water stress 30 – 35% soil water capacity</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Alternaria alternata Keissler Nees</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>2. Cladosporium cladosporoides (Fr.) de Wries</td>
<td>8</td>
<td>4</td>
</tr>
<tr>
<td>4. Fusarium poae (Peck) Wollenw.</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>5. Mucor spp.</td>
<td>–</td>
<td>1</td>
</tr>
<tr>
<td>5. Papularia sphaerosphera</td>
<td>–</td>
<td>1</td>
</tr>
<tr>
<td>6. Penicillium spp.</td>
<td>1</td>
<td>4</td>
</tr>
<tr>
<td>Total</td>
<td>11</td>
<td>15</td>
</tr>
</tbody>
</table>

SWC – soil water capacity; 60 – 70% SWC – control; 30 – 35% SWC – water stress
Measurements of the biometric traits of the water-stressed oats cv. Flämingsstern, conducted during this study, also showed that water stress had a significant impact on the reduction in the number of panicles per plant (Table 1). A declining trend was found also for TGW and number of grains per panicle, although these differences were not statistically proved (Table 1). The study of Michalski et al. (1999) on the effect of rainfall amounts and distribution on oat yields, carried out under field conditions during the period from April to June, showed that this species produced the highest yield when a cool May was followed by a wet June with rainfall levels of 80-100 mm. Rudnicki (1995) demonstrated a beneficial influence of the following rainfall and thermal conditions on the productivity of oats: “a warm and wet April, not very cool May with moderate rainfall levels, a warm June with above-average rainfall, and an averagely warm or warm July with high amounts of rainfall”. In the case of oats, average rainfall total close to the long-term average (190 mm from April to July) has the most beneficial effect on their productivity when rainfall distribution in particular months is as follows: 10% in April, about 21% in May, 19% in June, and 50% in July. Similar results obtained by Rudnicki (1995) show a clear increase in oat yields with increasing rainfall. The study of Koziala (2004) proved that there was a significant increase in yield under the influence of sprinkler irrigation, but this increase varied, ranging from 9.5% to 79.6%. The number of productive panicles and 1000 grain weight also increased. However, the aforementioned author did not demonstrate the impact of sprinkler irrigation on the number of grains per panicle. In turn, Michalski et al. (2003) found that a drought during the growing period (in the years 2000 and 2001) had an adverse effect on thousand grain weight.

Water stress resulted in reduced rates of the investigated gas exchange parameters in oats both in the first and second year of the study (Table 2 and 3). Significant differences were found in the rates of photosynthesis and transpiration at the plant growth stages in question. Olszewski et al. (2007, 2009A) as well as Pszczółkowska et al. (2003) also showed that water deficit in the soil resulted in a reduction of the gas exchange rates, primarily the rate of photosynthesis. The results of the present study showed that the values obtained for the rates of photosynthesis, stomatal conductance and transpiration under conditions of optimal soil moisture content did not significantly differ from those obtained for oats in the study of Piotrowska et al. (2003).

In the first year of the study, it was demonstrated that oat kernels from the control treatment were colonized by fungi to a greater degree than kernels that had grown under water stress conditions (Table 4). In 2004 Cladosporium cladosporioides was the dominant species in both treatment combinations. Under the control conditions, 15 isolates of F. poae were identified, whereas under water stress conditions 6 isolates belonging to this species (Table 4). A reverse correlation was found in the second year of the study (2005), since more fungal isolates were obtained from water-stressed kernels (Table 5). Cladosporium cladosporioides was also the most frequently isolated species. Among the investigated fungi of the genus Fusarium, similarly as in 2004, the presence of F. poae was only found – 1 isolate in the control treatment and 3 isolates under water deficit conditions (Table 5). However, it should be stated that in 2005 the number of fungal cultures isolated from the oat grain was relatively low, and Fusarium poae was represented by single isolates.

Cladosporium cladosporioides was the dominant species in both years of the study. But the study conducted by Kowalczyk and Maciorowski (2006) showed that the following fungi occurred most frequently on the grain of naked oats grown under field conditions: Alternaria alternata, Epicoccum purpurascens, Fusarium poae, Penicillium spp., and non-sporulating colonies. Burgiel and Pisulewska (2003) also confirmed that Alternaria alternata, Epicoccum purpurascens, Penicillium spp., and Fusarium culmorum were the dominant species. However, Michalski and Horoszkiewicz-Janka (2003) indicated Alternaria tenuis as the dominant species both on the husked and naked form.

