Diversity of the floral nectaries surface of four *Crataegus* L. species

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Summary

The performed studies focused on the surface structure of floral nectaries of four species from the following genus: *C. coccinea* L., *C. crus-galli* L., *C. curvisepala* Lindm and *C. prunifolia* (Poiret) Pers. The observations of the epidermis area were made in a scanning electron microscope (SEM). A nectary appears to be shaped like a slightly curved disk situated between the pistil style and the basal part of the stamens filaments. The nectary area of the studied species differed substantially as regards the cuticle sculpture and stomata number. The nectary secretion in *Crataegus* flowers proceeds through the stomata located below a level of the other epidermis cells, in the deep indents of the secretory tissue. The highest stomata number in 1 mm² nectary epidermis was recorded in *C. crus-galli, C. coccinea, C. prunifolia* and finally, *C. curvisepala*. Analyzing the nectary cuticular structure in respect of its increasing complexity (absence or presence of stripes), the investigated taxons can be ordered as following: *Crataegus curvisepala, C. coccinea, C. crus-galli* and *C. prunifolia*.

Key words: floral nectaries, epidermis, nectary stomata, cuticle, SEM, *Crataegus* L., Rosaceae

INTRODUCTION

Species from the *Crataegus* L. genus (f. Rosaceae, subf. Pomoideae) are found most frequently in North America. The following species, among others, are often cultivated in Poland: *C. coccinea* L., *C. curvisepala* Lindm., *C. crus-galli* L. and *C. prunifolia* (Poiret) Pers (Seneta and Dolatowski, 1997). Many of them are used in urban plantings on account of their ornamental flowers, fruit and leaves, as well as their medicinal properties (Rutkowski, 1998; Asgary et al., 1998; Wang, 1998). Trees
from the *Crataegus* genus are characterised by abundant nectar secretion and are included among valuable melliferous plants (Lipiński, 1976; Seneta and Dolutowski, 1997). Among honey types, hawthorn honey is also distinguished (Robert, 1994). Previously conducted studies show that within the *Crataegus* genus there are large differences in the amount of nectar secreted by flowers and the concentration of sugar in the nectar (Jabłoński and Kołtowski, 1996; Weryszko-Chmielewska et al., 1996). The sugar yield of several previously studied hawthorn species ranged between 0.33 and 4.22 mg of sugars from 10 flowers. The highest weight of sugars was found in *C. crus-galli*, while the lowest in *C. coccinea* (Weryszko-Chmielewska et al., 1996, 2003b). Due to the content of particular types of sugars, the nectar of hawthorns was included in the “Pyrus - type”, in which little sucrose occurs and the amount of glucose significantly exceeds the content of fructose (Maurizio and Grafl, 1969). Hitherto, it has been observed that cells of the nectary epidermis demonstrate large differences in the surface, also between the genera within the Pomioideae subfamily (Weryszko-Chmielewska and Konarska, 1995, 1996; Toth et al., 2000; Weryszko-Chmielewska et al., 2003a, 2004). The structure of the epidermis of the nectary gland is one of the factors affecting the amount of nectar secreted. The structural features of nectaries may also be of taxonomic value.

The aim of this study was to compare the surface structure of the secretory tissue of nectaries in four species from the *Crataegus* genus, with special attention to the cuticular ornamentation and the position of secretory stomata. The number of stomata found in mm² of the nectary epidermis was also determined.

**MATERIALS AND METHOD**

The studies conducted related to the structure of floral nectaries of *Crataegus coccinea* L., *C. crus-galli* L., *C. curvisepala* Lindm. and *C. prunifolia* (Poiret) Pers. Trees from which flowers were sampled grew in the Botanical Garden of the Maria Curie-Skłodowska University in Lublin.