It was found in the present study that the toxin-producing fungi of the genus Fusarium were represented only by one species, Fusarium poae. The research

<table>
<thead>
<tr>
<th>Soil water capacity (%)</th>
<th>albums + globulins</th>
<th>prolamins</th>
<th>glutelins</th>
</tr>
</thead>
<tbody>
<tr>
<td>control (60-70% SWC)</td>
<td>43149</td>
<td>22831</td>
<td>25664</td>
</tr>
<tr>
<td>water stress (30-35% SWC)</td>
<td>44416</td>
<td>23659</td>
<td>22361</td>
</tr>
</tbody>
</table>

Table 6
Protein fraction content in the grain of oats cv. Flämingsstern under different soil moisture conditions
(the peak area calculated as mAU × s)
on fungal colonization of grain (from field crops) of 23 breeding oat stocks, conducted by Kowalczyk and Maciorowski (2006), also showed the dominance of Fusarium poae. On the other hand, Burgiel and Pisulewska (2003) indicated the occurrence of Fusarium culmorum in great numbers. In the opinion of Mielniczuk (2001) and Kiecanan et al. (2005), Fusarium avenaceum and Fusarium poae belong to the fungi of the genus Fusarium which occur most frequently on oat grain. Kiecanan et al. (2005) also showed that Fusarium poae played a major role in inducing fusariosis of oat panicles. Packa (2005) demonstrated that the percentage of Fusarium fungi in the grain of naked oats was 16.2% in the control treatment (non-inoculated), and the following were found in greatest numbers: Fusarium avenaceum, Fusarium culmorum, and Fusarium poae. In addition to the above-mentioned species, this author also found the presence of Fusarium poae in the oat grain in the treatments in which panicles had been inoculated with the species Fusarium culmorum and Fusarium avenaceum.

Qualitative analysis of proteins in the oat grain showed an increase in albumin, globulin and prolamolin fractions, whereas in the case of glutelin fractions a reverse correlation was found (Table 6). According to Olszewski et al. (2009B), during a drought period, both throughout the entire growing season and in the period from flowering to full maturity, the grain of oats cv. Flämingsstern contained more prolamins to Olszewski et al. (2009B), during a drought period, both throughout the entire growing season and in the period from flowering to full maturity, the grain of oats cv. Flämingsstern contained more prolamins than the grain of oats cv. Flämingsstern. According to Kowalczyk and Maciorowski (2006), during a drought period, both throughout the entire growing season and in the period from flowering to full maturity, the grain of oats cv. Flämingsstern contained more prolamins than the grain from the control conditions. The study of Konopka et al. (2007) also showed changes in protein fractions in the grain of water-stressed cereals.

**CONCLUSIONS**

1. Water deficit in the period from panicle half emerged to the milk stage of grain development resulted in a decrease in grain weight per plant.
2. Under conditions of reduced soil moisture content, a decrease in the gas exchange rates was found, mainly in the rates of photosynthesis and transpiration.
3. Cladososporium cladosporioides was the dominant species on kernels in both experimental treatment options and in both years of the study; moreover, the presence of a toxin-producing species, Fusarium poae, was found in the oat grain.
4. Under soil water deficit conditions, the oat grain was characterized by higher contents of prolamin, albumins, and globulins.

This study was supported by the Ministry of Education and Science in Poland, grant no. PBZ-KBN-09/P06/2003.

**LITERATURE**


Gilman J. C., 1957. A manual of soil fungi. The Iowa State University, Ames USA.


**Produktywność i zdrowotność ziarnia owsa oplewionego (Avena sativa L.)**

**w warunkach zróżnicowanego uwilgotnienia gleby**

**Streszczenie**

Badano wpływ zróżnicowanego uwilgotnienia gleby (60 – 70% ppw – kontrola; 30 – 35% ppw stres wodny) na plonowanie, wskaźniki wymiany wodnej, zdrowotność ziarn oraz frakcję białek ziarnia owsa oplewionego. W badaniach wykazano, że niedobór wody przyczynił się do obniżenia masy ziaren z rośliiny, zmniejszenia intensywności parametrów wymiany gazowej, a głównie fotosyntezy i transpiracji. W zirniakach owsa w obu wariantach doświadczalnych i latach badań gatunkiem dominującym było Cladosporium cladosporioides. Stwierdzono również obecność Fusarium poae. W ziarnie owsa pochodzące z różliń uprawianych w warunkach niedoboru wody w podłożu stwierdzono większą zawartość frakcji prolamin oraz albumin i globulin.