For the scanning electron microscopy (SEM) examination of the surface of the nectary epidermis, fragments of flowers were fixed in 4 % glutaraldehyde 4 and (0.1 M) phosphate buffer with pH 7.0 for 4 hours at room temperature. After dehydrating the specimens in ethanol series and acetone, they were dried at critical point with liquid CO₂. Then the plant material was coated with gold by using the CS 100 Sputter Coater 100. Observations of the surface of the nectary epidermis in the aforementioned taxa were carried out by using the BS-300 Tesla microscope.

In the longitudinal sections, the thickness of the secretory tissue was compared based on semi-permanent slides from fresh plant material which were obtained by using the Vibratome 2000 (Technical Product International INC, USA). The 60 µm thick sections were embedded in glycerol-gelatine.

In order to prepare semi-thin sections, the plant material was postfixed with 1% osmium tetroxide, then dehydrated and embedded in Spurr’s epoxy resin. Sections of 0.75 µm thickness were cut by using the Reichert Ultracut S ultramicrotome, then stained with 1% methylene blue with 1% azur II. The sections were mounted in synthetic resin Eukitt.
RESULTS

The flowering of the aforementioned species occurs in May and June. The length of this period in the climatic conditions of Lublin was about 14 days. Flowers of the taxa studied were characterised by a white corolla and bright red anthers, as well as a diverse structure of the pistil. *C. coccinea*, *C. prunifolia* and *C. crus-galli* flowers develop a distylous pistil, whereas in *C. curvisepala* a monostylous pistil occurs (Figs 1-4).

Figs 1 4. Fragments of the flowers of three *Crataegus* species: 1, 4 *C. prunifolia*, 2 *C. curvisepala*, 3 *C. coccinea*; visible nectary surface (N) and dry nectar (arrow). Bars 2 mm (figs 1, 2, 3, 4).
Around the style of the pistil in the four species studied, numerous, strongly elongated unicellular hairs were observed in the epidermis of the nectary tissue (Figs 2, 4-6).

Nectar in the analysed flowers of the *Crataegus* representatives is easily accessible for insects, since stamens and elements of the perianth, when opening during flowering, uncover the nectary gland, situated on the adaxial surface of the concave flower receptacle. The nectary was yellow coloured, and it covered the surface between the style of the pistil and the basal part of filaments (Figs 1-6). The secretory tissue

Figs 5 6. Longitudinal sections of lower part of *Crataegus coccinea* flower with nectary (N), visible numerous trichomes (arrow) at the style base. Bars 150 µm (fig. 6), 200 µm (fig. 5).

Fig. 7. Longitudinal section of nectary tissue (N) *C. crus galli*. Bar 200 µm.
Figs 8 12. Fragments of the nectary epidermis surface of three *Crataegus* species: 8 *C. crus gali*; 9, 10 *C. curvisepala*; 11, 12 *C. prunifolia*; visible cells with smooth and striated (arrow) cuticle, stomata (arrow head) in depressions of secretion tissues and wax coating (double arrow). Bars 5 μm (fig. 8), 10 μm (figs 10,12); 50 μm (figs 9, 11).
Figs 13–17. Fragments of nectary epidermis surface of *Crataegus coccinea*, 13–15 the cells of epidermis from upper part of nectary with striped cuticle (arrows), 16–17 the cells from the lower part of nectary with smooth and corrugated cuticle at stomata (arrow head) laid in depressions. Bars: 5 μm (figs 14, 17); 20 μm (figs 13, 15, 16).
was characterised by a several-layers of cells, and it made up half of the thickness of the flower receptacle in this region (Figs 5-7).

Observed from above, the contour of the epidermis cells covering the nectary tissue in flowers of the taxa studied had a quadrangular shape (Figs 8-12, 16, 17). Nectar is secreted through stomata located below the level of the other cells of the nectary epidermis (Figs 8-12, 16, 17). As a result of a large depression, it was difficult to determine the aperture of the stomata, it was only found in *C. prunifolia* that they were open (Fig. 12). The modified stomata occurring in the epidermis of the nectaries of the observed species from the *Crataegus* genus were classified as anomocytic. They are characterised by the occurrence of a variable number of epidermis cells which surround the guard cells in a ring, and they do not differ from other cells of the epidermis (Figs 9-12, 16). The studied species of hawthorn developed a large number (175-308) of stomata in mm² of the nectary epidermis. The largest number of stomata was recorded in the epidermis of *C. crus-galli*, a much smaller number in *C. coccinea*, *C. prunifolia*, and *C. curvisepala* (tab. 1). The stomata were evenly distributed over the whole surface of the nectary (Figs 9, 11, 16).

<table>
<thead>
<tr>
<th>Species</th>
<th>Number of stomata in mm² epidermis</th>
<th>Cuticle</th>
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</thead>
<tbody>
<tr>
<td><em>C. coccinea</em></td>
<td>216</td>
<td>faintly striped</td>
</tr>
<tr>
<td><em>C. crus galli</em></td>
<td>308</td>
<td>faintly striped</td>
</tr>
<tr>
<td><em>C. curvisepala</em></td>
<td>175</td>
<td>smooth</td>
</tr>
<tr>
<td><em>C. prunifolia</em></td>
<td>185</td>
<td>greatly striped</td>
</tr>
</tbody>
</table>

The observations in scanning electron microscopy show that the outer wall of the nectary epidermis was covered by a layer of the cuticle differing in the ornamentation in particular taxa. The cuticle on the surface of the epidermis cells of the *C. curvisepala* nectary was smooth with wax coating which formed in some places an irregular, reticulate structure (Figs 9, 10). Some cells of the epidermis in *C. coccinea* and *C. crus-galli* showed clear striation of the cuticle, and other cells had a smooth surface (Figs 8, 13 17). On the surface of the *C. crus-galli* nectary, cuticular striae were observed in some places, undulate and interleaved, forming an ornamentation partly concealing the contours of the epidermis cells (Figs 8). But cells of the epidermis covering the *C. prunifolia* nectary tissue had clear cuticular sculpture in the form of longitudinal, slightly undulate striae running parallel within particular cells and radially around the stomata (Figs 11, 12). When analysing the cuticular sculpture of the nectary epidermis in terms of the increasing degree of complexity of structure, the taxa studied can be ranked as follows: *Crataegus curvisepala*, *C. coccinea*, *C. crus-galli* and *C. prunifolia*. 
DISCUSSION

The nectaries in flowers of the taxa studied from the *Crataegus* genus form on the surface of the flower receptacle, a glandular layer which, according to Smets (1986), developed from the reduced stamen whorl in the *Rosidae* subclass. Frey-Wyssling (1955) found that the disc nectary belongs to highly specialised structures occurring within the abovementioned plant subclass. In terms of the form in the Pomoideae, the following types of nectaries were distinguished: automorphic (*Crataegus, Pyrus, Chaenomeles*), epimorphic (*Cydonia*) and transitoric (*Malus*) (Weryszko-Chmielewska and Konarska, 1995; 1996).

Literature data show that nectaries of plants belonging to the *Rosidae* can be divided in terms of the presence or absence of modified stomata (Smets, 1988). Another division, relating to the tissue origin of nectaries, defines them as mesophilic (Vogel, 1977; Schmid, 1988).

In flowers of the studied taxa from the *Crataegus* genus the presence of the secretory tissue was found, secreting nectar through anomocytic stomata. The state of opening of the stomata in *C. curvisepala, C. crus-galli* and *C. coccinea* was difficult to determine due to their deep location. In the longitudinal sections of flowers, it was observed that the stomata in *C. crus-galli* and *C. coccinea* reach the second level of the glandular cells, which were characterised by a thick wall, numerous plastids with starch grains and a high degree of vacuolisation (Weryszko-Chmielewska et al., 2003b).

Open stomata in the epidermis of the nectaries analysed in the present study of hawthorns were only visible in *Crataegus prunifolia*. However, earlier studies of Weryszko-Chmielewska et al. (2003b) showed the presence of closed stomata in the secretory tissue of *Crataegus crus-galli*, in spite of the fact that observations were carried out during the secretion period, while in *Malus* (Toth et al., 2000) the stomata were characterised by different apertures, what indicates their ability to open and close. These observations suggest the active functioning of stomata in the epidermis of nectaries of the abovementioned taxa, what is related to the rhythmic secretion of nectar (Toth et al., 2000; Weryszko-Chmielewska et al., 2003b).

Among the analysed species from the *Crataegus* genus, the largest number of stomata on the surface area of 1 mm² of the nectary epidermis was observed in *C. crus-galli*, and the lowest in *C. curvisepala*. In many plant species, a positive correlation between the number of stomata in the nectary epidermis and the abundance of nectar secretion was found (Teuber et al., 1980; Murrell et al. 1982; Davis and Gunning, 1991; Weryszko-Chmielewska et al., 1997). One can expect that a similar dependence applies to nectaries of all the *Crataegus* species studied, since it was previously demonstrated for *C. crus-galli* and *C. coccinea* (Weryszko-Chmielewska et al., 2003b).

*Crataegus prunifolia* is a species developed as a result of crossing of *C. crus-galli* and *C. macracantha* (Rutkowski, 1998). In spite of the very close affinity
of *C. crus-galli* and *C. prunifolia*, significant differences were demonstrated in the structure of the surface of the nectar epidermis in these species. The cuticle was characterised by a different ornamentation, from partially striate in *C. crus-galli* to strongly striate over the whole surface in *C. prunifolia*. In the four *Crataegus* species studied, a total of 3 types of the nectar surface, related to the cuticular sculpture, were distinguished: smooth, striate in some places and striate over the whole surface. The striate sculpture of the cuticle of the nectary probably performs different functions, among others, protects nectar against drying and facilitates the distribution of the secretion over the surface of the nectary. According to Hejnowicz (2002), the rich cuticular sculpture more effectively “repels” from the epidermis rainwater drops which, when falling, do not flatten, and rolling over the surface, they enable it to self-clear. In its composition, the cuticle contains special carbohydrates, and on its surface waxes deflecting excessive UV radiation. It is particularly important, especially for the glandular tissue unprotected by stamens and the perianth. This layer also helps to limit the penetration of different chemical substances affecting plants.

At the same time, the cuticle, which is a complex compound made up of strongly polymerised fatty acids resistant to the action of water, is characterised by chemical stability (Esau, 1973; Hejnowicz, 2002), thanks to which it does not decompose. Esau (1973) states that no microorganism decomposes the cuticle, since it does not have any enzymes which degrade the cutin. As a result of this, the cuticle of the nectar epidermis is an important mechanical barrier against unfavourable conditions of the environment, among others: infection, damage by the wind and insects.

Significant differences, observed in this study, in the micromorphology of the nectaries of closely related species within the *Crataegus* genus may result from adaptations to different habitat conditions in which they occur in natural locations. According to Heywood (1974), the varied cuticular ornamentation may be used as a taxonomic value.

**CONCLUSIONS**

1. In flowers of the studied species from the *Crataegus* genus, nectar is secreted through anomocytic stomata located in deep depressions of the secretory tissue.

2. The largest amount of stomata for one unit of surface area was found in the epidermis of the *C. crus-galli* nectaries, then in *C. coccinea*, *C. prunifolia* and *C. curvisepala*.

3. The epidermis covering the floral nectaries in the studied species from the *Crataegus* genus formed three types of cuticular ornamentation: smooth, partially striate and strongly striate.

4. In terms of the increasing degree of complexity of cuticular sculpture, the *Crataegus* nectaries can be ranked as follows: *Crataegus curvisepala*, *C. coccinea*, *C. crus-galli* and *C. prunifolia*.
REFERENCES


Zróżnicowanie powierzchni nektarników kwiatowych czterech gatunków z rodzaju Crataegus L.

Streszczenie
